



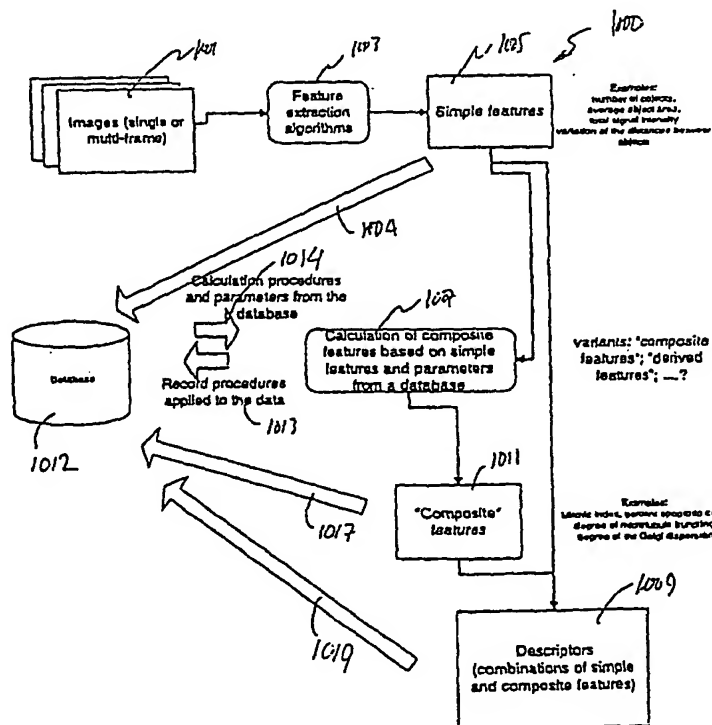
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(57) Abstract

Techniques for using information technology in therapeutics or drug discovery. In an exemplary embodiment, techniques for determining information about the properties of substances based upon information about structure of living or non-living cells exposed to substances are provided. A method according to the present invention enables researchers and/or scientists to identify promising candidates in the search for new and better medicines or treatments using, for example, a cellular informatics database. The present invention further teaches a system for acquiring knowledge from cellular information. The system has a database 1012 comprising a database management module ("DBMS"). The system also has a variety of modules, including a population module coupled to the DBMS for categorizing and storing a plurality of features (e.g., cell size, distance between cells, cell population, cell type) from an image acquisition device into the database. The system has a translation module coupled to the DBMS for defining a descriptor from a set of selected features from the plurality of features. In a specific embodiment, the descriptor is for a known or unknown compound, e.g., drug. A prediction module is coupled to the DBMS for selecting one of a plurality of descriptors from known and unknown compounds from the database based upon a selected descriptor from a selected compound. The selected compound may be one that is useful for treatment of human beings or the like.



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PATENT APPLICATION
METHOD AND APPARATUS FOR
PREDICTIVE CELLULAR BIOINFORMATICS

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10 computer codes, which may be used to implement aspects of the present invention. Assignee of the present invention reserves all rights with respect to these codes and provides notice herein. Notice is hereby given © Cytokinetics, Inc. 1999.

BACKGROUND OF THE INVENTION

 The present invention provides techniques for information
15 management using a database platform. More particularly, the present invention provides a system including computer code that couples to a database device. The system provides for image capturing of living, dead, or fixed cells or cell fractions used to identify information about substances used on the cells or information about the cells themselves. Accordingly, the present invention can enable researchers and
20 scientists to identify promising candidates in the search for new and better medicines, for example, in drug discovery and development. The principles enumerated herein may, with equal facility, be applied to other applications, including but not limited to use in environmental applications such as determining chemical toxicities and other non-pharmaceutical toxicology uses.

25 For a long time, researchers in the pharmaceutical field have sought for better ways of searching for substances possessing properties that make them suitable as medicines. In the early days, researchers generally relied upon extracts from plants, dyes, and microbiological extracts for such substances. Examples of such substances include the pain reliever aspirin, the anti-cancer drug paclitaxel (brand
30 name TaxolTM), and the heart medication called digoxin. The number of useful medicines has generally been limited.

Purified substances having desirable bio-active properties are also often difficult to discover. Advances in traditional organic chemistry and more recently the rapid chemical synthesis methods often referred to as combinatorial chemistry have increased the number of compounds that researchers test for biological activity. Originally, substances were often initially tested on animals or humans to determine their biological activity. While results from such tests may identify a good drug candidate, they are often time consuming and costly, thus a limited number of substances can be tested. Therefore, pharmaceutical companies have turned to testing their ever-increasing libraries of substances against isolated proteins (drug targets) in biochemical assays that can be carried out at high throughput and low cost. It should be noted that the substances need to be tested in numerous protein tests, each customized for a particular drug target. Therefore, although each protein test may be run at a high-throughput, the design of multiple protein tests can be time-consuming. Substances deemed promising based on results from the protein tests are then tested in lower throughput cellular and animal tests.

There have been some attempts to use image acquisition techniques to screen a large number of substances based upon biological cell information. One such attempt is described in International Application No. WO 98/38490 in the names of Dunlay, et al. Dunlay et al. generally describes a conventional image acquisition system. This conventional system collects and saves images based on certain criteria that are predefined, not on a fixed area of an imaging surface. Additionally, the conventional system has poor lighting design, which makes image processing for multiple cells difficult. Furthermore, the conventional system is not designed for capturing, populating and utilizing a large database design. The conventional system is designed for customized cellular assays, not as a tool for generation of a cellular informatics database. Without such database capabilities the conventional system cannot be used for screening, analyzing, and comparing large quantities of cells from multiple experiments on multiple days in a predictive, efficient and cost effective manner.

What is needed is a rapid assay to assess the activity of compounds against multiple drug targets simultaneously in a cellular context. What is also needed are techniques for finding the effects of substances on cell function based upon searching and analyzing cellular information.

SUMMARY OF THE INVENTION

According to at least one embodiment of the present invention, techniques for determining information about effects of potential substances on cells are provided. In another exemplary embodiment, the present invention provides a novel system including hardware, computer codes, user interfaces, and a database for acquiring, storing and retrieving cellular and substance information. The cells can include living, dead, or fixed cells or fractions of cells. The present invention enables, *inter alia*, researchers and/or scientists to identify promising candidates in the search for new and better medicines or treatments using, for example, a cellular informatics database.

According to the present invention, a computer program for identification and verification of biological properties of substances can include code that causes a sample of a substance to be administered to a cell. The code determines one or more features for two or more cell components, or markers, in the presence of the substance. The code can form one or more descriptors from the features. Descriptors can be formed by combining features of two or more cell components as identified using the markers. The code can then search one or more descriptors obtained from prior administered substances upon cells in order to locate descriptors having a relationship to the descriptors noted for the substance under study. The code predicts properties of the administered substance based upon the properties of the prior administered substances using the relationship between the descriptors. The code can provide for identifying properties of substances based upon effects on cell characteristics. Candidate drug mechanisms of action, potency, specificity, pharmacodynamic, and pharmacokinetic parameters, toxicity, and the like can be used as substance properties.

In a specific embodiment, the present invention provides a system for acquiring knowledge from cellular information. The system has a database comprising a database management module ("DBMS"). The system also has a variety of other modules, including a population module that is coupled to the DBMS and serves to categorize and store a plurality of features (including but not limited to cell size, distance between cells, cell population, as well as sub-cellular features such as organelle location, protein location and sub-cellular constituent location and

movement) from an image acquisition device into the database. The system has a translation module coupled to the DBMS for defining a descriptor from a set of selected features from the plurality of features. In a specific embodiment, the descriptor is for a known or unknown compound, e.g., drug. A prediction module is coupled to the DBMS for selecting one of a plurality of a descriptors from known and unknown compounds from the database based upon a selected descriptor from a selected compound. The selected compound may be one that is useful for treatment of human beings or the like.

In a specific embodiment, the present invention provides a system for populating a database with cellular information. The system includes a cell holder (e.g., multi-well plate, chip, microfluidic assembly, or other cell chamber) comprising a plurality of sites in a spatial orientation. Each of the sites is capable of holding a plurality of cells to be imaged. Note – the light guide is one embodiment, but we don't want to be limited to it.

According to one embodiment, the present system also has an illumination apparatus including a liquid light guide operably coupled to the imaging device for highlighting the plurality of cells in a relatively even spatial manner for image capturing and measurement purposes. Still further, the liquid light guide allows sub-elements (e.g., filter, lamp) of the illumination apparatus to be placed at a remote location to prevent mechanical interference of the cell holder during image capturing. Alternative lighting methodologies may, with equal facility, be implemented.

The system also has an image-capturing device (e.g., charge coupled device camera, translation stage, shutter, microscope, software, shutter control) coupled to a computing device (e.g., computer, network computer, work station, analog computing device, on-board image-processor, and laptop). The image-capturing device is adapted to capture at least one image in at least one of the plurality of sites. One some embodiments, multiple images can be captured, where each image represents a different cell component (or portion). The image-capturing device can be adapted to convert the image into a digital representation, which highlights the feature or features of the one site.

A database storage device (e.g., relational database, object oriented database, mixed object oriented database) includes a database management element. The

database is coupled to the image capturing device. In a specific embodiment, the present system includes modules for feature extraction, generation of descriptions, and data preparation and analysis.

In a specific embodiment, the present invention provides a novel
5 system for determining an effect of a manipulation of a cell using one or more image frames. The system has a plate comprising a plurality of sites in a spatial orientation. Each of the sites is capable of holding a plurality of cells to be imaged. The system also has an image capturing device to capture a plurality of images of at least one site from the plurality of sites. The image capturing device is coupled to the computing
10 device. The system also has an image processing device to combine the plurality of images of at least one site or plurality of sites. The image processing device is operably coupled to the plate. An image processing device is also included. The image processing device can be adapted to form a digitized representation of the plurality of images from the site or plurality of sites. Furthermore, the system has a
15 database storage device comprising a database management element. The database can be adapted to retrieve the descriptor or descriptors of the plurality of features from the computing processing device and storing them in a selected manner.

In a specific embodiment, the present invention provides a system for capturing cellular information. The system also has an image acquisition system
20 comprising a charged coupled device camera adapted to capture an image of a plurality of manipulated cells in various stages of the cell cycle. The stages of the cell cycle are currently understood to include interphase, G0 phase, G1 phase, S phase, G2 phase, M phase, prophase, prometaphase, metaphase, anaphase, and telophase. The principles of the present invention specifically contemplate the application thereof on
25 additional cell cycle stages when and if they are identified.

An optical source is coupled to the image acquisition system for highlighting the plurality of manipulated cells in the various stages of the cell cycle. The illumination apparatus provides for an acquisition of the image of the plurality of manipulated cells. In a specific embodiment, the illumination apparatus has a liquid
30 light guide coupled to a light source at a remote location.

A variety of user interfaces are utile for accessing the several features of the present invention. Those having ordinary skill in the art will appreciate that different user interfaces may be required to support different research scenarios. The

present invention specifically contemplates the utilization of a wide variety of user interfaces.

Numerous benefits are achieved by way of the present invention over conventional techniques. The present invention can provide techniques for predictive cellular bioinformatics that can streamline a number of important decisions made in the drug discovery industry. The present invention can be implemented using off the shelf hardware including databases. In other aspects, the present invention can find useful information about substances as well as cells or portions of cells. Furthermore, the present invention can acquire more than one feature using more than one manipulation. Moreover, the present invention can provide information about a wide variety of cellular information that is not conventionally available. This information includes information about different cell components, e.g., nuclei and Golgi apparatus. Still further, the present invention provides an automated or semi-automated technique for acquiring images and populating a database. The present database can be combined with others such as genomics, and the like. Moreover, the present invention can be implemented to predict, *inter alia*, a mechanism of action, toxicity, target validation, and pre-clinical disease model.

A further understanding of the nature and advantages of the invention herein may be realized by reference to the remaining sections of the specification and the attached drawings.

BRIEF DESCRIPTION OF THE DRAWING

For more complete understanding of the present invention, reference is
5 made to the accompanying Drawing in the following Detailed Description of the
Invention. In the drawing:

Fig. 1 is a simplified system diagram according to an embodiment
according to the present invention;

10 Figs. 1A-1B are more detailed diagrams of database systems according
to embodiments of the present invention;

Fig. 2 is a simplified block diagram according to an alternative
embodiment according to the present invention;

Figs. 3-6 are simplified diagrams of system elements according to
embodiments of the present invention;

15 Figs. 7A-7K illustrate representative block diagrams of simplified
process steps in a particular embodiment according to the present invention;

Fig. 8A-8F illustrate representative quantified descriptors of effects of
manipulations on images of cells in a particular experiment;

20 Fig. 9 illustrates example images for different types of morphologies in
a particular experiment;

Fig. 10 illustrates a distribution of various morphologies in a cell
population responsive to drug concentration in a particular experiment;

Fig. 11 illustrates a graph of quantified features of effects of
manipulations on cells in a particular experiment;

25 Fig. 12 illustrates effects of external agents on cells in a particular
experiment;

Fig. 13 illustrates 4 panels for each marker for a plurality of A549 cells
in a particular experiment;

30 Fig. 14 illustrates 4 panels for each marker for a plurality of OVCAR-3
cells in a particular experiment;

Fig. 15 illustrates 4 panels for each marker for a plurality of OVCAR-3
cells at 20x in a particular experiment;

Fig. 16 illustrates 4 panels for each marker for a plurality of OVCAR-3 cells at 40x in a particular experiment;

Fig. 17 illustrates a representative input for a morphometric analysis program in a particular embodiment according to the present invention; and

5 Figs. 18-19 illustrate examples of the generation of pseudo-sequences and clustering in a particular embodiment according to the present invention.

Fig. 20 is a block diagram for a first research scenario;

Fig. 21 is a block diagram for a second research scenario; and

Fig. 22 is a block diagram for a third research scenario.

10 Reference numbers refer to the same or equivalent parts of the invention throughout the several figures of the Drawing.

DETAILED DESCRIPTION OF THE INVENTION

According to the present invention, techniques for determining information about manipulated cells or substances based upon living, fixed, or dead cell structures or portions of cells are provided. In an exemplary embodiment, the present invention provides a novel system including computer codes coupled to a database and user interfaces for acquiring, storing and retrieving such information. Other embodiments provide a novel image capturing system for providing digitized representations of live and dead cell structures or the like.

Fig. 1 is a simplified system diagram 10 of a cellular knowledge-based system according to an embodiment of the present invention. This diagram is merely an example and should not limit the scope of the claims herein. One of ordinary skill in the art would recognize other variations, modifications, and alternatives. The present system 10 includes a variety of elements such as a computing device 13, which is coupled to an image processor 15 and is coupled to a database 21. The image processor receives information from an image capturing device 17, which image processor and image capturing device are collectively referred to as the imaging system herein. The image capturing device obtains information from a plate 19, which includes a plurality of sites for cells. These cells can be biological cells that are living, fixed, dead, cell fractions, cells in a tissue, and the like. The computing device retrieves the information, which has been digitized, from the image processing device and stores such information into the database. A user interface device 11, which can be a personal computer, a work station, a network computer, a personal digital assistant, or the like, is coupled to the computing device.

Fig. 1A is a simplified diagram of a database system 1000 according to an embodiment of the present invention. This diagram is merely an example and should not limit the scope of the claims herein. One of ordinary skill in the art would recognize many other variations, modifications, and alternatives. Database system 1000 includes a variety of techniques for processing images from biological cells, e.g., fixed, living, and dead cells, and cell portions. As shown, images are acquired 1001. These images can be from a single frame or multiple frames. As merely an example, an image processing system may analyze such images. One example of

such an image processing system is described below, but should not be construed as limiting certain claims.

In a specific embodiment, cell samples are manipulated using a compound (e.g., substance, drug). The cell samples are imaged for a simple portion or portions, e.g., manipulated cell substructure, manipulated spatial feature of cell, cell density. Image processing techniques are used to extract the feature or features from the image or images. The features can be an independent or a dependent set of cell characteristics (which may be predominately visual) including, for example, count, area, perimeter, length, breadth, fiber length, fiber breadth, shape factor, elliptical form factor, inner radius, outer radius, mean radius, equivalent radius, equivalent sphere volume, equivalent prolate volume, equivalent oblate volume, equivalent sphere surface, average intensity, total intensity, optical density, radial dispersion, texture difference, and others. Each of these features corresponds to a similar manipulation by a compound. Each manipulation forms a new set of features, which are identifiable to the compound. Once each set of features has been extracted, the feature set is populated into a database. Accordingly, the database includes many sets of features, where each set corresponds to a different manipulation for a selected cell. Each set of features corresponding to a manipulation provides a descriptor, which is also stored in the database. The descriptor is a "finger print" including each feature for the manipulation. Each descriptor may be unique, or may have similarities to other descriptors or may even be the same as other descriptors for known and unknown manipulations.

The present system retrieves features, which we define as simple features herein, and forms composite features from them. More than one feature can be combined in a variety of different ways to form these composite features. In particular, the composite feature can be any function or combination of a simple feature and other composite features. The function can be algebraic, logical, sinusoidal, logarithmic, linear, hyperbolic, statistical, and the like. Alternatively, more than one simple feature can be combined in a functional manner (e.g., arithmetic, algebraic). As merely an example, the composite feature equals a sum of feature 1 and feature 2, where these features correspond to the same manipulation. Alternatively, the composite feature equals feature 1 divided by feature 2. Alternatively, the composite feature equals feature 1 minus feature 2. Alternatively,

the composite feature equals a constant times feature 1 plus feature 2. Of course, there are many ways that the composite feature can be defined. The present system also stores 1017 these features in the database. The composite features can also be further combined with simple features. Once these features are defined as descriptors, they are stored 1019 in the database.

Fig. 1B is a simplified diagram of a database system engine 2000 according to an embodiment of the present invention. This diagram is merely an example and should not limit the scope of the claims herein. One of ordinary skill in the art would recognize many other variations, modifications, and alternatives. The engine can be implemented into the present database for populating, searching, and predicting compound or cell characteristics. As merely an example, engine 2001 includes an input/output module 2008. The input/output module is used to input and output information from the database. The information includes, among others, a plurality of feature sets, which correspond to many manipulations. Additionally, the information includes descriptors, which each corresponds to a set of features from the manipulation. The database also has a population module, which is used to configure the features based upon an entity relationship, which has been predetermined.

The database engine also has other modules. In particular, the database has a transcription module, which transfers a preselected set of features and creates a descriptor from them. The transcription module can be used to take a known compound, which has features, to transcribe them into a descriptor. Alternatively, the transcription module can be used to take an unknown compound, which has features, to transcribe them into a descriptor. These descriptors are provided into the database for subsequent use. Finally, the database engine has a prediction module, which can be used to potentially predict a property (e.g., mechanism of action) of an unknown compound. Here, the unknown compound is provided with a descriptor, but the property of the compound is unknown. In one embodiment, the prediction module compares a descriptor of an unknown compound with the many descriptors of known compounds, which were in the populated database. Depending upon the matching criteria, the prediction module will attempt to uncover one or more descriptors of known compounds. Once the prediction module finds the descriptors of the known compounds based upon the descriptor for the unknown compound, it identifies a potential property of such unknown compound for analysis and review. Here, it is

believed that certain features of the known compound, which are similar to those features of the unknown compound may uncover a property to the unknown compound. Details of the present software engine are described more fully below.

Fig. 2 is a simplified block diagram 20 of a cellular knowledge-based system according to an alternative embodiment of the present invention. This diagram is merely an example and should not limit the scope of the claims herein. One of ordinary skill in the art would recognize other variations, modifications, and alternatives. Like reference numerals are used in the present diagram as the previous diagram for easy cross-referencing, but are not intended to be limiting in any manner. The present diagram 20 includes a variety of elements such as a processor 13 or computing device coupled to a database 11. The processor can be used for retrieving and storing information from the database. The system also includes a plurality of system elements, such as a cleaner 23, a dispenser 25, and an image capturing system 27, which are also coupled to the database in some embodiments. These elements can be coupled to each other through a network or the like. As merely an example, the network can be a NetWareTM network from Novell Corporation or an internet network or the Internet but can also be others and any combination thereof. The system also has an output device 31, which can be used to output information from the database, processor, or other system elements. Details of these elements are described more fully below in reference to the Figs.

Figs. 3-5 are simplified drawings of system elements according to embodiments of the present invention. These diagrams are merely examples and should not limit the scope of the claims herein. One of ordinary skill in the art would recognize other variations, modifications, and alternatives. As merely an example, Fig. 3 is a simplified diagram of a processor or computing device 13. The computing device 13 includes a bus 112 which interconnects major subsystems such as a central processor 114, a system memory 116 (e.g., random access memory), an input/output ("I/O") controller 118, an external device such as a display screen 124 via a display adapter 126, a keyboard 132 and a mouse 146 via an I/O controller 118, a SCSI host adapter (not shown), and a floppy disk drive 136 operative to receive a floppy disk 138.

The computing device has other features. Storage Interface 134 may act as a storage interface to a fixed disk drive 144 or a CD-ROM player 140 operative

to receive a CD-ROM 142. Fixed disk 144 may be a part of computing device or may be separate and accessed through other interface systems. A network interface 148 may provide a direct connection to a remote server via a telephone link or to the Internet. Network interface 148 may also connect to a local area network ("LAN") or other network interconnecting many computer systems. Many other devices or subsystems (not shown) may be connected in a similar manner. Also, it is not necessary for all of the devices shown in Fig. 3 to be present to practice the present invention, as discussed below. The devices and subsystems may be interconnected in different ways from that shown in Fig. 3. The operation of a computer system such as that shown in Fig. 3 is readily known in the art and is not discussed in detail in this application. Computer code to implement the present invention, may be operably disposed or stored in computer-readable storage media such as system memory 116, fixed disk 144, CD-ROM 140, or floppy disk 138. The computer code can be organized in terms of processes or modules, depending upon the application. That is, the computer code can include a prediction module, a translation module, or other modules to carryout the functionality described herein, as well as others.

Figs. 4 and 5 are simplified diagrams of an imaging system 200 according to an embodiment of the present invention. As shown, the imaging system 200 includes a variety of features such as housing 203, which holds a stage assembly 204. The stage assembly includes an x-stage movement element 206, which is along an x-direction, and a y-stage movement element 207, which is along a y-direction. The imaging system also includes a z-direction movement element, which is perpendicular to the x-y plane. The z-direction movement motor can be attached to the stage, or to the objective nosepiece by way of the microscope housing, or as an external motor between the objective and the microscope housing. The stage can align in any one of the directions to an accuracy of one micron and less, or one-half micron and less, or one-quarter micron and less, depending upon the embodiment.

The stage holds a plate 202 or cell holder, which houses one of a plurality of samples. The plate includes a spatial array 209 of process sites. Each of the process sites can include a plurality of cells and solutions depending upon the embodiment. Each of the sites can carry a sufficient amount of solution to prevent substantial evaporation of the sample during processing in some embodiments. In embodiments for large scale analysis, the plate includes at least 96 sites, or more than

or equal to 384 sites, or more than or equal to 1,536 sites. The plate bottom is transparent and thin, which allows light to pass through the sample. Additionally, the plate is made of a suitable chemical resistant material. As merely an example, the plate can be either a 96, or 384, or 1536 or other formats from places such as Becton Dickinson of Franklin Lakes, NJ, or Corning Science Products of Corning, NY. In a preferred embodiment, the plate is a Corning Costar black-walled 96 well plate catalog #3904 from Corning Science Products of Corning, NY, but should not be limited to these in some applications, but can be others.

Also shown is the condenser for the microscope 201, which can be used to collect phase, DIC, or bright field images of the cells. Images resulting from the illumination of the samples to fluorescence, phase, DIC, or bright field techniques are collected using an image capturing device 208, which captures an image or images of cells from the plate. In a specific embodiment, the microscope is an inverted configuration with the objectives on the bottom of the plate and the condenser disposed overlying an upper surface of the sites, while the image capturing device underlies the sites. Images captured by the imaging device, whether analogue or digital, are viewed by a monitor or other devices. The image capturing device can be any camera assembly such as a charge coupled device camera, which is known as a CCD camera, or other high resolution camera capable of capturing images from the sites. In a specific embodiment, the camera is an interline CCD camera which does not require an external shutter.

In a specific embodiment, the present imaging system can be any suitable unit that is flexible for automated image collection using multi-well plastic plates. The imaging system also should be adapted to collect high-resolution images of cells on plastic or glass plates, cell growth chambers, or coverslips. The system also can be used for imaging multiple cell markers in multiple imaging conditions. To accomplish this, the microscope system has a variety of elements such as a light source, a motorized excitation filter wheel and shutter, x-y-z-motorized stage, excitation and emission filters, Fluor phase and DIC objectives, motorized objective nosepiece, dichroic filters, motorized dichroic filter cubes, phase and DIC rings and prisms, CCD camera, and software control. As merely an example, the present imaging system can have components such as those listed in the Table below.

DESCRIPTION	MAKER	MODEL
Microscope	Zeiss	100M
(x-y) motorized stage	Prior	
Xenon lamp	Sutter	Lambda
Filter wheel	Sutter	Lambda-10
Microtitre Plate holder	Prior	500-H223R
Isolation Table	Kinetic Systems	9101-24-85
Objective Spacers	Polytec PI	P-721.90
Camera	Hamamatsu	C47-95
Computer	IBM	IntelliStation
Software	Metamorph	v.4
Objectives	Zeiss	Achroplan 10x/0.25 LD-Achroplan 20x/0.4 LD-Achroplan 40x/0.6

Table: Image Acquisition System Elements

5 In a specific embodiment, the present system has the following capabilities, which are not intended to be limiting.

Image acquisition

1) Ability to automatically acquire multi-wavelength images from multiple sites on one multi-well plate, to sequentially name image files, and to log any
10 imaging parameter information with image files.

2) Ability to link images with a larger database/spreadsheet of information.

3) Ability to automatically collect multiple plates by interfacing the imaging system with a robotic arm.

15

X-Y control

1) Ability to place 96, 384, or 1536 well plates onto microscope stage and move to each well sequentially.

2) Ability to return to each well and collect another round of images (multi-site time-lapse) or ability to collect rapid time-lapse information at each well (time-lapse of many wells).

3) Ability to collect a low magnification image, automatically
5 determine features which may be of interest, automatically change the objective to a higher magnification, and collect high magnification images of a fixed number of those identified cells in the sample.

4) Ability to collect multiple frames in each site.

10 Z control

1. Ability to auto-focus with substantially minimal damage to biological specimen or fluorophore.

2. Ability to auto-focus rapidly.

15 The present embodiment of the imaging system is shown by way of Figs. 5A and 5B. These diagrams are merely examples and should not limit the scope of the claims herein. One of ordinary skill in the art would recognize other variations, modifications, and alternatives. The present imaging system 40 includes a variety of elements such as a microscope 41, which is preferably an epi-fluorescent microscope,
20 but can be confocal, multiphoton, or hybrid types. The microscope includes elements 41A, the motorized Z-axis; 41B, the motorized dichroic filter cube holder; and 41C, the motorized objective nosepiece. In one embodiment, the microscope is a Model 100M made by Zeiss. The microscope communicates to computer 51 through control lines 73, 75, and 76. The imaging system also has camera 50 coupled to controller
25 50A and computing device 51, which oversees and controls operations of the elements of the imaging system.

The present microscope includes drivers for spatially moving a stage in two dimensions, including an x-direction, a y-direction, and moving the objective nosepiece in a z-direction in a Cartesian coordinate system. The z-direction
30 movement is provided using a fast z-motor, which can make z-direction adjustments within a predetermined time. The z-direction movement generally provides for focussing of the sample to the camera. The focussing occurs within the predetermined time of preferably ten seconds and less, or five seconds and less, or one

second and less, depending upon the embodiment. As merely an example, the z-motor or positioner can be a model PIFOC objective nanopositioner made by a company called Physik Instrumente of Waldbronn, Germany, but also can be others. The z-motor couples to computer 51 through line 63, which may also include a controller. Depending upon the embodiment, a second z-motor 41A connected to the computer 51 by line 73 may be used to keep the z-motor 42 in the center of its travel. Alternatively, in other embodiments the stage could be provided with a z-motor allowing for movement of the stage in the z-direction.

The present stage also includes an x-y stage 43. The x-y stage moves plate 59, e.g., 96 site, 384 site, 1536 site. The x-y stage moves plate in an x-y spatial manner. The stage has an accuracy or repeatability of about 1 micron and less, or about 2 microns and less. The stage can move in a continuous manner or a stepped manner. The stage also can move up to 30 mm/sec. or faster. The stage also can move 1 mm/sec. and less, depending upon the embodiment. The stage can also step 0.1 micron and less or 1 micron and less, as well as other spatial dimensions. The stage can be one such as a Proscan Series made by Prior Scientific of Rockland, MA but can also be others. The stage is controlled via control line 61 through controller 43A, which couples to computer 51 through control line 65.

The stage includes plate holder 44. The plate holder can hold a single plate. In other embodiments, plate holder can also hold multiple plates. The plate holder can use mechanical, electrical, fluid, vacuum and other means for holding the plate or plates. The plate holder also is sufficiently stable for securing the plate. As merely an example, the plate holder is a Model 500-H223R made by Prior Scientific of Rockland, MA. In some embodiments, the plate holder may need adjustment in the z-direction to provide for a desirable focus of a sample on a plate. In these embodiments, the plate holder is supported by spacers 45 or a plurality of stage pins, which mechanically elevate the plate holder in the z-direction. These pins are generally made of a suitable material for supporting such plate holder and also are sufficiently resistant to chemicals and the like.

In some embodiments, the entire imaging system is placed on an isolation table 49. The isolation table is disposed between the microscope and support structure. The isolation table is designed to prevent excessive vibration of the plate. The isolation table is made of a suitable material such as steel and honeycomb but can

be others. The table has a thickness of about 8 inches or preferably less than about 24 inches. In one embodiment, the table is Model 9101-24-85 made by Kinetic Systems of Boston, MA.

The imaging system also has a lamp or illumination assembly 62. The lamp assembly provides for a light source (See reference letter B) to a plurality of elements in the imaging system. For easy reading, the light path is defined by the dotted lines, which are not intended to be limiting. The lamp assembly has a variety of elements such as a Xenon lamp 46. The Xenon lamp provides light at about 320 to 700 nanometers (Prefocused). The Xenon lamp is 175 or 300 Watts. As merely an example, the lamp can be a Lambda Model made by Sutter Instrument Company of Novato, CA.

Referring to Fig. 5B, the lamp assembly also has a cold mirror 58, an excitation filter wheel 48, excitation filter(s) 55, and an excitation light shutter 57. As shown, light is derived from the Xenon lamp, reflects off of the cold mirror 58, traverses through the excitation filter or filters 55, and is controlled by the excitation light shutter 57. The lamp assembly has filter wheel 48, which houses one of a plurality of filters, including excitation filters. The shutter and filter wheel are controlled via control lines 67, which are coupled to a computer 51 or other type of computing device. The control lines 67 are coupled through controller 57A (for element 57) and controller 48A (for element 48) via control line 69 to computer 51.

Preferably, light traverses from the lamp assembly through a light guide 47 to illuminate features within the plate. The light guide is suitably selected to have a flexible member, which can be used to place lamp source at a remote location away from the imaging device. The flexible member substantially keeps any vibration from the lamp assembly away from the imaging device. In some embodiments, the member is at least 1 foot away from the imaging device. The light guide is a guide, which is a flexible hose-type sleeve. The sleeve is filled with a liquid such as an aqueous solution containing chloride or phosphate. A thin layer may be formed on the inside of the sleeve. The layer can be a containing tetrafluoroethylene and hexafluoropropylene, or containing tetrafluoroethylene and perfluoromethyl vinyl ether, or tetrafluoroethylene and perfluoropropyl vinyl ether. An example of such a light guide is described in International Application No. WO/98/38537 filed February 29, 1997, and assigned to NATH, Gunther. The liquid

light guide has less than about 30% transmission loss of the light at a remote location such as the imaging system.

Light is derived from the lamp assembly and directs off of filter 56, which directs the light upward. Filter 56 can be a dichroic and emission filter, as well as others. The light traverses through microscope nosepiece 41C, and traverses through objective spacers 54. An objective 53 magnifies the light toward a predetermined point on the plate 59. The objective can be, for example, made by Zeiss of Jena, Germany, as well as other companies. The objective can be one of a plurality including 1X, 10X, 20X, 40X, and others, depending upon the application. Magnification can be further expanded or contracted by intermediate optics between the objective and the camera. Selection of filter or filters is controlled by computer 51 via control line 75.

The camera 50 captures an image of cells from plate 59. The image is obtained from light scattering off of cells or portions of cells in the plate through objective 53, through objective spacers, through filters 56, which are captured at camera 50. In this preferred embodiment, the camera is a digital camera, but can be an analogue camera. The digital camera is a CCD camera, which has 1280 by 1024 pixels, or more or less. The pixels can be 6.7 microns in dimension or more or less. The camera preferably is substantially free from an external shutter to quickly capture a plurality of images of cells from the plate. The camera is controlled via control line 71 through controller 50A, which connects to computer 51 through control line 70. The present invention can also include other types of image acquisition devices selected from at least an epifluorescence, a confocal, a total-internal reflection, a phase, a Hoffman, a bright field, a dark field, a differential interference contrast, an interference reflection, or multi-photon illumination device.

The present imaging system stores images on a high density memory device 60. The high density memory device is preferably optical, but can also be magnetic. The high density memory device can be any suitable unit that is capable of storing a plurality of images from a plurality of sites in the plate. The memory device can be a compact disk, which would generally use a compact disk burner or the like. Depending upon the embodiment, the high density memory device is used to archive the images that are captured from the camera in the imaging system. Further details

of the imaging system can be found throughout the present specification, and more particularly below.

As merely an example, the present invention can be implemented using the following sequence of steps, which have been described in a journal entry form.

- 5 Here, images are opened and objects are identified based on a background value that has been edited in starting image acquisition. Information is maintained in a spreadsheet or other database format, which has the following information for each object:

Image Name	Image Plane	Image Date and Time
Elapsed Time	Object #	Total area
Pixel area	Area	Hole area
Relative hole area	Standard area count	Perimeter
Length	Breadth	Fiber length
Fiber breadth	Shape factor	Ell. form factor
Inner radius	Outer radius	Mean radius
Average gray value	Total gray value	Optical density
Radial dispersion	Texture Difference Moment	EFA Harmonic 2, Semi-Major Axis
EFA Harmonic 2, Semi-Minor Axis	EFA Harmonic 2, Semi-Major Axis Angle	EFA Harmonic 2, Ellipse Area
EFA Harmonic 2, Axial Ratio	EFA Harmonic 3, Semi-Minor Axis	

10

After computations are done, the log file is saved. In particular, the file is saved in an appropriate place with an appropriate name.

In a specific embodiment, the present invention provides the following detailed example of journal entries, which should not limit the scope of the invention.

Set Up Sequential File Names	Interactive: user sets up prefix name and image storage directory
Open Data Log	Opens a DDE (Excel) File
Annotate Log File	Interactive: experimental information that will go into the first line of the log file of stage positions
Stage (Go to Origin)	Origin is set as the center of well A1
Stage (Move to Absolute Position)	Offset to upper left hand corner of well (1410, 1621)
Stage (Log Position)	
Stage (Scan Wells)	User picks wells to scan: runs 3x3 image collection.jnl.

3X3 IMAGE COLLECTION.jnl

Stage (Scan)	Takes 9 images of well, -1600 motor steps apart from left to right 3 columns and 3 rows, runs FOCUS, COLLECT IMAGE, SAVE SEQUENTIAL FILE NAME.JNL.
--------------	--

5

FOCUS, COLLECT IMAGE, SAVE SEQUENTIAL FILE NAME.jnl.

Stage (Log Position)	Logs stage position of each image
ADC – Focus	Opens up the manual focusing window with whatever focus time is current set
Show Message and Wait	Interactive: user hits enter to continue when done focusing

ADC-Acquire from Digital Camera	Takes Hoechst image
Save Using Sequential File Names	
Close	Closes image window

START IMAGE ANALYSIS.jnl

Low Pass	3x3 convolution of already opened image
Low Pass	3x3
Show Region Statistics	Interactive: Show entire image statistics. Calculate background subtraction value for step 4. by: INTENSITY Average + INTENSITY Std. Dev.
Arithmetic	Interactive: User inputs subtraction value from 3. into the constant Value field
Threshold image	Creates threshold 1 unit above 0 to 4096
Integrated Morphometry – Load State	Loads Start Image Analysis.ima Classifier 100 < area < 200000
Integrated Morphometry – Measure	Interactive: Shows area summary information about all objects. The average number is used as the Standard Area in 8.
Object Standards - Set Object Standards	Interactive: User inputs average area value from 7. into Standard Area box to be used by automated IMA for all images

IMA OBJECTS.jnl

Low Pass	3x3 convolution
Low Pass	3x3 convolution
Arithmetic	This background subtraction value needs to be manually entered into this journal from the value determined in START IMAGE ANALYSIS.jnl step 3
Threshold Image	1 unit above 0
Integrated Morphometry – Load State	Hoechst.IMA Classifier 200 < area < 200000
Integrated Morphometry – Measure	Measures statistical info for all objects
Run Journal	Runs log obj and sum data.jnl

Log obj and sum data.jnl

Integrated Morphometry – Log Data	Logs object data into Sheet 1
Integrated Morphometry – Log Data	Log summary data into Sheet 2

5

COLLECT AUTOMATED IMA DATA IN ONE SPREADSHEET.jnl

Run Journal	Runs OPEN OBJECT LOG DDE FILE.JNL
Loop for all Images in a Directory	Loops IMA OBJECTS.jnl
Close Summary Log	
Close Object Log	User must manually save Excel spreadsheet

OPEN OBJECT LOG DDE FILE.jnl

Open Object Log	Opens a DDE object log into sheet 1 of an Excel spreadsheet
Open Summary Log	Opens a summary log into sheet 2

COLLECT AUTOMATED IMA DATA IN ONE SPREADSHEET 16 BIT IMAGES.jnl

Arithmetic	Interactive: Opens Arithmetic window for user to input background subtraction level from START IMAGE ANALYSIS.jnl step 3
Run Journal	Runs OPEN OBJECT LOG DDE FILE.JNL
Loop for all Images in a Directory	Interactive: Runs IMA OBJECTS 16 bit.jnl. User picks directory from which to choose.

5

IMA OBJECTS 16bit.jnl

Low Pass	3x3 convolution
Low Pass	3x3 convolution
Copy to 8-bit Image	No autoscale, to new untitled image
Save Using Sequential File Name	Saves 8bit image using previously defined Sequential File names.
Arithmetic	This background subtraction value needs to be manually entered into this journal from the value determined in START IMAGE ANALYSIS 16 TO 8 BIT.jnl step 5
Threshold Image	1 unit above 0 to 255

Integrated Morphometry – Load State	Hoecsht. IMA Classifier 200 < area < 200000
Integrated Morphometry – Measure	Measures statistical info for all objects
Run Journal	Runs log obj and sum data.jnl

START IMAGE ANALYSIS 16 to 8 BIT.jnl

Copy to 8-bit Image	No autoscale, to new untitled image
Close	Closes 16 bit image
Low Pass	3x3 convolution
Low Pass	3x3 convolution
Show Region Statistics	Interactive: Show entire image statistics. Calculate background subtraction value for step 6. by: INTENSITY Average + INTENSITY Std. Dev.
Arithmetic	Interactive: User inputs subtraction value from 5. into the constant Value field
Threshold image	Creates threshold by 1 unit above 0 to 255
Integrated Morphometry – Load State	Loads Start Image Analysis.ima Classifier 100 < area < 200000
Integrated Morphometry – Measure	Interactive: Shows area summary information about all objects. The average number is used as the Standard Area in 10.
Object Standards - Set Object Standards	Interactive: User inputs average area value from 9. into Standard Area box to be used by automated IMA for all images

IMA OBJECTS WITH NEW LOG FILE.jnl

Run Journal	OPEN OBJECT LOG DDE FILE.JNL
Run Journal	IMA OBJECTS.jnl
Close Summary Log	
Close Object Log	User must manually save every Excel spreadsheet generated.

INTERACTIVE IMA OBJECTS.jnl

Threshold Image	User manually sets threshold
Integrated Morphometry – Load State	Hoechst.IMA Classifier $200 < \text{area} < 200000$
Integrated Morphometry – Measure	Objects
Integrated Morphometry – Log Data	Into open object.log file

5

COLLECT INTERACTIVE IMA DATA.jnl

Close Object Lo g	
Open Object Log	Interactive
Annotate Log File	Interactive: experimental information that will go into the first line of the object log file
Loop for all Images in Directory	Runs INTERACTIVE IMA OBJECTS.jnl

CHANGE FILTER, COLLECT IMAGE, SAVE SEQUENTIAL FILE
NAME.jnl

Stage (Log Position)	
ADC-Focus	

Show Message and Wait	Interactive – user presses Enter when done focusing
ADC – Acquire from Digital Camera	Hoechst
Save Using Sequential File Name	
Close	Close open image

COLLECT HOECHST AND FITC.jnl

Run Journal	FOCUS, COLLECT IMAGE, SAVE SEQUENTIAL FILE NAME.JNL
Run Journal	CHANGE FILTER, COLLECT IMAGE, SAVE SEQUENTIAL FILE NAME.jnl

3X3 IMAGE COLLECTION HOECHST FITC.jnl

Stage (Scan)	COLLECT HOECHST AND FITC.jnl
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5

AUTOMATED 3X3 IMAGE COLLECTION HOECHST FITC.jnl

Set Up Sequential File Names	Interactive: user sets up prefix name and image storage directory
Open Data Log	Excel DDL files
Annotate Log File	Interactive: experimental information that will go into the first line of the log file of stage positions
Stage (Go to Origin)	Origin is set as the center of well A1
Stage (Move to Absolute Position)	Offset to upper left hand corner of well (1410, 1621)

Stage (Log Position)	
Stage (Scan Wells)	Interactive: user picks wells to scan: runs 3X3 IMAGE COLLECTION HOECHST FITC.jnl

AUTOMATED IMAGE COLLECTION.jnl

Sct Up Sequential File Names	Interactive: user sets up prefix name and image storage directory
Open Data Log	Opens a DDE (Excel) File
Annotate Log File	Interactive: experimental information that will go into the first line of the log file of stage positions
Stage (Go to Origin)	Origin is set as the center of well A1
Stage (Log Position)	
Stage (Scan Wells)	Interactive: user picks wells to scan: runs FOCUS, COLLECT IMAGE, SAVE SEQUENTIAL FILE NAME.JNL. Well to well travel = (-9035, -9035)

5

STARTUP.jnl

Install and Configure Devices	Open Stage Meta Devices
Set Live Video Channel	

Preferences	<u>Measure Objects</u> : Draw failed classifier objects, Exclude objects that touch the edge of the image, Enable Elliptical Fourier Parameters, turn off Warn users when measurement data will be erased <u>Image Saving</u> : Save Tiff/stk using LZW compression <u>Image Windows</u> : Use transparent thresholds.
Configure Default Paths	C:\Metamorph Data C:\Metamorph Data\Common Settings
Load Journal Taskbar	Common.JTB

Nested Journals

Automated 3x3 Image Collection

5

Loop 3x3 image collection

Loop focus, collect image, save sequential file name

Automated 3x3 image collection Hoechst FITC

10

Loop 3x3 image collection Hoechst FITC

loop Collect Hoechst and FITC

focus, collect image, save sequential file name

change filter, collect image, save sequential file name

Automated image collection

15

Loop focus, collect image, save sequential file name

Collect automated IMA data in one Spreadsheet

Open object log DDE file

Loop IMA objects

Log obj and sum data

Collect automated IMA data in one spreadsheet 16 bit images

5 Open object log DDE file

Loop IMA objects 16 bit

Log obj and sum data

Although the above has been generally described in terms of a specific
10 user interface and software code, other user interfaces and code can also be used. One
of ordinary skill in the art would recognize many other variations, alternatives, and
modifications.

Fig. 6 is a simplified diagram 600 of a cleaning and dispensing system
according to an embodiment of the present invention. This system 600 includes a
15 variety of elements such as a dispensing head 609, which is coupled to a plurality of
pipettes 601. The pipettes input and output fluids or solutions from plate 603. The
plate has a plurality of sites, each of which can be used to input cells or a combination
of cells and solution. The system also has elements to house solutions 605, which are
used to manipulate cell samples in the plate. The dispensing head is supported
20 through a support member 607, which is sufficiently rigid to allow for movement of
the head. The dispenser is coupled to the present system in a mechanical and
electrical manner, which provides for a fully integrated system for providing cell
samples to the imaging system according to the present invention.

Fig. 7A illustrates a representative block flow diagram of simplified
25 process steps of a method for determining properties of a manipulation based upon
effects of the manipulation on one or more portions of one or more cells in a
particular embodiment according to the present invention. This diagram is merely an
illustration and should not limit the scope of the claims herein. One of ordinary skill
in the art would recognize other variations, modifications, and alternatives. In step
30 700, one or more samples of cells can be provided. These cells can be live, dead, or
fixed cells, or cell fractions. The cells also can be in one of many cell cycle stages,
including G0, G1, S, G2 or M phase, M phase including the following cell cycle
stages: interphase, prophase, prometaphase, metaphase, anaphase, and telophase.

Cell components tracked in presently preferable embodiments can include proteins, protein modifications, genetically manipulated proteins, exogenous proteins, enzymatic activities, nucleic acids, lipids, carbohydrates, organic and inorganic ion concentrations, sub-cellular structures, organelles, plasma membrane, adhesion complex, ion channels, ion pumps, integral membrane proteins, cell surface receptors, G-protein coupled receptors, tyrosine kinase receptors, nuclear membrane receptors, ECM binding complexes, endocytotic machinery, exocytotic machinery, lysosomes, peroxisomes, vacuoles, mitochondria, Golgi apparatus, cytoskeletal filament network, endoplasmic reticulum, nuclear membrane, proteosome apparatus, chromatin, nucleolus, cytoplasm, cytoplasmic signaling apparatus, microbe specializations and plant specializations.

The following table illustrates some markers and cell components commonly used by embodiments according to the present invention. Other markers can be used in various embodiments without departing from the scope of the invention.

Cell component	Marker	Disease State
Plasma membrane (including overall cell shape)	Carbocyanine dyes Phosphatidylserine Various lipids Glycoproteins	Apoptosis-Cancer Apoptosis-Neural degenerative Ds
Adhesion complexes	Cadherins Integrins Occludin Gap junction ERM proteins CAMs Catenins Desmosomes	Thrombosis Metastasis Wound healing Inflammatory Ds Dermatologic Ds
Ion Channels and Pumps	Na/K Atpase Calcium channels Serotonin reuptake pump CFTR	Cystic fibrosis Depression Congestive Heart Failure Epilepsy

G coupled receptors	β adrenergic receptor Angiotensin receptor	Hypertension Heart Failure Angina
Tyrosine kinase receptors	PDGF receptor FGF receptor IGF receptor	Cancer Wound healing Angiogenesis Cerebrovascular Ds
ECM binding complexes	Dystroglycan Syndecan	Muscular Dystrophy
Endocytotic machinery	Clathrin Adaptor proteins COPs Presenilins Dynamin	Alzheimer's Ds
Exocytotic machinery	SNAREs Vesicles	Epilepsy Tetanus Systemic Inflammation Allergic Reactions
Lysosomes	Acid phosphatase Transferrin	Viral diseases
Peroxisomes/Vacuoles		Neural degenerative Ds
Mitochondria	Caspases Apoptosis inducing factor F1 ATPase Fluorescein Cyclo-oxygenase	Apoptosis Neural degenerative Ds Mitochondrial Cytopathies Inflammatory Ds
Golgi Apparatus	Lens Culinaris DiOC6 carbocyanine dye COPs	

Cytoskeletal Filament Networks	Microtubules Actin Intermediate Filaments Kinesin, dynein, myosin Microtubule associated proteins Actin binding proteins Rac/Rho Keratins	Cancer Neural degenerative Ds Inflammatory Ds Cardiovascular Ds Skin Ds
Endoplasmic Reticulum	SNARE PDI Ribosomes	Neural degenerative Ds
Nuclear Membrane	Lamins Nuclear Pore Complex	Cancer
Proteosome Apparatus	Ubiquityl transferases	Cancer
Chromatin	DNA Histone proteins Histone deacetylases Telomerases	Cancer Aging
Nucleolus	Phase markers	
Cytoplasm	Intermediary Metabolic Enzymes BRCA1	Cancer
Cytoplasmic Signaling Apparatus	Calcium Camp PKC pH	Cardiovascular Ds Migraine Apoptosis Cancer
Microbe Specializations	Flagella Cilia Cell Wall components: Chitin synthase	Infectious Ds

Plant specializations	Choloroplast Cell Wall components	Crop Protection
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Then, in a step 702, one or more samples of the manipulation can be provided to the cells. Manipulations can comprise one or any combination of chemical, biological, mechanical, thermal, electromagnetic, gravitational, nuclear, or temporal factors, for example. For example, manipulations could include exposure to chemical compounds, including compounds of known biological activity such as therapeutics or drugs, or also compounds of unknown biological activity. Or exposure to biologics that may or may not be used as drugs such as hormones, growth factors, antibodies, or extracellular matrix components. Or exposure to biologics such as infective materials such as viruses that may be naturally occurring viruses or viruses engineered to express exogenous genes at various levels. Bioengineered viruses are one example of manipulations via gene transfer. Other means of gene transfer are well known in the art and include but are not limited to electroporation, calcium phosphate precipitation, and lipid-based transfection. Manipulations could also include delivery of antisense polynucleotides by similar means as gene transfection. Other genetic manipulations include gene knock-outs or gene mutations. Manipulations also could include cell fusion. Physical manipulations could include exposing cells to shear stress under different rates of fluid flow, exposure of cells to different temperatures, exposure of cells to vacuum or positive pressure, or exposure of cells to sonication. Manipulations could also include applying centrifugal force. Manipulations could also include changes in gravitational force, including sub-gravitation (the preferred embodiment in outer space). Manipulations could include application of a constant or pulsed electrical current. Manipulations could also include irradiation. Manipulations could also include photobleaching which in some embodiments may include prior addition of a substance that would specifically mark areas to be photobleached by subsequent light exposure. In addition, these types of manipulations may be varied as to time of exposure, or cells could be subjected to multiple manipulations in various combinations and orders of addition. Of course, the type of manipulation used depends upon the application.

Then, in a step 704, one or more descriptors of a state in the portions of the cells in the presence of the manipulation can be determined using the images

collected on the imaging system. Descriptors can comprise scalar or vector values, representing quantities such as area, perimeter, dimensions, intensity, gray level, aspect ratios, and the like. Other types of descriptors include, but are not limited to, one or any combination of characteristics such as a cell count, an area, a perimeter, a
5 length, a breadth, a fiber length, a fiber breadth, a shape factor, a elliptical form factor, an inner radius, an outer radius, a mean radius, an equivalent radius, an equivalent sphere volume, an equivalent prolate volume, an equivalent oblate volume, an equivalent sphere surface area, an average intensity, a total intensity, and an optical density. These descriptors can be average or standard deviation values, or frequency
10 statistics from the descriptors collected across a population of cells. These descriptors can be further reduced using other methods such as principal component analysis and the like. In some embodiments, the descriptors include features from different cell portions or cell types. That is, a first feature can be from a nuclei and a second feature is from another cell structure such as Golgi apparatus, mitochondria, spacing between
15 cell structures or cells themselves, as well as many others.

A presently preferable embodiment uses descriptors selected from the following table. Other descriptors can also be used without departing from the scope of the invention.

Name of Parameter	Explanation/Comments
Count	Number of objects
Area	
Perimeter	
Length	X axis
Width	Y axis
Shape Factor	Measure of roundness of an object
Height	Z axis
Radius	
Distribution of Brightness	
Radius of Dispersion	Measure of how dispersed the marker is from its centroid
Centroid location	x-y position of center of mass
Number of holes in closed objects	Derivatives of this measurement might include, for

	example, Euler number (= number of objects - number of holes)
Elliptical Fourier Analysis (EFA)	Multiple frequencies that describe the shape of a closed object
Wavelet Analysis	As in EFA, but using wavelet transform
Interobject Orientation	Polar Coordinate analysis of relative location
Distribution Interobject Distances	Including statistical characteristics
Spectral Output	Measures the wavelength spectrum of the reporter dye. Includes FRET
Optical density	Absorbance of light
Phase density	Phase shifting of light
Reflection interference	Measure of the distance of the cell membrane from the surface of the substrate
1,2 and 3 dimensional Fourier Analysis	Spatial frequency analysis of non closed objects
1,2 and 3 dimensional Wavelet Analysis	Spatial frequency analysis of non closed objects
Eccentricity	The eccentricity of the ellipse that has the same second moments as the region. A measure of object elongation.
Long axis/Short Axis Length	Another measure of object elongation.
Convex perimeter	Perimeter of the smallest convex polygon surrounding an object
Convex area	Area of the smallest convex polygon surrounding an object
Solidity	Ratio of polygon bounding box area to object area.
Extent	proportion of pixels in the bounding box that are also in the region
Granularity	
Pattern matching	Significance of similarity to reference pattern
Volume measurements	As above, but adding a z axis

Then, in a step 705, a database of cell information can be provided.

Next, in a step 706, a plurality of descriptors can be searched from a database of cell information in order to locate descriptors based upon one of the descriptors of the manipulation. Then, in a step 708, properties of the manipulation are predicted based upon the properties of the located descriptors. Properties can comprise toxicity, specificity against a subset of tumors, mechanisms of chemical activity, mechanisms of biological activity, structure, adverse biological effects, biological pathways, clinical effects, cellular availability, pharmacological availability, pharmacodynamic properties, clinical uses and indications, pharmacological properties, such as absorption, excretion, distribution, metabolism and the like.

In a particular embodiment, step 706 comprises determining matching descriptors in the database corresponding to a prior administration of the manipulation to the descriptors of the present administration of the manipulation. In a particular embodiment according to the present invention, combinations of measurements of scalar values can provide predictive information. A database can be provided having one or more "cellular fingerprints" comprised of descriptors of cell-substance interactions of drugs having known mechanisms of action with cells. Such descriptors can be analyzed, classified, and compared using a plurality of techniques, such as statistical classification and clustering, heuristic classification techniques, a technique of creating "phylogenetic trees" based on various distance measures between descriptors from various drugs. In this embodiment, numeric values for the descriptors can be used by comparison techniques. A phylogenetic tree can be created that illustrates a statistical significance of the similarity between descriptors for the drugs in the database. Because the drugs used to build the initial database are of known mechanism, it can be determined whether a particular scalar value in a descriptor is statistically predictive. Finally, a compound descriptor with no known mechanism of action can be queried against the database and be statistically compared and classified among the drugs in the database that the compound most resembles.

In a particular embodiment, relationships between measured morphological properties of images and physiological conditions can be determined. Relationships can include, for example, treatment of different cell lines with chemical compounds, or comparing cells from a patient with control cells, and the like. In a presently preferable embodiment, comparisons can be performed on acquired image

features. Some embodiments can comprise statistical and neural network - based approaches to perform comparisons of various features. The foregoing is provided as merely an example, and is not intended to limit the scope of the present invention.

Other techniques can be included for different types of data.

- 5 In some embodiments, classification, clustering and other types of predictive data analysis can be performed on features extracted from cell images. In a presently preferable embodiment, statistical procedures for comparisons, classification and clustering are performed on data obtained from imaging cells.

Fragments of data preparation and pre-formatting (S language):

```
10       >tmp.frame <- Generic.Summary  
      >names1 <- paste("Cell.line.5", tmp.names, sep=".")  
      > by.compound.matrix <- as.matrix(arranged.by.compound)
```

- Example of the code for principal component analysis (data
15 preparation) using S language:

```
      all.data.princomp <- menuPrincomp(data =  
      by.compound.matrix, scores = T, cor = "Correlation",  
      na.action = T, print.short = T, print.importance = T,  
      print.loadings = T, cutoff.loadings = 0.1,  
20       plot.screepplot = T, plot.loadings = T, plot.biplot = T,  
      plot.biplot.choices = c(1,2), predict.p = F)
```

Example of clustering using a divisive hierarchical clustering
algorithm:

```
25       > div.hier.2.manhattan.cluster$call  
      diana(x = tmp.sum.by.comp, diss = F, metric =  
      "manhattan",  
          stand = T, save.x = T, save.diss = T)
```

- 30 Another embodiment utilizes existing tools for biological sequence similarity searches, classification, and phylogenetic analysis. In a particular embodiment, numbers in a numerical descriptor can be substituted by one or more of nucleic acid or amino acid codes according to a one of several sets of rules. Once

converted into a corresponding nucleotide or amino acid sequence representation, the fingerprints can be analyzed and compared using software and algorithms known in the art for genetic and peptide sequence comparisons, such as GCG, a product of Genetics Computer Group, with company headquarters in Madison WI. Select

5 embodiments comprising such approaches enable the use of a broad array of sophisticated algorithms to compare, analyze, and cluster gene and protein sequences. Many programs performing this task are known to those of ordinary skill in the art, such as for example, the PHYLIP (PHYlogeny Interference Package) a package of programs for inferring phylogenies (evolutionary trees) described in (Feldenstein, J.
10 1996 Methods Enzymol 266:418-427 and Feldenstein, J. 1981 J. Mol. Evol. 17(6):368-376).

Embodiments can perform such analysis based upon factors such as numerical value, statistical properties, relationships with other values, and the like. Further details of a step of manipulation are noted more particular below.

15 Fig. 7B illustrates a representative block flow diagram of simplified process steps for determining one or more descriptors of a state in the portions of the cells in the presence of the manipulation of step 704 of Fig. 7A in a particular embodiment according to the present invention. This diagram is merely an illustration and should not limit the scope of the claims herein. One of ordinary skill
20 in the art would recognize other variations, modifications, and alternatives. In a step 712, an image of a cell portion is obtained. In some embodiments, the cell portion is visualized with a fluorescently labeled marker that is specific for the portion or portions of interest. A cell portion can include, for example, one or more of the following: nuclei, Golgi apparatus, and other features. The cell portion may vary in
25 select embodiments according to the invention. Then, in a step 714, a digitized representation of the image obtained in step 712 is determined. In some embodiments, steps 714 and step 712 can comprise a single step. These embodiments use a digital imaging means such as a digital camera, to obtain a digital image of the target directly. Next, in a step 716, the digital representation of the image is
30 processed to obtain image features. Image features can include such quantities as area, perimeter, dimensions, intensity, aspect ratios, and the like. Then, in a step 718 descriptors can be determined from the image features. Descriptors can comprise scalar or vector quantities and can comprise the image features themselves, as well as

composed features, such as shape factor derived by a relationship $4\pi * \text{area} / \text{perimeter}$, and the like. Descriptors can also comprise statistical quantities relating to feature characteristics across a population of cells, such as a standard deviation, and average, and the like.

5 In a preferred embodiment, cells can be placed onto a microscope, such as a Zeiss microscope, or its equivalent as known in the art. A starting point, named Site A01, is identified to the microscope. A plurality of exposure parameters can be optimized for automated image collection and analysis. The microscope can automatically move to a new well, automatically focus, collect one or more images, at
10 one or more wavelengths, move to a next well, and repeat this process for all designated wells in a multiple well plate and for multiple plates. A file having a size and an intensity distribution measurement for each color and rank for each well can then be created for the images acquired. Based on this information, a user or a computer can revisit sites of interest to collect more data, if desired, or to verify
15 automated analysis. In a presently preferred embodiment, image automatic focus and acquisition can be done using computer software controlling the internal Z-motor of the microscope. Images are taken using a 10x, 20x, or 40x air long working distance objectives. Sometimes multiple images are collected per well. Image exposure times can be optimized for each fluorescent marker and cell line. The same exposure time
20 can be used for each cell line and fluorescent marker to acquire data.

Fig. 7C illustrates a representative block flow diagram of simplified process steps for obtaining images of cell portions of step 712 of Fig. 7B in a particular embodiment according to the present invention. This diagram is merely an illustration and should not limit the scope of the claims herein. One of ordinary skill
25 in the art would recognize other variations, modifications, and alternatives. The method is generally outlined by the steps below:

(1). In a step 720, a sample is provided to the imaging device. Samples can be provided in 96 well plates and the like. The sample may be loaded into a microscope, such as a Zeiss microscope or equivalent.

30 (2). In a step 722, a set of optical filters is selected to shine light of the appropriate wavelength to illuminate the first sample, which may be contained in a first well designated A01.

(3). In a step 724, an automatic focusing procedure is performed for the site. In a particular embodiment, the internal z-motor of the microscope which is attached to the objective nosepiece is used for automatic focusing of the microscope. In an alternative embodiments, the plate holding the samples is moved to perform
5 automatic focusing of the microscope, or focusing can be performed by moving optical components attached to the microscope and the like.

(4). In a step 726, images are collected for the site. Images can be collected for every color at every site. Present embodiments can provide images for up to four colors. However, embodiments are contemplated that can provide more
10 colors by using either a monochromator coupled with excitation filters which are on a filter wheel, or by digitally separating overlapping fluorophores. Those knowledgeable in the field will know that given calibration images of single fluorophores, a look-up table can be devised which will allow for the digital removal of fluorescence bleed-through of fluorescence which may occur in optical channels
15 other than the one for which that filter has been optimized in instances of using more than one fluorophore at once. Cell growth and density information is also collected. Cell density is determined by what percentage of the area being imaged is inhabited by cells. In some embodiments, imaging can be facilitated using one or more biosensors, molecules such as non-proteins, i.e., lipids and the like, that are
20 luminescently tagged. However, some embodiments can also use fluorescence polarization and the like. Fluorescence polarization is a homogeneous fluorescence technology where the excited state of the molecule lasts much longer than in normal fluorescence, taking seconds to minutes to reach equilibrium, obliterating the need to wash away fluorescence markers that are not specifically bound to a marker. Further,
25 embodiments can detect differences in spectral shifts of luminescent markers. Some fluorescence markers, such as Nile Red sold by Molecular Probes of Eugene, OR, will change its emission peak wavelength depending on its environment. One can detect these changes by monitoring the level of fluorescence at both wavelengths and reading out at ratio of the two.

(5). In a step 728, a determination is made whether more fields of view
30 need to be taken for a particular color. If this is so, then processing continues at step 726 at a new site. Otherwise, processing continues with a decisional step 730.

Images can now be taken by repeating step 726. In a preferred embodiment 4 to 9 images are collected at each site.

(5). In a step 730, a determination is made whether more optical configurations need to be taken in order to obtain images for all differently-marked cell portions the sample. If this is so, then in a step 732 a new optical configuration is determined. Images for the new optical configuration can now be taken by repeating steps 726 and 728.

(6). In a decisional step 734, after all optical configurations and images for fields of view in a sample have been obtained, a determination is made whether any further samples remain to be analyzed. If so, a new sample is brought into view and processing continues with step 720. Otherwise, image processing is complete. In a presently preferable embodiment, image data can be stored on a CD ROM using a CD ROM burner, such as CRW4416 made by Yamaha of Japan. However, other mass storage media can also be used.

Fig. 7D illustrates a representative block flow diagram of simplified process steps for processing digitized representations of step 716 of Fig. 7B in a particular embodiment according to the present invention. This diagram is merely an illustration and should not limit the scope of the claims herein. One of ordinary skill in the art would recognize other variations, modifications, and alternatives. The method is generally outlined by the steps below:

(1). In a step 740, a digitized image input is preprocessed . Preprocessing might include, but is not limited to, such operations as background subtraction, thresholding, smoothing, adoptive filtering, edge enhancements, contrast enhancements, histogram equalization. A particular combination of preprocessing steps can be applied to images in successive steps or in parallel to copies of the image.

A simplified example of a smoothing and background subtraction procedure in a MatLab language is presented in computer code below:

```
function Isubtracted = cmBackgrSubtrl(I,k)
% cmBackgrSubtrl(I,k) - simple flat background (=modal*k)
subtraction
% Y = cmBackgrSubtrl(I, k) - image Y is generated by
```



```

    % subtraction (with saturation) of modal pixel value of I
    multiplied by k
    % DEFAULT - k=1
    %
5   if (nargin == 1)
        k=1;
    end
    if (size(k)~=1)
        error('cmBackgrSubtrl: parameter k should be a number.
10  Exiting...');
    end

    %modpixnum = floor(size(I(:),1)/2);
    %sortedval = sort( double(I(:)) );
15  %modpixel = sortedval(modpixnum);
    modpixel = median(double(I(:)));
    bg = k*modpixel;

    Isubtracted = mmsubm( uint8(I), uint8(round(ones(
20  size(I))*k*modpixel )) );

```

An example of a procedure for thresholding in computer code (MatLab) is presented below:

```

function thresh = GetThreshByPerim1(I, M)
25 % GetThreshByPerim1(I) Finds optimal thresholding value
    for image I
    % N = GetThreshByPerim1(I) Finds thresholding value N for
    image I
    % N = GetThreshByPerim1(I, M) - tests threshold values up
30  to M
    % DEFAULT M = maximum pixel value in I
    % note that GetThreshByArea is significantly faster
    % finds a threshold value that causes the maximal change
    in the

```

```

% total perimeter of the objects (Russ ????)
% see Matlab_Auto_threshold1_1-23-99.doc for more details
% Note: works somewhat better on SMOOTH images (i.e.
medfilt2(I, [3 3]) two times

5
if (nargin == 0)
    error (strcat( mfilename, ' : at least one parameter
required')));
elseif (nargin == 1)
10    M = double(max(I(:)));      %test thresholds up to
    maximum pixel value in I
    elseif (nargin > 2)
        error (strcat (mfilename, ' : too many parameters'));
    end
15
    if (size(M)>1)
        error (strcat(mfilename, ' : argument M should be a
number')));
    end
20
    Minval = double( min(I(:)));
    step = 1;

    %generate vertical vector perims with total perimeters of
25 objects at different
    %threshold values
    for i=Minval : step : M
        bwI = im2bw(I, i/255);
        prI = bwperim(bwI);
30        pr = sum(prI(:));
        if (exist('perims', 'var') == 0) %perims is yet
undefined
            perims = pr;
        else

```

```

        perims = cat(1, perims, pr);
    end
end

5  % vector prdiffs contains differences between successive
    perimeters
    prdiffs = diff(perims);
    mindecrease = min(prdiffs);
    minvalues = find(prdiffs == mindecrease);
10  index_of_mindecrease = minvalues(1);
    thresh = index_of_mindecrease + 1;

    % =====end GetThresh1=====

15  Thresholding provides a specific intensity, such that pixels darker than
    the threshold are deemed black, and pixels lighter than the threshold are considered
    white. The thresholded image can be processed using binary image processing
    techniques in order to extract regions.

    (2). In a step 742+744, the digitized image input is subjected to object
20  identification. This can be accomplished by a variety of procedures, for example by
    thresholding or edge detection and subsequent morphological opening and closing.
    Edge detection can be accomplished by means of gradient-based or zero-crossing
    methods, such as Sobel, Canny, Laplassian, Perwitt, and other methods.

    An example of object identification procedure based on Canny edge
25  detection (in MatLab language) is presented below:

    function Imask = cmMaskDNA1(I);
    % cmMaskDNA1 - generates binary mask for cell nuclei
    through edge detection
30  % Imask = cmMaskDNA1(I)
    % PARAMETERS
    %   I - intensity image (grayscale)
    % OUTPUT
    %   Imask - BW image with objects from I

```

```

%
% For more details see Notebook Matlab_DNA_masking1_1-22-
99.doc
% Uses SDC Morphology Toolbox V0.7
5
if (nargin ~= 1)
    error('Wrong number of input parameters');
end
if (nargout ~= 1)
10    error('Wrong number of output parameters: one output
argument should be provided');
end

15  Imask = edge(I, 'canny');
    Imask = mm dil(Imask, mmsecross(1));
    Imask = mmero ( mmc lohole(Imask,mmsecross(1)));
    Imask = mmedgeoff(Imask, mmsecross(1));
    % note that mmedgeoff this command removed FILLED OBJECTS
20  but not touching OUTLINES.
    % these outlines can be removed by filtering:
    Imask = medfilt2(Imask, [5 5]);

    %=====end cmMaskDNA1
25  =====

```

However, embodiments can also use other techniques, such as Fast Fourier Transforms (FFT) and the like as known in the art without departing from the scope of the present invention.

30 (3). In a step 746, a plurality of region features can be determined. For example, in a representative embodiment, image features can include such quantities as area, perimeter, dimensions, intensity, aspect ratios, and the like. Features not directly related to individual objects are also being extracted.

An example of a procedure for extraction of some of the features (MatLab language) is presented below:

```

function OData = cmGetObjectsData(I, Ilabel)
5  % cmGetObjectsData returns array measurements of objects
   in image "I" masked by "Ilabel"
   % EV 2-3-99; 2-10-99
   % OData = cmGetObjectsData(I, Ilabel) returns an array of
   morphological and intensity measurements
10  %   taken from a grayscale image "I". Objects are
   identified on a mask image Ilabel, usually
   %   created by bwlabel()
   % OUTPUT:
   % Each row in the output array OData represents
15  individual object
   % columns contain the following measurements:
   %
   %   1 - Index ("number" of an object);      8 -
   Solidity;
20  %   2 - X coordinate of the center of mass; 9 - Extent;
   %   3 - Y coordinate      "-"; 10 - Total
   Intensity;
   %   4 - Total Area (in pixels);              11 - Avg.
   Intensity;
25  %   5 - Ratio of MajorAxis/MinorAxis;        12 - Median
   Intensity;
   %   6 - Eccentricity;                        13 - Intensity of
   20% bright pixel
   %   7 - EquivDiameter;                      14 - Intensity of
30  80% bright pixel
   %
   % For details on morphological parameters see information
   on MatLab imfeature();

```

```
% Intensity parameters are either obvious or are
documented in comments in this file.

if (nargin ~= 2)
5   error ('function requires exactly 2 parameters');
end
if (nargout ~= 1)
    error ('function has 1 output argument (array X by
14) ');
10 end

% finished checking arguments

% first collect morphological parameters in a structure
15 array:
ImStats = imfeature(Ilabel, 'Area', 'Centroid',
'MajorAxisLength',...
    'MinorAxisLength', 'Eccentricity', 'EquivDiameter',
...
20 'Solidity', 'Extent', 8 );

% now convert it into array (matrix) while collecting
intensity data for each object:

25 %preallocate output array:
numobjects = size(ImStats, 1);
OData = zeros(numobjects, 14);
%now convert ImStats into array and add intensity data to
it
30 for k=1:numobjects
    OData(k, 1) = k;
    OData(k, 2) = ImStats(k).Centroid(1);
    OData(k, 3) = ImStats(k).Centroid(2);
    OData(k, 4) = ImStats(k).Area;
```

```
    OData(k, 5) = (ImStats(k).MajorAxisLength) /  
    (ImStats(k).MinorAxisLength);  
    OData(k, 6) = ImStats(k).Eccentricity ;  
    OData(k, 7) = ImStats(k).EquivDiameter;  
5    OData(k, 8) = ImStats(k).Solidity;  
    OData(k, 9) = ImStats(k).Extent;  
  
    % now collect and assign intensity parameters from  
    image I  
10  
    object_pixels = find( Ilabel == k);  
    object_area = size(object_pixels, 1); %same as total  
    number of pixels in the object  
    object_intensities = double(I(object_pixels)); %  
15 need to convert to double to do math  
    sorted_intensities = sort(object_intensities); %  
    will need to get median, 20% and 80% pixels  
    total_intensity = sum(object_intensities, 1);  
    avg_intensity = total_intensity / object_area;  
20    median_intensity = sorted_intensities( floor(  
    object_area/2 ) + 1 );  
    pix20 = sorted_intensities( floor(object_area*0.2)+1  
    ) ; %brightest pixel among dimmest 20%  
    pix80 = sorted_intensities( floor(object_area*0.8)+1  
25    ) ;  
  
    OData(k, 10) = total_intensity;  
    OData(k, 11) = avg_intensity;  
    OData(k, 12) = median_intensity;  
30    OData(k, 13) = pix20; %brightest pixel among dimmest  
    20%  
    OData(k, 14) = pix80; %dimmest pixel among brightest  
    20%  
end %for
```

```
%===== end function
cmGetObjectsData() =====
```

5 (4). In a step 748, quantitative descriptors, characterizing cell state are calculated based on the feature measurements extracted at step 746. For example, histogram distribution of intensities of cell nuclei provides information about the population cell cycle stages.

10 In a particular embodiment according to the present invention, data analysis techniques for describing the fluorescence patterns of cell portions in multiple cell lines in the presence and absence of compounds are provided. Automated image analysis techniques can include determining one or more regions from around nuclei, individual cells, organelles, and the like, called "objects" using a thresholding function. Objects that reside on the edge of an image can be included or
15 excluded in various embodiments. An average population information about an object can be determined and recorded into a database, which can comprise a database text file or Excel spreadsheet, for example. However, embodiments can use any recording means without departing from the scope of the present invention. Values measured can be compared to the visual image. One or more types of numerical
20 descriptors can be generated from the values. For example, descriptors such as a number of objects, an average, a standard deviation of objects, a histogram (number or percentage of objects per bin, average, standard deviation), and the like can be determined.

25 In a particular embodiment according to the present invention, data can be analyzed using morphometric values derived from any of a plurality of techniques commonly known in the art. For example, a software package called MetaMorph Imaging System, provided by Universal Imaging Corporation, a company with headquarters in West Chester, PA and NIH Image, provided by Scion Corporation, a company with headquarters in Frederick, Maryland.

30 Fluorescent images can be described by numerical values, such as for example, an area, a fluorescence intensity, a population count, a radial dispersion, a perimeter, a length, and the like. Further, other values can be derived from such measurements. For example, a shape factor can be derived according to a relationship

$4\pi \times \text{area} / \text{perimeter}$. Other values can be used in various embodiments according to the present invention. Such values can be analyzed as average values and frequency distributions from a population of individual cells.

In a particular embodiment according to the present invention, techniques for the automatic identification of mitotic cells are provided. Image analysis techniques employing techniques such as multidimensional representations, frequency-based representations, multidimensional cluster analysis techniques and the like can be included in various embodiments without departing from the scope of the present invention. Techniques for performing such analyses are known in the art and include those embodied in MatLab software, produced by MathWorks, a company with headquarters in Natick, MA.

Scalar values providing efficacious descriptors of cell images can be identified using the techniques of the present invention to perform predictive analysis of drug behavior. In a presently preferred embodiment, a plurality of heterogeneous scalar values can be combined to provide descriptors for each manipulation. By applying predictive analysis routines to the collections of these descriptors, predictive information about any number of manipulations and cell interactions can be extracted.

Fig. 7E illustrates a representative block flow diagram of simplified process steps for analyzing image feature values to obtain descriptors of cell state of step 718 of Fig. 7B in a particular embodiment according to the present invention. This diagram is merely an illustration and should not limit the scope of the claims herein. One of ordinary skill in the art would recognize other variations, modifications, and alternatives. Fig. 7E illustrates an input data of descriptors of known manipulations 319. A step 320 of reformatting and transforming data 319 to formats suitable for analysis is performed. Additionally, a "cleaning" process can eliminate outlying data points and the like in the data. Then, in a step 322, a decision is made whether to continue with step 324 or with step 326 based upon determining a particular type of analysis appropriate for the present application or particular type of prediction. If decisional step 322 determines processing should continue with step 324, then, in that step, an error estimate using a set of test descriptors is performed to estimate the quality of a prediction and processing continues with step 320. Once an optimal prediction is achieved, processing continues with step 326. In step 326, optimal transformation parameters and prediction methods are selected for use in

steps 328 and 330 which analyze data about an unknown manipulation. In a step 328, a solution is generated based upon any of techniques including training a neural network, solving a mathematical equation, applying decision tree rules and/or the like. In a step 330, an input data set of unknown descriptors 318 is reformatted and
5 transformed based upon the optimal transformation parameters selected in step 326 using the transformation procedures in steps 320, 322 and 324. In a step 332, predictions techniques are applied to the reformatted manipulations from step 330 and the solution generated in step 328 and a plurality of properties of known manipulations 317 (e.g., therapeutic properties, and the like) in order to determine a
10 prediction of properties of unknown manipulation 316.

Fig. 7F illustrates a representative block flow diagram of simplified process steps for a method of mapping a manipulation of cells to a physiological characteristic in a particular embodiment according to the present invention. This diagram is merely an illustration and should not limit the scope of the claims herein.
15 One of ordinary skill in the art would recognize other variations, modifications, and alternatives. The method is generally outlined by the steps below:

(1) In a step 750, a plurality of cells, e.g., dead, live, cell fractions or mixtures of cells are provided.

(2) Then, in a step 752, the plurality of cells is manipulated, where
20 manipulation occurs using a source(s) from one or a combination selected from an electromagnetic, electrical, chemical, thermal, gravitational, nuclear, temporal, or a biological source.

(3) Next, in a step 754, a feature value is captured from the plurality of cells. The feature value can include one or any combination of characteristics such as
25 cell count, area, perimeter, length, breadth, fiber length, fiber breadth, shape factor, elliptical form factor, inner radius, outer radius, mean radius, equivalent radius, equivalent sphere volume, equivalent prolate volume, equivalent oblate volume, equivalent sphere surface area, average intensity, total intensity, and optical density. This list is not meant to be limiting.

(4) Then, in a step 756, a degree of presence of one or more feature values is assigned for each manipulation.
30

(5) In a step 758, the feature values from the plurality of cells are stored in memory locations. From the memory locations the values can be used for

statistical analyses to produce predictive information about the relatedness of the descriptors of the manipulations to one another. This information is used to infer properties of the manipulations.

Fig. 7G illustrates a representative block flow diagram of a simplified process steps for a method for populating a database with manipulated biological cell information in a particular embodiment according to the present invention. This diagram is merely an illustration and should not limit the scope of the claims herein. One of ordinary skill in the art would recognize other variations, modifications, and alternatives. The method is generally outlined by the steps below:

(1) In a step 760, a plurality of cells in various stages of the cell cycle, A montage image that was used as a source to generate data in Appendix A is presented in Fig. 12., such as for example, the stages of interphase, prophase, metaphase, anaphase, and telophase are provided.

(2) Then, in a step 762, each of the cells in the various stages of mitotic development is manipulated.

(3) Next, in a step 764, an image of the plurality of manipulated cells is captured using image acquisition techniques in order to provide a morphometric characteristic of each of the manipulated cells.

(4) As a preferable option, in a step 766, an image database may be populated with the image of the plurality of manipulated cells.

(5) Following step 764 or optional step 766, a morphological value is calculated from the image in a step 768.

(6) In a step 770, the database is populated with the morphological value.

Fig. 7H illustrates a representative block flow diagram of simplified process steps for a method for populating a database with manipulated biological information, e.g., image acquisition parameters, image feature summary information, and well experimental parameters in a particular embodiment according to the present invention. This diagram is merely an illustration and should not limit the scope of the claims herein. One of ordinary skill in the art would recognize other variations, modifications, and alternatives. Fig. 7H illustrates a step 780 in which cells are placed into site on a plate and a manipulation is applied. Then, in a step 781 an image is taken of the cells. In step 782, the image is transferred to an image archive

database. Then, in a step 783, well experimental parameters are entered into the database 787. Well experimental parameters can include cell type, manipulation and the like. In a step 784, image acquisition parameters are transferred to database 787. Image acquisition parameters can include file name, fluorophores and the like. In a step 785, the image acquired in step 781 is analyzed. Then, in step 786, an image feature summary from the analysis step 785 is transferred to database 787.

In step 788, a lookup table for all analyses is provided to database 787. The lookup table provides information about the analyses. In a step 789, a query of database 787 for process data is performed. The results are reformatted. Then in a step 790, the database 787 is queried. Next, in a step 791, features of the manipulations stored in the database are combined and reduced. Next, in a step 793, reduced features of step 791 can be compared. In a step 792, the results of step 793 are recorded in database 787. Then, in a step 794, a report of predictions based on comparisons performed in step 793 is generated.

Fig. 7I illustrates a representative block flow diagram of simplified process steps for acquiring images of manipulated biological information, e.g., cells, cell tissues, and cell constituents in a particular embodiment according to the present invention. This diagram is merely an illustration and should not limit the scope of the claims herein. One of ordinary skill in the art would recognize other variations, modifications, and alternatives. Fig. 7I illustrates a step 770 in which a user sets up an image analysis procedure. Then, in a step 772, an image is read into image analysis software. Next, in a step 774, patterns and objects are identified in the image using one or more algorithms. Next, in a step 776, sets of features are extracted from the image. Then, in a step 778, feature information, descriptor values and the like are exported to the database, such as database 787 of Fig. 7H, for recording. Next, in a decisional step 779, a determination is made whether any more images should be taken. If this is so, processing continues with step 772. Otherwise, image acquisition processing is completed.

Fig. 7J illustrates a representative block flow diagram of simplified process steps for populating, acquiring and analyzing images of manipulated biological information in a particular embodiment according to the present invention. This diagram is merely an illustration and should not limit the scope of the claims herein. One of ordinary skill in the art would recognize other variations,

modifications, and alternatives. Fig. 7J illustrates a step 300 of placing a plate onto an imaging stage and reading a bar code. Then, in a step 301 an autofocus procedure is performed. Next, in a step 302, a first optical filter configuration is selected and an image is collected. Then, in a decisional step 303, a determination is made whether
5 more than one image per optical configuration can be taken. If so, then, in a step 304, a new position within the well is targeted and another image is collected. Then, in a decisional step 305, a determination is made whether any more images need to be collected. If this is so, step 304 is repeated until all images for a particular well have been collected. After one or more images are collected for the well, in a step 306, the
10 stage is returned to a starting position within the well, and a montage is created from collected images. The results are named with a unique file name and stored.

In a decisional step 307, a determination is made whether any more optical channels in the well can be imaged. If this is so, then in a step 308 the next optical filter configuration is selected and an image is collected. Processing then
15 continues with decisional step 303, as described above. Otherwise, if no further optical channels in the well can be imaged, then in a decisional step 309 a determination is made whether any wells remain to be imaged. If not all wells have been imaged, then in a step 310, the stage moves to the next well and processing continues with step 301, as described above. Otherwise, if all wells on the plate have
20 been imaged, then in a decisional step 311, a determination is made whether any more plates can be processed. If this is so, then processing continues with step 300 as described above. Otherwise, in a step 312, the information is stored to a CD or other storage device as a backup.

Fig. 7K illustrates a representative block flow diagram of simplified
25 process steps compound based upon information about effects of one or more known compounds on a cell population in a particular embodiment according to the present invention. This diagram is merely an illustration and should not limit the scope of the claims herein. One of ordinary skill in the art would recognize other variations, modifications, and alternatives. Fig. 7K illustrates a step 340 of populating a database
30 with descriptors for known compounds. Such descriptors can be determined from imaging the cell population. However, in some embodiments, descriptors can be derived by measurements and combinations of measurements and the like. Then, in a step 342, descriptors for the unknown compound are determined from imaging a

second cell population. The second cell population has been treated with the unknown compound. Then, in a step 344, a relationship between the descriptors determined from the unknown compound with the descriptors determined from the known compounds can be determined. Finally, in a step 346, an inference can be made about the unknown compound based upon the descriptors of the known compounds from the relationship determined in step 344.

Accordingly, the present invention provides a novel database design. In a particular embodiment according to the present invention, a method for providing a database comprises measurement of a potentially large number of features of one or more sub-cellular morphometric markers. Markers can be from any of a large variety of normal and transformed cell lines from sources such as for example, human beings, fungi, or other species. The markers can be chosen to cover many areas of cell biology, such as, for example markers comprising the cytoskeleton of a cell. The cytoskeleton is one of a plurality of components that determine a cell's architecture, or "cytoarchitecture". A cytoarchitecture comprises structures that can mediate most cellular processes, such as cell growth and division, for example. Because the cytoskeleton is a dynamic structure, it provides a constant indication of the processes occurring within the cell. The cytoarchitecture of a cell can be quantified to produce a one or more scalar values corresponding to many possible cellular markers, such as cytoskeleton, organelles, signaling molecules, adhesion molecules and the like. Such quantification can be performed in the presence and absence of drugs, peptides, proteins, anti-sense oligonucleotides, antibodies, genetic alterations and the like. Scalar values obtained from such quantification can provide information about the shape and metabolic state of the cell.

In a presently preferred embodiment, scalar values can comprise morphometric, frequency, multi-dimensional parameters and the like, extracted from one or more fluorescence images taken from a number of cellular markers from a population of cells. Two or more such scalar values extracted from a plurality of cell lines and markers grown in the same condition together comprise a unique "fingerprint" or descriptor that can be incorporated into a database. Such cellular descriptors will change in the presence of drugs, peptides, proteins, antisense oligonucleotides, antibodies or genetic alterations. Such changes can be sufficiently unique to permit a correlation to be drawn between similar descriptors. Such

correlations can predict similar properties or characteristics with regard to mechanism of action, toxicity, animal model effectiveness, clinical trial effectiveness, patient responses and the like. In a presently preferred embodiment, a database can be built from a plurality of such descriptors from different cell lines, cellular markers, and compounds having known mechanisms of action (or structure, or gene response, or toxicity).

The present invention also provides database and descriptor comparisons according to other embodiments. In a particular embodiment according to the present invention, measurement of scalar values or features can provide predictive information. A database can be provided having one or more "cellular fingerprints" comprised of descriptors of cell substance interactions of drugs having known mechanisms of action with cells. Such descriptors can be compared using a plurality of techniques, such as a technique of creating "phylogenetic trees" of a statistical similarity between the descriptors from various drugs. In a present embodiment, scalar, numeric values can be converted into a nucleotide or amino acid letter. Once converted into a corresponding nucleotide representation, the descriptors can be analyzed and compared using software and algorithms known in the art for genetic and peptide sequence comparisons, such as GCG, a product of Genetics Computer Group, with company headquarters in Madison WI. In an alternative embodiment, numeric values for the fingerprints can be used by comparison techniques. A phylogenetic tree can be created that illustrates a statistical significance of the similarity between descriptors for the drugs in the database. Because the drugs used to build the initial database are of known mechanism, it can be determined whether a particular scalar value in a descriptor is statistically predictive. Finally, a compound fingerprint with no known mechanism of action can be queried against the database and be statistically compared and classified among the drugs in the database that the compound most resembles.

In a particular embodiment, relationships between measured morphometric properties and features of images and physiological conditions can be determined. Relationships can include, for example, treatment of different cell lines with chemical compounds, or comparing cells from a patient with control cells, and the like. In a presently preferable embodiment, a clustering can be performed on acquired image descriptors. Some embodiments can comprise statistical and neural

network - based approaches to perform clustering and comparisons of various descriptors. The foregoing is provided as merely an example, and is not intended to limit the scope of the present invention. Other techniques can be included for different types of data. In some embodiments, clustering and comparing can be performed on features extracted from cell images. In a presently preferable embodiment, procedures for comparisons and phylogenetic analysis of biological sequences can be applied to data obtained from imaging cells.

Select embodiments comprising such approaches enable the use of a broad array of sophisticated algorithms to compare, analyze, and cluster gene and protein sequences. Many programs performing this task are known to those of ordinary skill in the art, such as for example, the program Phylip, available at <http://evolution.genetics.washington.edu/phylip.html>, and other packages listed at <http://evolution.genetics.washington.edu/phylip/software.html>. However, select embodiments according to the present invention can comprise a technique of statistical classification, statistical clustering, distance based clustering, linear and non-linear regression analysis, self-organizing networks, and rule-based classification.

Embodiments can perform such analysis based upon factors such as numerical value, statistical properties, relationships with other values, and the like. In a particular embodiment, numbers in a numerical descriptor can be substituted by one or more of nucleic acid or amino acid codes. Resulting "pseudo-sequences" can be subjected to analysis by a sequence comparison and clustering program.

Other types of databases can also be provided according to other embodiments. The database includes details about the properties of a plurality of standard drugs. When the descriptor of a test compound is compared to the database, predictions about the properties of the test compound can be made using any known property of the other compounds in the database. For example, properties about a compound in the database could include structure, mechanism of action, clinical side effects, toxicity, specificity, gene expression, affinity, pharmacokinetics, and the like. The descriptor of a compound of unknown structure from a natural products library could be compared to the descriptors of compounds with known structure and the structure could be deduced from such a comparison. Similarly, such information could lead to better approaches to drug discovery research including target validation

and compound analogizing, as well as pre-clinical animal modeling, clinical trial design, side effects, dose escalation, patient population and the like.

According to the present invention, databases can be integrated with and complementary to existing genomic databases. Differential genomic expression strategies can be used for drug discovery using database technology. In one particular embodiment, cell data and cellular response data can be associated with a genetic expression profile assay to form a single assay. Live cells expressing fluorescence markers can be treated with a drug, imaged and analyzed for morphometry; and then analyzed for mRNA for expression. Such embodiments can provide rapid development of tools to link cellular behavior with functional genomics.

Database methods according to the present invention can be used to predict gene function and to assist in target validation. Databases that include genetic diversity, i.e., having cellular descriptors from cells of differing genetic backgrounds (tumor, tissue specific, and gene knock out cell lines), can provide the capability to compare cells of unknown genetic background to those in the database. Similarly, the descriptor of an unknown cellular portion in the presence of multiple drugs can be queried against the descriptors of the known markers in the database. For example, if an unknown gene is tagged with Green Fluorescent Protein (GFP), the database may be used to identify the cellular portions for which that unknown gene encodes.

According to the present invention, target validation and specialized cell-based assay screening can be performed using database systems and methods to serve as a universal high-throughput cell-based assay that can evaluate the molecular mechanism of drug action. As new genes are isolated and identified, a large collection of available gene-based knowledge is becoming available. From this large collection of new genes, potential protein targets can be identified using the genomic tools of sequence analysis and expression profiling. However, unless a gene mutation is tightly linked to a disease state, further validation of individual targets is a time consuming process, becoming a bottleneck in drug discovery. Furthermore, robotics and miniaturization are making "High Throughput Screening (HTS)" the industry standard, substantially reducing the time and cost of running a target-based biochemical assay. Therefore, it is now possible to routinely screen large libraries and use a resulting "hit" to validate the target. In such approaches, a specialized cell-based assay would be developed to test hits for each target. Since this often involves

the creation of cell lines expressing new markers, this stage may also become a bottleneck that cannot keep pace with HTS. In addition, these cell-based assays may not be amenable to high-throughput screening, making it difficult to test the increasing number of analogs arising from combinatorial chemistry.

5 In a particular embodiment according to the invention, a rapid characterization of large compound libraries for potential use as pharmaceutical products can be provided by predicting properties of compounds that relate to the compounds' potential as bioactive drugs. In many drug discovery situations, virtually millions of compounds can be passed through a HTS assay against a small number of
10 validated targets. These assays produce hundreds to thousands of potential hits. These hits can then be subsequently screened by a pipeline of secondary and tertiary screens to further characterize their specificity, often time completely missing non-specific interactions with other proteins. Techniques according to the present invention can provide a replacement to such screening operations by providing
15 information about cellular accessibility and mechanism of action for the hits coming from a HTS system. Furthermore, it can replace the biochemical HTS assay and allow rapid and accurate identification of attractive compounds from large libraries without an intervening biochemical assay. The cell information can be predictive of whether to continue into an animal model for each compound, and which animal model to
20 pursue.

 The principles of the present specifically contemplate a wide variety of research methodologies, or usage scenarios, implementing these principles. The following discussion of three such scenarios is by way of illustration and not limitation. Study of the principles enumerated herein will render evident to those
25 skilled in the art certain additional methodologies or usage scenarios enabled by the teachings hereof. The present invention specifically contemplates all such modifications. The following description presents some specific embodiments and scenarios that represent a broader use of cellular phenotypic data and characterizations to deduce mechanisms of action and other features of cellular
30 responses to various stimuli. Such procedures generally involve producing a quantitative cellular phenotype based upon two or more cellular attributes and then comparing that phenotype to phenotypes previously stored and indexed. Such

procedures make use of databases or other repositories of biological information. The invention is not limited to the specific embodiments described here.

Considering first the procedure 2000 depicted in Figure 20, a compound has been identified as having a particular cellular activity. See 2004. For example, a compound may be found to inhibit the growth of certain cancer cell *in vitro* by a specific and desired mechanism of action. This may be a particular company's "gold standard."

Next, the compound is analyzed at 2006 in terms of its effect on one or more cell lines. More specifically, the compound is linked, virtually, to a particular phenotype. Two or more values or measures of cellular attributes characterize that phenotype. These attributes are quantified in the context of specific cellular markers.

In one example, the cellular marker is an organelle such as a nucleus or Golgi apparatus. Measured attributes useful for characterizing an associated phenotype include geometric parameters (e.g., size, shape, and/or location of the organelle) and composition (e.g., concentration of particular biomolecules within the organelle).

The phenotype may be characterized by administering the compound of interest to various cell lines and in various concentrations. In each example within this matrix, the attributes of interest are measured. Ultimately, certain phenotypic features (combinations of attribute values) are associated with the compound of interest. These features provide a template for the phenotype.

Next, using the phenotype as identified at 2006, the process identifies other compounds providing similar features. The goal here is to present a list of compounds having a mechanism of action similar to that of the compound that started the process. This allows researchers to identify a mechanism of action, if not already known, for their compound and to draw conclusions based upon their compound's link to other known compounds (which may not be chemically/structurally similar to the compound of interest).

Identifying similar compounds based upon phenotype can take many paths. Most will involve some mathematical basis. For example, the phenotype defined at 2006 can be represented as a fingerprint or vector comprised of multiple scalar values of cellular attributes (as described above). The phenotype representation can then be compared against known phenotypes characterized by the same format

(e.g., they are all characterized as vectors having the same attribute set, but with different values of the attributes). The comparison may be as simple as a Euclidean distance or more sophisticated as a neural network or multivariate statistical correlation.

5 The known compounds and associated phenotypes may be stored as database records or other data structures that can be queried or otherwise accessed as part of the identification procedure. The compounds may also be associated with other relevant data such as clinical toxicity, cellular toxicity, hypersensitivity, mechanism of action, etc. (when available).

10 Compounds found to be sufficiently similar to the starting compound are returned for consideration by researchers. A data processing system may rank such compounds based on degree of similarity to the starting compound. In some cases, the system may even provide similarity scores associated with the listed compounds.

15 Often researchers wish to determine whether their particular compound has clinical or biochemical effects beyond those that they are already aware of. In a typical scenario, the compound of interest was selected based upon its strong binding a target or its stimulation or inhibition of cell growth in a particular cell line. The process associated with 2010 has likely identified the compound of interest as having
20 a particular mechanism of action based on phenotypic similarity to other compounds having a similar mechanism of action. However, within the region of biochemical space, there may be subspaces (characterized by subphenotypes) that correspond to separate properties. For example, within the phenotypic space associated with one mechanism of action, there may be subspaces associated with clinical toxicity,
25 cellular toxicity (likely overlapping the clinical toxicity space), and little or no toxicity. Obviously, a researcher would like to know whether her compound is likely to be toxic.

 Thus, the process 2000 may include characterizing the compound of interest in terms of its distance from (i.e., similarity to) specific phenotypes having
30 known characteristics. In a typical example, the known characteristic is toxicity. This feature allows the researcher to quantify her compound in terms of mechanism of action AND toxicity (or in terms of two or more other relevant properties associated

with phenotype). To allow simple ranking or characterization, compounds of interest may be scored according to a simple or weighted Boolean expression.

A second scenario of interest is depicted in Figure 21. This scenario again defines a phenotype in terms of a quantifiable vector or other measure.

5 However, rather than using a compound of interest to generate the phenotype, some other cellular stimulus is used to generate the phenotype.

As shown, a process 2100 begins with receipt of cells of interest. See 2104. In many situations, the cells are produced by a genetic or epigenetic process that affects the expression level or activity of a particular protein. More generally, 10 any cellular stimulus (e.g., radiation level and type, gravity level, magnetic field, acoustic perturbations, etc.) can be used to generate the cell line of interest. Importantly, this stimulus affects the phenotype and can be correlated therewith.

In the context of drug discovery, a gene encoding for a particular target can be genetically knocked out, underexpressed, overexpressed, expressed in a non- 15 native state, etc. This may be accomplished via standard procedures involving genomic modification, translation or transcription apparatus modification (e.g., use of antisense nucleic acids), blocking target activity (using antibodies to a receptor site for example), and the like. These processes will generally affect the phenotype in some quantifiable way. Importantly, they clearly and unambiguously define a cellular 20 phenotype associated with altering the activity of the target protein.

At 2106, the process involves measuring one or more cellular features from the cell line of interest to define/quantify the phenotype. This may be accomplished as described above with reference to 2006. Next, at 2108, the cellular phenotype generated in this manner is used to identify and rank a set of compounds 25 associated with the phenotype. This operation may proceed in the manner of operations 2008 and/or 2010 from Figure 20.

Finally, at 2110, the process clusters the compounds returned at 2108 by a mechanism of action. The operation 2106 has tightly bound a mechanism of action to a phenotype. Various compounds characterized and stored in a system 30 database may be tentatively assigned a mechanism of action or may have no suggested mechanism of action. By matching their virtual phenotype to the phenotype generated at 2106, one can create or strengthen an association between the compounds and mechanism of action relevant to the stimulus at 2104.

Considering now Figure 22, a third scenario is depicted. This scenario again involves using a virtual phenotype to glean information relevant to a mechanism of action or other cellular activity. In this case, assay data from a group of compounds (e.g., a primary or focused library) is used to elucidate a phenotype.

5 As shown, a process 2200 begins by identifying a target protein. See 2204. Then, at 2206, the process involves identifying positive and negative biochemical hits. More generally, this may involve ranking a number of compounds based upon their interaction with the target. In a specific case, the compounds are ranked based upon their binding affinities to or ability to inhibit the enzymatic activity
10 of the target protein.

 After the compounds have been characterized in some manner based upon their interaction with the target, they are used to define a cellular phenotype. See 2208. Generally, the techniques to accomplish are the same as described with reference to operation 2006 of Figure 20. In this case however, one may obtain a
15 strong correlation between mechanism of action (involving the target) and phenotype by using multiple of the compounds identified at 2206. For example, some of the "best hits" may be administered to cell lines in various concentrations. And some of the least effective compounds may also be administered. Cellular attributes that are more strongly exhibited with increasing concentration of the best hits (and not
20 exhibited or exhibited only weakly upon administration of the negative hits) can be used to define the virtual phenotype. In a related approach, compounds having widely varying levels interaction with the target are administered to cells. Those cellular attributes that vary linearly or at least monotonically with the degree of interaction between the target and compound represent attributes that can be used to define the
25 virtual phenotype.

 After the cellular phenotype has been defined, previously characterized compounds may be clustered with that phenotype. See 2210. As with operation 2110 of Figure 2, this may create or strengthen an association between a mechanism of action and various compounds in a database.

30 Finally, and optionally, procedure 2200 may provide a "higher resolution" mechanism of action for the compounds identified at 2206. See 2212. Presumably interaction with the target suggests a specific mechanism of action or at least some aspect of a mechanism of action. However, a given target may participate

in a larger cellular mechanism of action – unknown to researchers. Further, a compound may that binds with the target may participate in multiple mechanisms of action – some of which do not involve the target. By linking the target (and its positive hits) to a particular phenotype, some of these additional cellular level activities can be elucidated. The defined phenotype may have been previously identified as associated with other mechanisms of action or higher resolution mechanisms of action. Thus, the phenotype identified at 2208 can be leveraged to generate a higher resolution mechanism of action at 2212.

As suggested in the above discussion, compounds and associated phenotypes may be stored as database records. Such databases can take on many flavors. In one example, a database includes various pieces of information relevant to oncology. Such database may include numerous compounds classified by cellular phenotype, mechanism of action, toxicity, etc. More specifically, the database may include data on commercially available compounds clustered by cellular phenotypes corresponding to mechanisms of action. Further the databases of interest may extended or combined (via standard relational tables and algebra for example) to include additional data such as pharmacology data, cellular genomics data, gene expression data, protein expression data, etc. In a specific example, the database includes measurements made on a subset of the NCI60 cell lines, using DNA, Golgi apparatus, and/or microtubules as markers for defining the phenotypes. Other data includes dosage response information, variation in effect over time, etc. The compounds populating the database could include known National Cancer Institute oncology study compounds. In a specific embodiment, the compound set includes some or all of the compounds mentioned in the article “A gene expression database for the molecular pharmacology of cancer,” Nature Genetics, 24, pp. 236-244 (March 2000).

Various biological analyses may be conducted to develop additional information for characterizing compound mechanisms of action, etc. For example, a cell count analysis may be used to develop dose response curves, GI 50 data, etc. The cell cycle may also be analyzed to find out how various stages in the cycle vary in response to particular stimuli. The Golgi apparatus may be analyzed to determine whether it is in a normal state, a dispersed state, a diffused state, etc. As another example, tubulin may be analyzed to determine whether it is normal, de-polymerized,

over-polymerized, bundled, etc. Obviously, combinations of such analyses may be performed. For example, properties of the Golgi apparatus or tubulin may be analyzed over one or more cell cycles.

In some embodiments, techniques according to the present invention
5 can provide tools for the later stages of drug development such as clinical trial design and patient management. The properties of known drugs, such as clinical trial and patient response information, will be used in a similar fashion as the pre-clinical information to provide predictions about the properties of novel compounds. Because
10 the human cell is the locus of drug action, a database containing drug-cell interactions will be able to provide predictive value for this aspect of drug development.

Although the above has generally been described in terms of specific hardware, software, and methods, it is understood that many alternatives can exist. In particular, the present invention is not limited to a particular kind of data about a cell, but can be applied to virtually any cellular data where an understanding about the
15 workings of the cell is desired. Thus, in some embodiments, the techniques of the present invention could provide information about many different types or groups of cells, substances, and genetic processes of all kinds. Of course, one of ordinary skill in the art would recognize other variations, modifications, and alternatives. Some examples according to the present invention are provided below.

20

EXPERIMENTS

To prove the principle and demonstrate the objects of the present invention, experiments have been performed to determine the effects of manipulations on cell structure using imaging and analysis techniques applied to a variety of
25 situations. These experiments were performed by growing multiple cell lines in the presence of multiple compounds, or substances. Cells were fixed and stained with fluorescent antibodies or labels to multiple cellular portions. One or more images of the cells were then obtained using a digital camera. Descriptors were built by quantifying and/or qualifying patterns of one or more feature from each image in the
30 cell lines under study. A database was built from the descriptors. As the database grows, it should be able to predict the mechanism of action of an unknown drug by comparing its effect with the effects of known compounds or to identify data clusters within large libraries of compounds.

In a first experiment, an automated method to count the number of cells and differentiate normal, mitotic, and apoptotic cells was created.

Approximately, 5,000 HeLa cells were plated per well in a 96 well plate and grown for 3.5 days. The cells were fixed with -20° MEOH for 5 minutes, washed with TBS for 15 minutes, and then incubated in 5 mg/ml Hoechst 33342 in TBS for 15 minutes. Then, 72 images were collected with a 40x objective and 75 ms exposure time.

The analysis was performed on objects that met a certain size criteria that was based on 1) measuring the size of objects in the image that were clearly not cells and 2) excluding the first peak of the area histogram (Fig. 8B values 1-4654).

Histograms of the individual object data were generated for each type of feature. Fig. 8A shows the histogram for average intensity, and Fig. 8B shows histogram data for the area of each object. Fig. 8C shows the scatter plot of the average intensity vs. the area of all of the objects. The pattern of the scatter plot showed an interesting pattern: a large cluster of cells in one region of the graph, with a scattering of object points in other regions. Because mitotic structures are identified as particularly bright objects, most likely due to the biological fact that the chromatin is condensed, the original Hoechst images could be used to identify which cells were either undergoing mitosis, or otherwise looked abnormal. Manual inspection of 917 cells resulted in the classification of each object. Fig. 8D shows a graph where each type of cellular classification is delimited. This graph clearly shows that the mitotic nuclei are brighter than the interphase nuclei. Further, the different phases of the cell cycle can be separated using these two features. Figs. 8E-8F show bar graphs of the average and standard deviations of the areas and average intensities for each cell classification type. These graphs show that interphase nuclei are statistically less bright than mitotic nuclei and that telophase nuclei are statistically smaller than other mitotic nuclei.

Each image was thresholded to an intensity level of 20. A standard area value was set at 9500 pixels. Automated information gathering about all of the objects was done and collected into an Excel spreadsheet (for more information see, section on imaging system). The following information was recorded:

IMAGE NAME
OBJECT #

AREA
STANDARD AREA COUNT
PERIMETER
FIBER LENGTH
FIBER BREADTH
SHAPE FACTOR
ELL. FORM FACTOR
INNER RADIUS
OUTER RADIUS
MEAN RADIUS
AVERAGE INTENSITY
TOTAL INTENSITY
OPTICAL DENSITY
RADIAL DISPERSION
TEXTURE DIFFERENCE MOMENT
EFA HARMONIC 2, SEMI-MAJOR AXIS
EFA HARMONIC 2, SEMI-MINOR AXIS
EFA HARMONIC 2, SEMI-MAJOR AXIS
ANGLE
EFA HARMONIC 2, ELLIPSE AREA
EFA HARMONIC 2, AXIAL RATIO
EFA HARMONIC 3, SEMI-MINOR AXIS

The following results were obtained:

- 1,250 objects were counted
- 201 of those objects has standard area counts > 2 (area > 19000 pixels)
- 195 objects had areas < 6000 pixels
- 1529 objects estimated in total
- 1328 object areas are > 6000 pixels
- The data was reduced to 917 objects that were $6000 < \text{area} < 19000$
- For the 917 objects a scatter plot of area vs. average intensity and a histogram of the average intensity were generated.

- 116 objects that had average intensity intensities > 60 were manually looked at to determine their morphology.

- Of those 116 objects:

6 were dead or indistinguishable

5 4 were interphase

30 were prophase

32 were metaphase

24 were anaphase

20 were telophase (10 pairs)

10

- 12 prophase objects were missed because of gray scale cut off. (8 of those prophase cells had gray scale values > 57 , as did 7 interphase)

- 1 telophase object was missed because it was too small (< 6000)

- 1 prophase object was missed because it was too big (> 1900)

15

- 16 mitotic objects were missed because they were parts of objects with standard count > 2 .

In sum, out of 917 single objects, the analysis correctly identified 106 out of 130 mitotic objects, or (81% predictive, 91% of identified mitotics). Out of 917 single objects, the analysis incorrectly identified only 10 non-mitotics as mitotics (1% total, 8% of identified mitotics); 14 mitotics as interphase (1.4% total, 1% interphase). An automated classification system that would automatically assign values to each object using these or other measurement features can thus be developed, utilizing the principles set forth herein.

In a second experiment, the effects of Taxol on MDCK cells and the different types of morphological effects were observed. A plurality of MDCK cells grown in 96 well plates were treated with Taxol for 4.5 hours at different concentrations (10 uM-1pM). They were then fixed, labeled with Hoechst, and imaged.

This experiment used a labeling protocol comprising: MEOH fix at – 20°, Wash in PBS, Block in PBS/BSA/Serum/Triton-X 100, Incubate with 5 µg/ml Hoechst 10 minutes, and wash.

Cells were inspected for different morphologies and manually counted at each different drug concentration in one well. Fig. 9 shows example images from each drug concentration and the different types of morphologies and cells are highlighted. Fig. 10 shows the distribution of each morphology within the cell population as a function of drug concentration. The higher the concentration of Taxol, the larger proportion of cells underwent apoptosis, and the fewer number of normal mitotic cells were detected.

In a third experiment, the purpose was to determine whether the automated analysis methods developed in the first experiment can detect differences in Hoechst morphology in the presence of 6 known compounds at one concentration and exposure time in one cell line. In this experiment, HeLa cells were separately treated with 6 compounds with known mechanism of action. The quantitative methods described in the first experiment were applied to the Hoechst images.

Approximately 5,000 HeLa cells per well were plated in a Costar black-walled 96 well tissue culture treated plate and left to recover in the incubator for 24 hours. After this time, 10 ug/mL of cytochalasin D (CD), Taxol, hydroxyurea, vinblastine, nocodazole, and staurosporine was added to different wells at a 1:100 addition in DMSO.

The cells were incubated in the presence of drug for 24 more hours. After 24 hours, the cells were removed and fixed as in the first experiment. Then, 9 images per well were collected of the Hoechst staining using a 10x objective.

5 The low magnification images taken of Hoechst were run through the automated image analysis method described in the first experiment. Plots of the average intensity and area were made of each compound. Fig. 11 shows the scatter plots of the compounds. The scatter plots of each compound are visually distinct. For example, cells treated with CD are smaller than control, and cells treated with Hydroxyurea are larger and brighter. Furthermore, the number of cells per well was
10 very different (data not shown).

The effects of different compounds can be clearly and automatically distinguished by identifying changes in cellular morphology. This method can also be used to count adherent cells.

The next experiment was to develop clustering algorithms that assign
15 statistically meaningful values to the representative two dimensional data shown in Fig. 10, and even more complicated clustering of all of the multidimensional data that can be extracted across one, and multiple images.

A fourth experiment was performed to obtain high magnification images of two markers in the presence of drugs. In this experiment, HeLa cells were
20 treated with 80 generic compounds with known mechanism of action. The quantitative methods described in the first experiment were applied to the Hoechst images.

Approximately 5,000 HeLa cells per well were plated in a Costar black walled 96 well tissue culture-treated plate and left to recover in the incubator for 24
25 hours. After this time, 10 ug/mL of each compound from the Killer Plate from Microsource Discovery Systems (Gaylordsville, CT) was added to different wells at a 1:100 addition in DMSO. The cells were incubated in the presence of drug for 24 more hours. After 24 hours, the cells were removed and fixed as in the first experiment. In addition to being labeled with Hoechst 33342 (against chromatin),
30 cells were also labeled with 1 unit of rhodamine-conjugated phalloidin (against actin) for 30 minutes.

The 96 well plate was imaged twice. Once, 9 images per well were collected of the Hoechst staining using a 10x objective. After this, one image per well of both the phalloidin and Hoechst staining was collected using a 40x objective.

The resulting high magnification images were analyzed qualitatively and distinct pattern differences were detected in both the Hoechst and phalloidin images. Fig. 12 shows three example images from the experiment. The top row is the Hoechst staining, and the bottom row is the phalloidin staining from the same well. The columns show the images from wells treated with just DMSO (control), cytochalasin D, and Colchicine. The morphology of each marker is different in the presence of each drug. Interestingly, there is an effect in the morphology of the chromatin in the Hoechst image of cytochalasin D, which directly targets the actin cytoskeleton (and thus there is an expected effect in the phalloidin image). Also, there is an effect on the actin cytoskeleton, compared to control, in the presence of colchicine that directly targets the microtubule network.

The low magnification images were analyzed as described in the first experiment, and different patterns were seen in both the average intensity vs. area plots, and in the number of cells per well (data not shown). Thus, changes in patterns of a marker that is "down-stream" from the direct target of a compound are detectable. Automated image analysis protocols for actin and other markers can be developed similarly, again utilizing the principles set forth herein.

A fifth experiment was performed to test quadruple labeling of 9 different cell lines grown in normal conditions. In this experiment, NCI-H460, A549, MDA-MD-231, MCF-7, SK-OV-3, OVCAR-3, A498, U-2 OS, and HeLa cells were plated. Then, the cells were fixed and stained for portions of the each cell known as DNA, tubulin, actin, and Golgi.

The following table summarizes the procedures for this experiment:

Action	Active Ingredient/Notes	Buffer	Vol/ well	Desired Time	Temp
Remove media	NOTE: gently by pipetting, not aspiration				
Fix	4% Formaldehyde	PBS	100µl	20 min	rt
Wash		TBS	100µl	5 min	rt
Wash		TBS	100µl	5 min	rt

Permeablize	0.1% Triton X-100	TBS	100 μ l	10 min	rt
Permeablize	0.1% Triton X-100	TBS	100 μ l	10 min	rt
Block	% BSA % Serum Filter sterilize before use	TBS w/azide	100 μ l	1 hr or o/n	rt or 4°C
Primary Antibody	1:1000 dilution of DM1 α	TBS + 1% BSA + 0.1% TX-100	50 μ l	1 hr -- or o/n	rt or 4°C
Wash		TBS	100 μ l	5 min	rt
Wash		TBS	100 μ l	5 min	rt
Wash		TBS	100 μ l	5 min	rt
Fluorescent Stain	FITC lens culinaris 1:500 Rhodamine-Phalloidin 1:500 CY5 goat anti-mouse 1:100	TBS + 1% BSA + 0.1% TX-100	50 μ l	1 hr.	rt, dark
Wash		PBS	100 μ l	5 min	rt, dark
Hoechst	1:1000 dilution of 5mg/ml	TBS	100 μ l	15 min	rt, dark
Wash		PBS	100 μ l	5 min	rt, dark
Wash		PBS	100 μ l	5 min	rt, dark
Wash		PBS	100 μ l	5 min	rt, dark
Store		PBS	200 μ l	1 month	4°C

Cells were plated out at different densities for 48 hours. Cells were fixed and labeled by the above method. Cells were imaged using an automated imaging system that collected 9 images from each marker using a 10x objective.

Higher magnification images were collected of a few cells for demonstration purposes.

In this experiment, each cell line demonstrated different morphological patterns as determined by phase. For example, A549 cells are much more compacted than OVCAR-3 cells as determined by phase contrast imaging (data not shown). The different fluorescent markers showed even bigger differences between different cell lines. Figs. 13 and 14 show 4 panels of each marker for A549 (Fig. 13) and OVCAR-3 cells (Fig. 14). The markers are Hoechst (upper left), Phalloidin (upper right), Lens culinaris (lower left), and DM1a antibody (lower right). The following table summarizes the qualitative differences between these images:

MARKER	A549	OVCAR3
Hoechst DNA	small	large
Phalloidin actin	fuzzy	crisp - many stress fibers
Lens culinaris Golgi	compact	Disperse/punctate
DM1alpha Tubulin	perinuclear	evenly distributed

Higher magnification images were taken of the OVCAR3 cells. Fig. 15 shows the same markers at 20x, and Fig. 16 shows the markers at 40x. While the highest magnification images show the most detail, these images illustrate that very little morphological or feature information is lost in the 10x images.

These data exemplify the differences in morphology seen between different cell types. Thus the automated image analysis software can be customized for each marker in each cell type. Different drugs should effect these morphologies differentially.

An automated quantification method for each marker and cell line can be similarly developed.

A sixth experiment was conducted with a more sophisticated software package and to develop more flexible image recognition algorithms. In this experiment, prototype image features extraction was performed using MatLab programming language with image toolbox and SDC morphology toolboxes. Algorithms are being developed that will automatically identify objects on images and

to measure various morphological and feature parameters of these objects. Many different features for each of the cellular markers were acquired.

An example of a MatLab program called "AnalyseDNA" that takes as an input an unlimited number of images, identifies individual objects in these images based on either their intensities, or based on edge-detection algorithms, and extracts a number of morphological and intensity characteristics of these objects. A copy of this program follows:

Listing of the AnalyseDNA.m program and of some of the
supporting subroutines

```
10 function files_analysed = AnalyseDNA(filemask, outpath,
    nx, ny, filter_range, dext, modifier, sfname)
% AnalyseDNA performs measurements on files of DNA images
% V1. EV 2-11-99; 2-15-99; 2-16-99
15 %
% files_analysed = AnalyseDNA(filemask, outpath, nx, ny,
    filter_range, dext, modifier, sfname)
%
% PARAMETERS:
20 %   ALL PARAMETERS ARE OPTIONAL
%
%   FILEMASK - mask for file names to be analyzed
    INCLUDING PATH(for example c:\images\*.tif)
%   DEFAULT '.\*.tif' (all *.tif files in the current
25 directory).
%
%   OUTPATH - path to a directory where all the output
    files will be placed.
%   DEFAULT - output is saved in the same directory
30 which contains images
%
%   NX, NY - number of individual images in montage
    images along X and Y axes (DEFAULT 1)
%
```

```
%    FILTER_RANGE - 3 col-wide array (or[]). Specifies
%    how data is filtered when summary is calculated
%    this parameter internally is passed to GetDNADData
%    and then to GetSummaryData - see these
5 %    functions for details. For example: [2 2 Inf; 6 100
%    8000] will case all raws of data for which
%    values in column 2 are less than 2 and all raws
%    where values in column 6 are less than 100 or
%    more than 8000 to be excluded from all
10 calculations of a summary.
%    DEFAULT - [] (means do not filter, summarize all
%    data)
%
%    DEXT - string. Extension for data files being saved.
15 %    DEFAULT 'dat';
%
%    MODIFIER - this modifier is 'SUMMARY', summary file
%    is created;
%    'SUMMARY ONLY' - only summary is generated,
20 data for individual files are not saved
%
%    sfname - string. File name of a summary file
%    DEFAULT 'summary[date].dat'
%
25 % OUTPUT:
%
%    AnalysedDNA works on image files or montages. For
%    each image file it creates a tab-delimits file of
%    measured
30 %    parameters of all the objects in the montage with
%    the same base name as a montage file and extension
%    specified
```

```
%    by dext parameter (or .dat by default) and file
'errors[date].err' - with the list of files that matched
the
%    filemask but could not be processed.
5 %    If 'summary' or 'summary only' modifier is
specified, it also creates a single file
'summary[date].dat' (or
%    different extension, if specified by DEXT) which
contains summary information for all analyzed files.
10 %
%    ALL OUTPUT FILES are saved in a directory specified
by OUTPATH parameter
%
%    RETURNS *files_analysed* - number of files that have
15 been successfully processed.
%
%    Column designations in the output files are
described in GetDNADData
%
20 % FILE NAME CONVENTIONS
%    AnalyseDNA attempts to identify a number for each
file to identify the file in summary output.
%    It does that by looking for the first space or
underscore, followed by a number and then takes
25 %    as many successive numbers as it can find. If it
fails to identify a number it assigns a
%    default which is -1
%
%
30 % SEE ALSO GetDNADData, GetSummaryData
%
% TO DO    improve error handling in opening and writing
files (GLOBAL error_file ?)
```

```
%          include procedures for writing text headers
into the output files

if nargin > 8
5   error ('Wrong number of input parameters');
end
if nargin >1
    error ('Wrong number of output parameters: only one
allowed');
10  end

% set defaults
need_summary = 0;
summary_only = 0;
15  use_default_outpath = 0;
datestring = datestr(floor(now));
if nargin == 7      % set default summary file name
    sfname = ['summary' deblank(datestring)]; % extension
will be appended later based on dext
20  if deblank(upper(modifier)) == 'SUMMARY'
        need_summary = 1;
    elseif deblank(upper(modifier)) == 'SUMMARY ONLY'
        need_summary = 1;
        summary_only = 1;
25  else
        error (['Wrong parameter: unknown modifier '
modifier]);
    end
end
30

if nargin == 5
    % default data file extension
    set_dext = 'dat';
end
```

```
    if nargin == 4
        % default filter range
        filter_range = [];
    end
5   if nargin == 3
        ny = 1; % default number of images in montage along Y
    end
    if nargin == 2
        nx = 1;
10  end
    if nargin == 1
        use_default_outpath = 1;
    end
    if nargin == 0
15  filemask = '*.tif'
    end

    % check parameters
    if ( ~ischar(filemask) | ~ischar(dext) | ~ischar(sfname)
20  )
        error('Wrong parameter type: filename, filepath,
dext and sfname should be strings');
    end
    if ( ( size(nx) ~= [1 1] ) | ( size(ny) ~= [1 1] ) )
25  error ('Wrong parameter type: nx and ny should be
scalars (1x1 arrays)');
    end
    if (~isempty(filter_range) & size(filter_range, 2) ~= 3)
        error ('Wrong parameter type: filter range should be
30  [] or 3 - cols-wide array');
    end
    % end testing parameters

    % Generate list of files to process
```

```
datapath = getpath(filemask);
if use_default_outpath == 1
    outpath = datapath;
5  end
if exist(outpath, 'dir') ~= 7
    error(['Path ' outpath, 'not found. Exiting..']);
elseif exist(datapath, 'dir') ~= 7
    error(['Path ' datapath, 'not found. Exiting..']);
10 end

sfname = makefullname(outpath, sfname, dext);
if need_summary == 1
    if exist(sfname, 'file')
15         disp(['File ', sfname, 'already exists!']);
        input ('Press ^C to abort, Enter to delete and
continue');
        delete(sfname);
    end
20 end

flist = FileList(getfname(filemask), datapath);
numfiles = size(flist, 1); % total number of files to
25 process
disp(['About to process ', num2str(numfiles), ' files']);
%DEBUG - commented out "input" to run from Wrod
input('Press ^C to abort, Enter to continue');

30 % main loop where the job gets done:
error_file = makefullname(outpath, ['error' datestring
'.err']);
num_processed = 0;
num_error = 0;
```

```
for i = 1:numfiles
    % first generate file name for a data output file
    current_fullname = flist(i, :); % full name with path
    and extension
5    current_datafile = makefullname(outpath,
    makefname(getbasefname(current_fullname), dext) );

    %extract number from a filename
    fnumber = getfilenumber(current_fullname);
10
    % load an imagefile, record errors
    read_error = 0;
    try
        I = imread(current_fullname);
15        %DEBUG
        disp(['Image file #', num2str(fnumber), '
loaded']);
    catch
        % record file-opening error in an error_file
20        read_error = 1;
        num_error = num_error +1;
        msg = [current_fullname ': ' lasterr];
        add_error_msg(error_file, msg);
    end
25
    % extract and write data to a file in outpath
    if read_error ~=1
        if (need_summary == 0)
            %DEBUG
30            disp(['Starting analysis of file #',
num2str(fnumber), '.']);
            current_data = GetDNADData(I, nx, ny, fnumber);
            %DEBUG
```

```

        disp(['Finished analysis of file #',
num2str(fnumber), '.']);
        %load current_data.mat 'current_data';
        write_data(current_data, current_datafile);
5      else      %summary needed
        %DEBUG
        [current_data, current_summary] = GetDNADData(I,
nx, ny, fnumber, filter_range);
        %load current_data.mat 'current_data';
10      %load current_summary.mat 'current_summary';
        write_summary (current_summary, sfname);
        if summary_only ~= 1
            write_data(current_data, current_datafile);
        end
15      end
    end
end % of the main for loop
num_processed = numfiles - num_error;

20 %=====end function AnalyseDNA()
=====

%=====
=====

25 function result = add_error_msg(filename, msg)
% adds string MSG to an errorfile FILENAME
% returns 1 if success, 0 if failure

err_FID = fopen(filename, 'at');
30 if err_FID == -1
    warning(['Can not open error file ' filename]);
else
    fprintf(err_FID, '%s\n', msg);
    fclose(err_FID);

```



```

end
%=====end function add_error_masg()
=====

5 %=====
=====

function N = getfilenumber(fname)
% returns the first number extracted from a file name
(string) or -1 if fails to extract any number
10 numbers = NumbersFromString( getfname(fname) ); % vector
of all numbers encoded in the name

                                % (but not in the path, even if
present)
15 if isempty(numbers)
    N = (-1); % return -1 if no numbers found in the
name
else
    N = numbers(1);
20 end

%===== end function getfilenumber()
=====

25 %=====
=====

function result = write_data(data_array, file_name)
% writes data in a data_array in a tab-delimited ascii
file.
30 % result is 0 if success and -1 if failure
% if file_name exists, overwrites it
result = -1;
try
    fid = fopen(file_name, 'wt');

```

```

        if fid ~= -1
            for k = 1:size(data_array, 1)
                fprintf(fid, '%g\t', data_array(k, :));
                fprintf (fid, '\n');
5         end
            test = fclose(fid);
            result = -1;
        catch
            result = -1;
10    end

%===== end function write_data()
=====

15  %=====
=====

function result = write_summary (s_vector, file_name)
% appends summary vector s_vector to a file_name (ASCII
tab-delimited file).
20  % if file_name does not exist, creates it.
% result is 0 if success and -1 if failure
%
result = -1;
try
25    % debug
        fid = fopen(file_name, 'at');
        result = fprintf(fid, '%g\t', s_vector);
        result = fprintf(fid, '\n');
        result = fclose(fid);
30    result = 0;
    catch
        result = -1;
    end

```

```

% ===== end function write_summary()
=====

function Data = GetObjectsData(I, Ilabel)
5 % GetObjectsData returns array measurements of objects in
  image "I" masked by "Ilabel"
  % EV 2-3-99; 2-10-99
  % OData = GetObjectsData(I, Ilabel) returns an array of
  morphological and intensity measurements
10 %   taken from a grayscale image "I". Objects are
  identified on a mask image Ilabel, usually
  %   created by bwlablel()
  % OUTPUT:
  % Each row in the output array OData represents
15 individual object
  % columns contain the following measurements:
  %
  %   1 - Index ("number" of an object);      8 -
  Solidity;
20 %   2 - X coordinate of the center of mass; 9 - Extent;
  %   3 - Y coordinate      "-"; 10 - Total
  Intensity;
  %   4 - Total Area (in pixels);              11 - Avg.
  Intensity;
25 %   5 - Ratio of MajorAxis/MinorAxis;        12 - Median
  Intensity;
  %   6 - Eccentricity;                        13 - Intensity of
  20% bright pixel
  %   7 - EquivDiameter;                       14 - Intensity of
30 80% bright pixel
  %
  % For details on morphological parameters see information
  on MatLab imfeature();

```

WO 00/70528

```

% Intensity parameters are either obvious or are
documented in comments in this file.
% Procedures in this file are documented in notebook file
"MATLAB Measuring Nuclei (1) 1-29-98.doc"

5
if (nargin ~= 2)
    error ('function requires exactly 2 parameters');
end
if (nargout ~= 1)
10    error ('function has 1 output argument (array X by
    14)');
end

% finished checking arguments

15
% first collect morphological parameters in a structure
array:
ImStats = imfeature(Ilabel, 'Area', 'Centroid',
'MajorAxisLength',...
20    'MinorAxisLength', 'Eccentricity', 'EquivDiameter',
    ...
    'Solidity', 'Extent', 8 );

% now convert it into array (matrix) while collecting
25 intensity data for each object:

%preallocate output array:
numobjects = size(ImStats, 1);
OData = zeros(numobjects, 14);
30 %now convert ImStats into array and add intensity data to
    it
    for k=1:numobjects
        OData(k, 1) = k;
        OData(k, 2) = ImStats(k).Centroid(1);

```

```

        OData(k, 3) = ImStats(k).Centroid(2);
        OData(k, 4) = ImStats(k).Area;
        OData(k, 5) = (ImStats(k).MajorAxisLength) /
        (ImStats(k).MinorAxisLength);
5         OData(k, 6) = ImStats(k).Eccentricity ;
        OData(k, 7) = ImStats(k).EquivDiameter;
        OData(k, 8) = ImStats(k).Solidity;
        OData(k, 9) = ImStats(k).Extent;

10         % now collect and assign intensity parameters from
        image I

        object_pixels = find( Ilabel == k);
        object_area = size(object_pixels, 1); %same as total
15 number of pixels in the object
        object_intensities = double(I(object_pixels)); %
        need to convert to double to do math
        sorted_intensities = sort(object_intensities); %
        will need to get median, 20% and 80% pixels
20         total_intensity = sum(object_intensities, 1);
        avg_intensity = total_intensity / object_area;
        median_intensity = sorted_intensities( floor(
        object_area/2 ) + 1 );
        pix20 = sorted_intensities( floor(object_area*0.2)+1
25 ) ; %brightest pixel among dimmest 20%
        pix80 = sorted_intensities( floor(object_area*0.8)+1
        ) ;

        OData(k, 10) = total_intensity;
30         OData(k, 11) = avg_intensity;
        OData(k, 12) = median_intensity;
        OData(k, 13) = pix20; %brightest pixel among dimmest
        20%

```

```

        OData(k, 14) = pix80; %dimmeest pixel among brightest
    20%
    end %for

5  %===== end function
    GetObjectsData()=====

function Imask = MaskDNA1(I);
10 % MaskDNA1 - generates binary mask for cell nuclei
    through edge detection
    % EV 1-22-99; 2-6-99; 2-10-99
    % Imask = MaskDNA1(I)
    % PARAMETERS
15 %     I - intensity image (grayscale)
    % OUTPUT
    %     Imask - BW image with objects from I
    %
    % For more details see Notebook Matlab_DNA_masking1_1-22-
20 99.doc
    % Uses SDC Morphology Toolbox V0.7

    if (nargin ~= 1)
        error('Wrong number of input parameters');
25 end
    if (nargout ~= 1)
        error('Wrong number of output parameters: one output
        argument should be provided');
    end
30

    Imask = edge(I, 'canny');
    Imask = mmddil(Imask, mmsecross(1));
    Imask = mmnero ( mmclohole(Imask,mmsecross(1)));

```

```

Imask = mmedgeoff(Imask, mmsecross(1));
% note that mmedgeoff this command removed FILLED OBJECTS
but not touching OUTLINES.
% these outlines can be removed by filtering:
5  Imask = medfilt2(Imask, [5 5]);

%=====end MaskDNA1 =====

```

Given the list of image files or montages of images as an input, this
 10 program creates an individual file for each image that contains the following
 quantitative measurements for all objects identified in the image:

1 - Index ("number" of an object);	8 - Solidity;
2 - X coordinate of the center of mass;	9 - Extent;
15 3 - Y coordinate "-";	10 - Total Intensity;
4 - Total Area (in pixels);	11 - Avg. Intensity;
5 - Ratio of MajorAxis/MinorAxis;	12 - Median Intensity;
6 - Eccentricity;	13 - Intensity of 20% bright pixel
7 - EquivDiameter;	14 - Intensity of 80% bright pixel

20 A fragment of an output for a single file, containing 9 images of cells
 stained for DNA and acquired with a 10x objective. A montage image that was used
 as a source to generate data in A is presented in Fig. 17.

The same program also summarizes measurements across many files
 and performs statistical analysis of the summary data. It creates a summary file with
 25 the following data:

1 - Image file number;	
2 - Average object Area (in pixels);	3 - STD (standard deviation) of
2;	
30 4 - Avg. of Ratio of MajorAxis/MinorAxis;	5 - STD of 4;
6 - Avg. Eccentricity;	7 - STD of 6;
8 - Avg. EquivDiameter;	9 - STD of 8;
10 - Avg. of Solidity;	11 - STD of 10;

- | | |
|---|----------------|
| 12 - Avg. of Extent; | 13 - STD of 11 |
| 14 - Avg. of objects Total Intensity; | 15 - STD of 14 |
| 16 - Avg. of objects Avg Intensity; | 16 - STD of 15 |
| 18 - Avg. of objects Median intensity; | 19 - STD of 18 |
| 20 - Avg. of objects intensity of 20% bright pixel; | 21 - STD of 19 |
| 22 - Avg. of objects intensity of 80% bright pixel; | 23 - STD of 21 |

An example of summary output obtained by running AnalyseDNA against 10 montage files also is shown in Appendix B.

10 A seventh experiment was conducted in order to use sequence analysis algorithms to analyze features of cell images. In this experiment, HeLa cells were treated for 24 hours with several different compounds, and then fixed, and stained with a fluorescent DNA dye. One image of these cells was acquired for each of the treatments and morphometric parameters and features were measured:

15 Resulting measurements were arranged into a string of numbers and reduced to a pseudo- nucleic acid sequence using following rules: At any given position in the sequence a number was substituted by "t" (a code for thymidine) if its value is among highest 25% of the values at the corresponding position in the data set, "g" if it is between 50% and 25%, "c" if it is between 75% and 50%, and "a" if it
20 belongs to lowest 25% of values. Thus one descriptor or sequence was generated per treatment as illustrated in Fig. 18.

 Resulting sequences were clustered using an AlignX module commercial software package Vector NTI (<http://informaxinc.com>), which uses a Neighbor Joining algorithm for sequence clustering.

25 The resulting dendrogram is presented in Fig 18. On the dendrogram the closest "leafs" correspond to the closest pseudo-sequences. Interestingly, compounds with similar mechanisms of action cluster together on the dendrogram. Another example of the generation of pseudo-sequences and clustering is shown in Fig. 19.

30 In some embodiments, techniques according to the present invention can provide tools for the later stages of drug development such as clinical trial design and patient management. The properties of known drugs such as clinical trial and patient response information will be used in a similar fashion as the pre-clinical

information to provide predictions about the properties of novel compounds. Because the human cell is the locus of drug action, a database containing drug-cell interactions can be able to provide predictive information for this aspect of drug development.

Although the above has generally described the present invention

- 5 according to specific systems, the present invention has a much broader range of applicability. In particular, the present invention is not limited to a particular kind of data about a cell, but can be applied to virtually any cellular data where an understanding about the workings of the cell is desired. Thus, in some embodiments, the techniques of the present invention could provide information about many
- 10 different types or groups of cells, substances, and genetic processes of all kinds. Of course, one of ordinary skill in the art would recognize other variations, modifications, and alternatives.

APPENDIX A

EV Table 1.doc

Example of the output of AnalyseDNA.m program
(measurements for a single 3 by 3 montage image)

File	Subimage	object	X coord	Y coord	Area	Area ratio	Excitance	Equilibrium	Solidity	Extant	Intensity	Avg. Intensity	Median Intensity	20% pla.	80% pla.
1	1	1	12,2897	152,455	145	1.13253	0.337621	12.5075	0.923567	0.730756	1405	31.7586	31	75	37
1	1	2	16,7352	416,032	125	1.46594	0.762471	12.4137	0.903197	0.781755	1406	36.810	31	30	45
1	1	3	20,1073	72,079	177	1.09845	0.413705	15.0121	0.917098	0.691406	1769	28.9125	29	22	31
1	1	4	21,4186	402,764	13	1.32215	0.478004	7.3928	0.914994	0.747157	1690	85.811	81	67	105
1	1	5	27,0918	181	96	1.30887	0.445134	11.0538	0.881869	0.671329	1502	48.1938	49	38	56
1	1	6	30,2252	259,534	206	2.23106	0.903309	16.1953	0.927928	0.715278	1630	30.9709	33	24	37
1	1	7	31,4629	187,572	85	1.31981	0.471656	10.4531	0.927003	0.711667	1725	17.4719	50	39	56
1	1	8	35,0411	16,3726	166	1.23176	0.601695	12.4313	0.929336	0.710718	1615	37.089	40	29	44
1	1	9	37,786	364,023	47	1.40402	0.493562	7.73578	0.87037	0.652718	1667	141.831	142	113	171
1	1	10	49,1078	170,004	232	1.90491	0.851127	11.187	0.852911	0.70303	1832	42.3789	45	33	51
1	1	11	54,0749	126,534	271	1.25704	0.455955	16.7746	0.921666	0.666235	1700	31.8152	33	25	37
1	1	12	57,7155	44,9922	167	1.24677	0.461301	12.4809	0.907407	0.706731	1765	32.415	34	26	38
1	1	13	58,4444	366,854	272	2.27225	0.893953	16.7555	0.923468	0.706612	1978	51.8121	56	43	68
1	1	14	58,6029	282,272	206	1.82782	0.454914	18.1953	0.923167	0.64375	1737	31.4156	37	28	48
1	1	15	57,6618	227,136	108	1.21885	0.412266	20.0967	0.913254	0.701289	1614	42	45	33	51
1	1	16	68,1714	323,181	315	1.11184	0.427264	20.0967	0.75	0.536256	13151	48.0984	50	31	42
1	1	17	71,4449	463,733	185	1.27618	0.49066	16.7346	0.925092	0.670959	1809	44.5864	48	35	56
1	1	18	73,624	184,854	122	1.2588	0.411622	15.2476	0.91133	0.69323	1841	29.3577	41	20	29
1	1	19	78,7377	207,513	208	1.4378	0.781918	12.7386	0.923591	0.723231	1859	32.3577	41	20	29
1	1	20	81,0786	525,912	177	1.23513	0.461301	12.4738	0.91048	0.719194	1882	36.7159	40	28	42
1	1	21	81,0786	525,912	177	1.23513	0.461301	12.4738	0.91048	0.719194	1882	36.7159	40	28	42
1	1	22	81,0786	525,912	177	1.23513	0.461301	12.4738	0.91048	0.719194	1882	36.7159	40	28	42
1	1	23	81,0786	525,912	177	1.23513	0.461301	12.4738	0.91048	0.719194	1882	36.7159	40	28	42
1	1	24	81,0786	525,912	177	1.23513	0.461301	12.4738	0.91048	0.719194	1882	36.7159	40	28	42
1	1	25	81,0786	525,912	177	1.23513	0.461301	12.4738	0.91048	0.719194	1882	36.7159	40	28	42
1	1	26	81,0786	525,912	177	1.23513	0.461301	12.4738	0.91048	0.719194	1882	36.7159	40	28	42
1	1	27	81,0786	525,912	177	1.23513	0.461301	12.4738	0.91048	0.719194	1882	36.7159	40	28	42
1	1	28	81,0786	525,912	177	1.23513	0.461301	12.4738	0.91048	0.719194	1882	36.7159	40	28	42
1	1	29	81,0786	525,912	177	1.23513	0.461301	12.4738	0.91048	0.719194	1882	36.7159	40	28	42
1	1	30	81,0786	525,912	177	1.23513	0.461301	12.4738	0.91048	0.719194	1882	36.7159	40	28	42
1	1	31	102,47	155,307	122	1.3708	0.733131	12.4619	0.917808	0.691739	1958	28.2272	34	27	36
1	1	32	105,356	57,1271	118	1.70239	0.450499	12.2573	0.901892	0.691119	1695	39.7601	42	30	48
1	1	33	121,23	285,08	324	1.70239	0.450499	12.2573	0.901892	0.691119	1695	39.7601	42	30	48
1	1	34	125,532	170,645	141	1.53045	0.757011	12.3988	0.927432	0.736275	1829	45.3557	49	37	53
1	1	35	128,98	60,3355	152	1.75489	0.827208	12.3116	0.921212	0.747477	1875	45.2303	47	36	54
1	1	36	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	37	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	38	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	39	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	40	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	41	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	42	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	43	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	44	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	45	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	46	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	47	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	48	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	49	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	50	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	51	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	52	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	53	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	54	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	55	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	56	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	57	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	58	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	59	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	60	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	61	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	62	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	63	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	64	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	65	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	66	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	67	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43
1	1	68	137,092	128,083	266	1.865	0.810275	18.1033	0.956407	0.538652	1910	36.8797	38	26	43

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1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60	61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77	78	79	80	81	82	83	84	85	86	87	88	89	90	91	92	93	94	95	96	97	98	99	100	101	102	103	104	105	106	107	108	109	110	111	112	113	114	115	116	117	118	119	120	121	122	123	124	125	126	127	128	129	130	131	132	133	134	135	136	137	138	139	140	141	142	143	144	145	146	147	148	149	150	151	152	153	154	155	156	157	158	159	160	161	162	163	164	165	166	167	168	169	170	171	172	173	174	175	176	177	178	179	180	181	182	183	184	185	186	187	188	189	190	191	192	193	194	195	196	197	198	199	200	201	202	203	204	205	206	207	208	209	210	211	212	213	214	215	216	217	218	219	220	221	222	223	224	225	226	227	228	229	230	231	232	233	234	235	236	237	238	239	240	241	242	243	244	245	246	247	248	249	250	251	252	253	254	255	256	257	258	259	260	261	262	263	264	265	266	267	268	269	270	271	272	273	274	275	276	277	278	279	280	281	282	283	284	285	286	287	288	289	290	291	292	293	294	295	296	297	298	299	300	301	302	303	304	305	306	307	308	309	310	311	312	313	314	315	316	317	318	319	320	321	322	323	324	325	326	327	328	329	330	331	332	333	334	335	336	337	338	339	340	341	342	343	344	345	346	347	348	349	350	351	352	353	354	355	356	357	358	359	360	361	362	363	364	365	366	367	368	369	370	371	372	373	374	375	376	377	378	379	380	381	382	383	384	385	386	387	388	389	390	391	392	393	394	395	396	397	398	399	400	401	402	403	404	405	406	407	408	409	410	411	412	413	414	415	416	417	418	419	420	421	422	423	424	425	426	427	428	429	430	431	432	433	434	435	436	437	438	439	440	441	442	443	444	445	446	447	448	449	450	451	452	453	454	455	456	457	458	459	460	461	462	463	464	465	466	467	468	469	470	471	472	473	474	475	476	477	478	479	480	481	482	483	484	485	486	487	488	489	490	491	492	493	494	495	496	497	498	499	500	501	502	503	504	505	506	507	508	509	510	511	512	513	514	515	516	517	518	519	520	521	522	523	524	525	526	527	528	529	530	531	532	533	534	535	536	537	538	539	540	541	542	543	544	545	546	547	548	549	550	551	552	553	554	555	556	557	558	559	560	561	562	563	564	565	566	567	568	569	570	571	572	573	574	575	576	577	578	579	580	581	582	583	584	585	586	587	588	589	590	591	592	593	594	595	596	597	598	599	600	601	602	603	604	605	606	607	608	609	610	611	612	613	614	615	616	617	618	619	620	621	622	623	624	625	626	627	628	629	630	631	632	633	634	635	636	637	638	639	640	641	642	643	644	645	646	647	648	649	650	651	652	653	654	655	656	657	658	659	660	661	662	663	664	665	666	667	668	669	670	671	672	673	674	675	676	677	678	679	680	681	682	683	684	685	686	687	688	689	690	691	692	693	694	695	696	697	698	699	700	701	702	703	704	705	706	707	708	709	710	711	712	713	714	715	716	717	718	719	720	721	722	723	724	725	726	727	728	729	730	731	732	733	734	735	736	737	738	739	740	741	742	743	744	745	746	747	748	749	750	751	752	753	754	755	756	757	758	759	760	761	762	763	764	765	766	767	768	769	770	771	772	773	774	775	776	777	778	779	780	781	782	783	784	785	786	787	788	789	790	791	792	793	794	795	796	797	798	799	800	801	802	803	804	805	806	807	808	809	810	811	812	813	814	815	816	817	818	819	820	821	822	823	824	825	826	827	828	829	830	831	832	833	834	835	836	837	838	839	840	841	842	843	844	845	846	847	848	849	850	851	852	853	854	855	856	857	858	859	860	861	862	863	864	865	866	867	868	869	870	871	872	873	874	875	876	877	878	879	880	881	882	883	884	885	886	887	888	889	890	891	892	893	894	895	896	897	898	899	900	901	902	903	904	905	906	907	908	909	910	911	912	913	914	915	916	917	918	919	920	921	922	923	924	925	926	927	928	929	930	931	932	933	934	935	936	937	938	939	940	941	942	943	944	945	946	947	948	949	950	951	952	953	954	955	956	957	958	959	960	961	962	963	964	965	966	967	968	969	970	971	972	973	974	975	976	977	978	979	980	981	982	983	984	985	986	987	988	989	990	991	992	993	994	995	996	997	998	999	1000
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1	136	105.00	356.469	237	2.80072	0.331005	19.5115	0.89221	0.64891	11065	43.0401	50	38	57	
2	137	106.132	356.267	190	1.73179	0.292039	15.5935	0.852556	0.61667	9786	46.2121	50	38	57	
3	138	107.330	357.207	195	1.3628	0.267935	15.757	0.820018	0.77281	5506	27.7322	50	38	57	
4	139	108.090	358.076	102	1.21531	0.346019	11.3961	0.815919	0.71287	5508	51	50	40	60	
5	140	109.165	359.585	167	1.49992	0.450132	13.6808	0.86021	0.57677	5099	36.4893	51	39	41	61
6	141	110.515	361.281	176	1.49894	0.741034	14.9556	0.876316	0.67059	8466	55.0569	46	45	44	62
7	142	111.715	363.431	176	1.20928	0.562256	10.2179	0.881172	0.693333	4120	53.8924	56	45	44	63
8	143	112.915	365.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	64
9	144	114.115	367.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	65
10	145	115.315	369.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	66
11	146	116.515	371.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	67
12	147	117.715	373.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	68
13	148	118.915	375.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	69
14	149	120.115	377.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	70
15	150	121.315	379.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	71
16	151	122.515	381.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	72
17	152	123.715	383.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	73
18	153	124.915	385.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	74
19	154	126.115	387.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	75
20	155	127.315	389.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	76
21	156	128.515	391.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	77
22	157	129.715	393.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	78
23	158	130.915	395.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	79
24	159	132.115	397.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	80
25	160	133.315	399.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	81
26	161	134.515	401.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	82
27	162	135.715	403.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	83
28	163	136.915	405.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	84
29	164	138.115	407.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	85
30	165	139.315	409.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	86
31	166	140.515	411.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	87
32	167	141.715	413.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	88
33	168	142.915	415.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	89
34	169	144.115	417.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	90
35	170	145.315	419.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	91
36	171	146.515	421.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	92
37	172	147.715	423.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	93
38	173	148.915	425.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	94
39	174	150.115	427.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	95
40	175	151.315	429.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	96
41	176	152.515	431.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	97
42	177	153.715	433.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	98
43	178	154.915	435.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	99
44	179	156.115	437.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	100
45	180	157.315	439.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	101
46	181	158.515	441.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	102
47	182	159.715	443.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	103
48	183	160.915	445.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	104
49	184	162.115	447.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	105
50	185	163.315	449.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	106
51	186	164.515	451.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	107
52	187	165.715	453.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	108
53	188	166.915	455.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	109
54	189	168.115	457.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	110
55	190	169.315	459.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	111
56	191	170.515	461.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	112
57	192	171.715	463.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	113
58	193	172.915	465.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	114
59	194	174.115	467.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	115
60	195	175.315	469.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	116
61	196	176.515	471.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	117
62	197	177.715	473.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	118
63	198	178.915	475.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	119
64	199	180.115	477.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	120
65	200	181.315	479.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	121
66	201	182.515	481.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	122
67	202	183.715	483.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	123
68	203	184.915	485.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	124
69	204	186.115	487.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	125
70	205	187.315	489.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	126
71	206	188.515	491.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	127
72	207	189.715	493.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	128
73	208	190.915	495.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	129
74	209	192.115	497.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	53.8924	56	45	44	130
75	210	193.315	499.431	176	1.49712	0.741034	14.9556	0.881172	0.693333	4120	5				

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1	1	270.231	432.417	177	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	2	272.404	432.243	221	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	3	274.577	432.069	265	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	4	276.750	431.895	309	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	5	278.923	431.721	353	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	6	281.096	431.547	397	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	7	283.269	431.373	441	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	8	285.442	431.199	485	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	9	287.615	431.025	529	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	10	289.788	430.851	573	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	11	291.961	430.677	617	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	12	294.134	430.503	661	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	13	296.307	430.329	705	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	14	298.480	430.155	749	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	15	300.653	429.981	793	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	16	302.826	429.807	837	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	17	304.999	429.633	881	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	18	307.172	429.459	925	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	19	309.345	429.285	969	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	20	311.518	429.111	1013	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	21	313.691	428.937	1057	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	22	315.864	428.763	1101	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	23	318.037	428.589	1145	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	24	320.210	428.415	1189	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	25	322.383	428.241	1233	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	26	324.556	428.067	1277	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	27	326.729	427.893	1321	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	28	328.902	427.719	1365	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	29	331.075	427.545	1409	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	30	333.248	427.371	1453	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	31	335.421	427.197	1497	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	32	337.594	427.023	1541	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	33	339.767	426.849	1585	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	34	341.940	426.675	1629	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	35	344.113	426.501	1673	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	36	346.286	426.327	1717	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	37	348.459	426.153	1761	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	38	350.632	425.979	1805	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	39	352.805	425.805	1849	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	40	354.978	425.631	1893	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	41	357.151	425.457	1937	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	42	359.324	425.283	1981	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	43	361.497	425.109	2025	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	44	363.670	424.935	2069	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	45	365.843	424.761	2113	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	46	368.016	424.587	2157	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	47	370.189	424.413	2201	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	48	372.362	424.239	2245	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	49	374.535	424.065	2289	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	50	376.708	423.891	2333	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	51	378.881	423.717	2377	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	52	381.054	423.543	2421	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	53	383.227	423.369	2465	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	54	385.400	423.195	2509	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	55	387.573	423.021	2553	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	56	389.746	422.847	2597	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	57	391.919	422.673	2641	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	58	394.092	422.499	2685	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	59	396.265	422.325	2729	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	60	398.438	422.151	2773	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	61	400.611	421.977	2817	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	62	402.784	421.803	2861	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	63	404.957	421.629	2905	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	64	407.130	421.455	2949	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	65	409.303	421.281	2993	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	66	411.476	421.107	3037	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	67	413.649	420.933	3081	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	68	415.822	420.759	3125	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	69	417.995	420.585	3169	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	70	420.168	420.411	3213	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	71	422.341	420.237	3257	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	72	424.514	420.063	3301	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	73	426.687	419.889	3345	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	74	428.860	419.715	3389	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	75	431.033	419.541	3433	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	76	433.206	419.367	3477	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	77	435.379	419.193	3521	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	78	437.552	419.019	3565	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	79	439.725	418.845	3609	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	80	441.898	418.671	3653	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	81	444.071	418.497	3697	1.27886	0.421112	15.0121	0.317098	0.715457	97169	35.0739	35	69
1	82	446.244	418.323	3741	1.27886	0.421112	15.0121	0.317098	0.715457	97169			

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1	150	151	152	153	154	155	156	157	158	159	160	161	162	163	164	165	166	167	168	169	170	171	172	173	174	175	176	177	178	179	180	181	182	183	184	185	186	187	188	189	190	191	192	193	194	195	196	197	198	199	200	201	202	203	204	205	206	207	208	209	210	211	212	213	214	215	216	217	218	219	220	221	222	223	224	225	226	227	228	229	230	231	232	233	234	235	236	237	238	239	240	241	242	243	244	245	246	247	248	249	250	251	252	253	254	255	256	257	258	259	260	261	262	263	264	265	266	267	268	269	270	271	272	273	274	275	276	277	278	279	280	281	282	283	284	285	286	287	288	289	290	291	292	293	294	295	296	297	298	299	300	301	302	303	304	305	306	307	308	309	310	311	312	313	314	315	316	317	318	319	320	321	322	323	324	325	326	327	328	329	330	331	332	333	334	335	336	337	338	339	340	341	342	343	344	345	346	347	348	349	350	351	352	353	354	355	356	357	358	359	360	361	362	363	364	365	366	367	368	369	370	371	372	373	374	375	376	377	378	379	380	381	382	383	384	385	386	387	388	389	390	391	392	393	394	395	396	397	398	399	400	401	402	403	404	405	406	407	408	409	410	411	412	413	414	415	416	417	418	419	420	421	422	423	424	425	426	427	428	429	430	431	432	433	434	435	436	437	438	439	440	441	442	443	444	445	446	447	448	449	450	451	452	453	454	455	456	457	458	459	460	461	462	463	464	465	466	467	468	469	470	471	472	473	474	475	476	477	478	479	480	481	482	483	484	485	486	487	488	489	490	491	492	493	494	495	496	497	498	499	500	501	502	503	504	505	506	507	508	509	510	511	512	513	514	515	516	517	518	519	520	521	522	523	524	525	526	527	528	529	530	531	532	533	534	535	536	537	538	539	540	541	542	543	544	545	546	547	548	549	550	551	552	553	554	555	556	557	558	559	560	561	562	563	564	565	566	567	568	569	570	571	572	573	574	575	576	577	578	579	580	581	582	583	584	585	586	587	588	589	590	591	592	593	594	595	596	597	598	599	600	601	602	603	604	605	606	607	608	609	610	611	612	613	614	615	616	617	618	619	620	621	622	623	624	625	626	627	628	629	630	631	632	633	634	635	636	637	638	639	640	641	642	643	644	645	646	647	648	649	650	651	652	653	654	655	656	657	658	659	660	661	662	663	664	665	666	667	668	669	670	671	672	673	674	675	676	677	678	679	680	681	682	683	684	685	686	687	688	689	690	691	692	693	694	695	696	697	698	699	700	701	702	703	704	705	706	707	708	709	710	711	712	713	714	715	716	717	718	719	720	721	722	723	724	725	726	727	728	729	730	731	732	733	734	735	736	737	738	739	740	741	742	743	744	745	746	747	748	749	750	751	752	753	754	755	756	757	758	759	760	761	762	763	764	765	766	767	768	769	770	771	772	773	774	775	776	777	778	779	780	781	782	783	784	785	786	787	788	789	790	791	792	793	794	795	796	797	798	799	800	801	802	803	804	805	806	807	808	809	810	811	812	813	814	815	816	817	818	819	820	821	822	823	824	825	826	827	828	829	830	831	832	833	834	835	836	837	838	839	840	841	842	843	844	845	846	847	848	849	850	851	852	853	854	855	856	857	858	859	860	861	862	863	864	865	866	867	868	869	870	871	872	873	874	875	876	877	878	879	880	881	882	883	884	885	886	887	888	889	890	891	892	893	894	895	896	897	898	899	900	901	902	903	904	905	906	907	908	909	910	911	912	913	914	915	916	917	918	919	920	921	922	923	924	925	926	927	928	929	930	931	932	933	934	935	936	937	938	939	940	941	942	943	944	945	946	947	948	949	950	951	952	953	954	955	956	957	958	959	960	961	962	963	964	965	966	967	968	969	970	971	972	973	974	975	976	977	978	979	980	981	982	983	984	985	986	987	988	989	990	991	992	993	994	995	996	997	998	999	1000
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1	1	36	214.803	44.7106	334	1.70545	0.810101	20.4219	0.805912	0.702158	15564	46.508	19	38	55
1	2	37	341.431	115.89	145	1.57559	0.772771	12.3975	0.917722	0.491113	4489	32.2379	32	45	39
1	3	38	345.761	264.232	172	1.43232	0.725724	12.8815	0.905759	0.468508	2922	51.731	36	44	64
1	4	39	359.265	297.404	146	1.74553	0.806127	13.2723	0.907916	0.468633	4158	41.4041	41	34	50
1	5	40	365.892	35.5	186	1.49233	0.806127	13.2723	0.907916	0.468633	4158	41.4041	41	34	50
1	6	41	367.611	474.44	208	1.49233	0.806127	13.2723	0.907916	0.468633	4158	41.4041	41	34	50
1	7	42	374.033	254.171	152	1.46026	0.794253	13.3116	0.91018	0.462087	12451	41.875	41	34	50
1	8	43	388.878	481.764	123	1.4018	0.731849	13.4143	0.91018	0.462087	12451	41.875	41	34	50
1	9	44	395.075	481.764	123	1.4018	0.731849	13.4143	0.91018	0.462087	12451	41.875	41	34	50
1	10	45	310.406	479.341	122	1.72398	0.810231	12.4434	0.917293	0.723274	5724	36.2053	40	30	46
1	11	46	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	12	47	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	13	48	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	14	49	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	15	50	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	16	51	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	17	52	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	18	53	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	19	54	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	20	55	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	21	56	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	22	57	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	23	58	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	24	59	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	25	60	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	26	61	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	27	62	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	28	63	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	29	64	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	30	65	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	31	66	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	32	67	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	33	68	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	34	69	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	35	70	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	36	71	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	37	72	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	38	73	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	39	74	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	40	75	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	41	76	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	42	77	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	43	78	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	44	79	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	45	80	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	46	81	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	47	82	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	48	83	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	49	84	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	50	85	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	51	86	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	52	87	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	53	88	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	54	89	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	55	90	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	56	91	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	57	92	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	58	93	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	59	94	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	60	95	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	61	96	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	62	97	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	63	98	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	64	99	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46
1	65	100	314.624	419.341	122	1.55255	0.755216	12.3919	0.917293	0.723274	5724	36.2053	40	30	46

1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60	61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77	78	79	80	81	82	83	84	85	86	87	88	89	90	91	92	93	94	95	96	97	98	99	100	101	102	103	104	105	106	107	108	109	110	111	112	113	114	115	116	117	118	119	120	121	122	123	124	125	126	127	128	129	130	131	132	133	134	135	136	137	138	139	140	141	142	143	144	145	146	147	148	149	150	151	152	153	154	155	156	157	158	159	160	161	162	163	164	165	166	167	168	169	170	171	172	173	174	175	176	177	178	179	180	181	182	183	184	185	186	187	188	189	190	191	192	193	194	195	196	197	198	199	200	201	202	203	204	205	206	207	208	209	210	211	212	213	214	215	216	217	218	219	220	221	222	223	224	225	226	227	228	229	230	231	232	233	234	235	236	237	238	239	240	241	242	243	244	245	246	247	248	249	250	251	252	253	254	255	256	257	258	259	260	261	262	263	264	265	266	267	268	269	270	271	272	273	274	275	276	277	278	279	280	281	282	283	284	285	286	287	288	289	290	291	292	293	294	295	296	297	298	299	300	301	302	303	304	305	306	307	308	309	310	311	312	313	314	315	316	317	318	319	320	321	322	323	324	325	326	327	328	329	330	331	332	333	334	335	336	337	338	339	340	341	342	343	344	345	346	347	348	349	350	351	352	353	354	355	356	357	358	359	360	361	362	363	364	365	366	367	368	369	370	371	372	373	374	375	376	377	378	379	380	381	382	383	384	385	386	387	388	389	390	391	392	393	394	395	396	397	398	399	400	401	402	403	404	405	406	407	408	409	410	411	412	413	414	415	416	417	418	419	420	421	422	423	424	425	426	427	428	429	430	431	432	433	434	435	436	437	438	439	440	441	442	443	444	445	446	447	448	449	450	451	452	453	454	455	456	457	458	459	460	461	462	463	464	465	466	467	468	469	470	471	472	473	474	475	476	477	478	479	480	481	482	483	484	485	486	487	488	489	490	491	492	493	494	495	496	497	498	499	500	501	502	503	504	505	506	507	508	509	510	511	512	513	514	515	516	517	518	519	520	521	522	523	524	525	526	527	528	529	530	531	532	533	534	535	536	537	538	539	540	541	542	543	544	545	546	547	548	549	550	551	552	553	554	555	556	557	558	559	560	561	562	563	564	565	566	567	568	569	570	571	572	573	574	575	576	577	578	579	580	581	582	583	584	585	586	587	588	589	590	591	592	593	594	595	596	597	598	599	600	601	602	603	604	605	606	607	608	609	610	611	612	613	614	615	616	617	618	619	620	621	622	623	624	625	626	627	628	629	630	631	632	633	634	635	636	637	638	639	640	641	642	643	644	645	646	647	648	649	650	651	652	653	654	655	656	657	658	659	660	661	662	663	664	665	666	667	668	669	670	671	672	673	674	675	676	677	678	679	680	681	682	683	684	685	686	687	688	689	690	691	692	693	694	695	696	697	698	699	700	701	702	703	704	705	706	707	708	709	710	711	712	713	714	715	716	717	718	719	720	721	722	723	724	725	726	727	728	729	730	731	732	733	734	735	736	737	738	739	740	741	742	743	744	745	746	747	748	749	750	751	752	753	754	755	756	757	758	759	760	761	762	763	764	765	766	767	768	769	770	771	772	773	774	775	776	777	778	779	780	781	782	783	784	785	786	787	788	789	790	791	792	793	794	795	796	797	798	799	800	801	802	803	804	805	806	807	808	809	810	811	812	813	814	815	816	817	818	819	820	821	822	823	824	825	826	827	828	829	830	831	832	833	834	835	836	837	838	839	840	841	842	843	844	845	846	847	848	849	850	851	852	853	854	855	856	857	858	859	860	861	862	863	864	865	866	867	868	869	870	871	872	873	874	875	876	877	878	879	880	881	882	883	884	885	886	887	888	889	890	891	892	893	894	895	896	897	898	899	900	901	902	903	904	905	906	907	908	909	910	911	912	913	914	915	916	917	918	919	920	921	922	923	924	925	926	927	928	929	930	931	932	933	934	935	936	937	938	939	940	941	942	943	944	945	946	947	948	949	950	951	952	953	954	955	956	957	958	959	960	961	962	963	964	965	966	967	968	969	970	971	972	973	974	975	976	977	978	979	980	981	982	983	984	985	986	987	988	989	990	991	992	993	994	995	996	997	998	999	1000
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EV Table 1.doc

1	106	359,016	332,35	116	1,70064	0,477012	12,0018	0,491628	0,475376	4181	36,4354	38	78	43
2	107	361,716	195,031	129	1,40135	0,701018	12,1153	0,502451	0,461538	7538	36,4186	40	16	71
3	108	370,43	126,784	132	1,127164	0,472767	11,9416	0,502226	0,717949	4216	37,4007	30	46	64
4	109	379,866	101,258	57	1,215377	0,548277	11,1132	0,538110	0,714410	4116	42,7016	14	34	51
5	110	382,727	60,484	187	1,748828	0,820329	15,1204	0,912153	0,411111	8530	44,1093	44	34	51
6	111	388,96	193,188	201	1,187034	0,635633	19,4416	0,907198	0,411735	11210	36,9315	39	39	81
7	112	387,155	331,22	174	1,735584	0,811993	15,4416	0,915109	0,715	8157	36,3716	32	39	81
8	113	384,664	261,441	136	1,46281	0,801233	13,1559	0,891337	0,413816	15371	32,4346	35	27	61
9	114	401,478	202,432	414	1,512937	0,737039	12,3551	0,844109	0,913731	151684	31,5339	30	32	61
10	115	385,517	273,516	149	2,06818	0,872145	12,7716	0,925166	0,411731	151684	36,1136	38	29	61
11	116	411,693	273,516	149	1,50902	0,748901	11,7186	0,873371	0,411731	151684	36,0766	37	29	61
12	117	421,436	70,7642	159	1,6316	0,790105	11,2783	0,910128	0,411731	151684	38,1804	37	30	41
13	118	425	14	109	1,24376	0,591149	11,7186	0,911628	0,411731	151684	35,1031	37	30	41
14	119	416,501	176,255	132	1,32341	0,603166	12,7661	0,922077	0,723333	10217	35,8515	24	30	38
15	120	402,616	371,2	393	1,44131	0,40148	12,1211	0,915107	0,411731	151684	35,8515	24	30	38
16	121	435,448	104,209	91	1,47731	0,733353	10,1601	0,911318	0,438844	8120	41,3176	42	33	51
17	122	435,448	104,209	91	1,71053	0,818188	11,3119	0,911136	0,727718	4281	31,7023	34	25	39
18	123	435,448	245,441	131	1,51612	0,735103	11,3119	0,914327	0,763316	4245	36,4091	40	29	47
19	124	401,904	87,4919	118	1,54828	0,735416	15,0515	0,912728	0,458172	4184	35,1497	27	20	30
20	125	430,265	484,51	113	1,27401	0,464112	11,9948	0,911647	0,802143	4153	38,4106	34	29	44
21	126	451,705	140,813	103	1,25242	0,464112	11,9948	0,911647	0,802143	4153	36,2364	34	29	44
22	127	456,132	139,5	100	1,27418	0,464112	11,9948	0,911647	0,802143	4153	36,2364	34	29	44
23	128	461,271	112,424	123	1,7004	0,809351	12,3131	0,915107	0,719231	3958	38,58	41	31	47
24	129	463,54	184,828	227	1,7029	0,810337	16,0183	0,911318	0,438844	8120	36,4339	38	28	44
25	130	472,742	297,525	101	1,12931	0,441536	11,2701	0,90931	0,715151	8020	36,4339	38	28	44
26	131	472,742	297,525	101	1,21653	0,549329	12,1525	0,923567	0,723264	4208	36,4339	38	28	44
27	132	472,742	312,506	116	1,10233	0,423339	12,1525	0,913386	0,715559	4237	36,4339	38	28	44
28	133	472,742	312,506	116	1,47045	0,710199	11,2706	0,923238	0,720531	4276	36,4339	38	28	44
29	134	472,742	312,506	116	2,09652	0,816005	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
30	135	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
31	136	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
32	137	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
33	138	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
34	139	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
35	140	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
36	141	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
37	142	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
38	143	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
39	144	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
40	145	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
41	146	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
42	147	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
43	148	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
44	149	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
45	150	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
46	151	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
47	152	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
48	153	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
49	154	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
50	155	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
51	156	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
52	157	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
53	158	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
54	159	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
55	160	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
56	161	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
57	162	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
58	163	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
59	164	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
60	165	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
61	166	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
62	167	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
63	168	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
64	169	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
65	170	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
66	171	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
67	172	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
68	173	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
69	174	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
70	175	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
71	176	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
72	177	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
73	178	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
74	179	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
75	180	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
76	181	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
77	182	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
78	183	472,742	312,506	116	1,47045	0,710199	11,2706	0,914201	0,710551	4276	36,4339	38	28	44
79	184													

1	1	131.431	616.093	116	1.53941	0.77101	12.3333	0.30331	0.3235	4170	29	39	37.039	29	41
2	1	161.089	179.484	232	1.64547	0.81191	15.4333	0.31666	0.42743	4755	19	29	26.1656	19	31
3	1	168.375	177.243	298	1.61175	0.83873	17.4788	0.403204	0.325532	5510	33	39	32.8128	33	39
4	1	245.757	212.466	161	1.1501	0.69161	13.3988	0.315594	0.725072	3111	24	33	30.5937	24	33
5	1	148.47	16.2418	203	1.35581	0.67093	16.9759	0.401448	0.748156	4182	47	55	45.2319	47	55
6	1	132.921	219.331	199	1.60944	0.80053	13.3338	0.335641	0.35	4303	26	30	25.8111	26	30
7	1	150.319	93.4716	149	1.91944	0.85132	11.4668	0.351037	0.470833	4649	29	31	27.5099	29	31
8	1	153.972	245.497	187	2.01842	0.87125	13.1394	0.321112	0.431034	7571	40	48	40.1856	40	48
9	1	155.578	139.748	135	1.54598	0.76104	13.1166	0.312163	0.407083	4282	31	39	27.7926	31	39
10	1	144.222	248.369	188	2.46046	0.91383	13.9773	0.31053	0.407083	4282	31	39	27.7926	31	39
11	1	177.049	501.09	316	3.04239	0.91161	20.4835	0.433821	0.718462	9428	21	21	26.4394	21	21
12	1	179.35	180.975	160	1.74376	0.83223	11.273	0.31954	0.431741	4174	27	31	27.915	27	31
13	1	182.463	312.953	177	2.04101	0.87101	13.0121	0.307621	0.516131	6096	34	41	34.4107	34	41
14	1	182.642	312.948	173	1.33589	0.71432	13.4739	0.481279	0.709539	8159	43	51	43.829	43	51
15	1	183.963	145.944	143	1.62117	0.80117	13.4735	0.489371	0.410559	4742	21	26	20.2457	21	26
16	1	182.925	326.351	131	1.32067	0.63064	11.9149	0.315714	0.719742	4343	31	34	31.1521	31	34
17	1	191.743	254.019	348	1.6183	0.80153	22.7215	0.461441	0.446447	4974	32	40	42.8255	32	40
18	1	187.464	444.44	155	1.91591	0.78121	13.1166	0.312163	0.407083	4282	31	39	27.7926	31	39
19	1	179.35	180.975	160	1.74376	0.83223	11.273	0.31954	0.431741	4174	27	31	27.915	27	31
20	1	182.463	312.953	177	2.04101	0.87101	13.0121	0.307621	0.516131	6096	34	41	34.4107	34	41
21	1	182.642	312.948	173	1.33589	0.71432	13.4739	0.481279	0.709539	8159	43	51	43.829	43	51
22	1	183.963	145.944	143	1.62117	0.80117	13.4735	0.489371	0.410559	4742	21	26	20.2457	21	26
23	1	182.925	326.351	131	1.32067	0.63064	11.9149	0.315714	0.719742	4343	31	34	31.1521	31	34
24	1	191.743	254.019	348	1.6183	0.80153	22.7215	0.461441	0.446447	4974	32	40	42.8255	32	40
25	1	187.464	444.44	155	1.91591	0.78121	13.1166	0.312163	0.407083	4282	31	39	27.7926	31	39
26	1	179.35	180.975	160	1.74376	0.83223	11.273	0.31954	0.431741	4174	27	31	27.915	27	31
27	1	182.463	312.953	177	2.04101	0.87101	13.0121	0.307621	0.516131	6096	34	41	34.4107	34	41
28	1	182.642	312.948	173	1.33589	0.71432	13.4739	0.481279	0.709539	8159	43	51	43.829	43	51
29	1	183.963	145.944	143	1.62117	0.80117	13.4735	0.489371	0.410559	4742	21	26	20.2457	21	26
30	1	182.925	326.351	131	1.32067	0.63064	11.9149	0.315714	0.719742	4343	31	34	31.1521	31	34
31	1	191.743	254.019	348	1.6183	0.80153	22.7215	0.461441	0.446447	4974	32	40	42.8255	32	40
32	1	187.464	444.44	155	1.91591	0.78121	13.1166	0.312163	0.407083	4282	31	39	27.7926	31	39
33	1	179.35	180.975	160	1.74376	0.83223	11.273	0.31954	0.431741	4174	27	31	27.915	27	31
34	1	182.463	312.953	177	2.04101	0.87101	13.0121	0.307621	0.516131	6096	34	41	34.4107	34	41
35	1	182.642	312.948	173	1.33589	0.71432	13.4739	0.481279	0.709539	8159	43	51	43.829	43	51
36	1	183.963	145.944	143	1.62117	0.80117	13.4735	0.489371	0.410559	4742	21	26	20.2457	21	26
37	1	182.925	326.351	131	1.32067	0.63064	11.9149	0.315714	0.719742	4343	31	34	31.1521	31	34
38	1	191.743	254.019	348	1.6183	0.80153	22.7215	0.461441	0.446447	4974	32	40	42.8255	32	40
39	1	187.464	444.44	155	1.91591	0.78121	13.1166	0.312163	0.407083	4282	31	39	27.7926	31	39
40	1	179.35	180.975	160	1.74376	0.83223	11.273	0.31954	0.431741	4174	27	31	27.915	27	31
41	1	182.463	312.953	177	2.04101	0.87101	13.0121	0.307621	0.516131	6096	34	41	34.4107	34	41
42	1	182.642	312.948	173	1.33589	0.71432	13.4739	0.481279	0.709539	8159	43	51	43.829	43	51
43	1	183.963	145.944	143	1.62117	0.80117	13.4735	0.489371	0.410559	4742	21	26	20.2457	21	26
44	1	182.925	326.351	131	1.32067	0.63064	11.9149	0.315714	0.719742	4343	31	34	31.1521	31	34
45	1	191.743	254.019	348	1.6183	0.80153	22.7215	0.461441	0.446447	4974	32	40	42.8255	32	40
46	1	187.464	444.44	155	1.91591	0.78121	13.1166	0.312163	0.407083	4282	31	39	27.7926	31	39
47	1	179.35	180.975	160	1.74376	0.83223	11.273	0.31954	0.431741	4174	27	31	27.915	27	31
48	1	182.463	312.953	177	2.04101	0.87101	13.0121	0.307621	0.516131	6096	34	41	34.4107	34	41
49	1	182.642	312.948	173	1.33589	0.71432	13.4739	0.481279	0.709539	8159	43	51	43.829	43	51
50	1	183.963	145.944	143	1.62117	0.80117	13.4735	0.489371	0.410559	4742	21	26	20.2457	21	26
51	1	182.925	326.351	131	1.32067	0.63064	11.9149	0.315714	0.719742	4343	31	34	31.1521	31	34
52	1	191.743	254.019	348	1.6183	0.80153	22.7215	0.461441	0.446447	4974	32	40	42.8255	32	40
53	1	187.464	444.44	155	1.91591	0.78121	13.1166	0.312163	0.407083	4282	31	39	27.7926	31	39
54	1	179.35	180.975	160	1.74376	0.83223	11.273	0.31954	0.431741	4174	27	31	27.915	27	31
55	1	182.463	312.953	177	2.04101	0.87101	13.0121	0.307621	0.516131	6096	34	41	34.4107	34	41
56	1	182.642	312.948	173	1.33589	0.71432	13.4739	0.481279	0.709539	8159	43	51	43.829	43	51
57	1	183.963	145.944	143	1.62117	0.80117	13.4735	0.489371	0.410559	4742	21	26	20.2457	21	26
58	1	182.925	326.351	131	1.32067	0.63064	11.9149	0.315714	0.719742	4343	31	34	31.1521	31	34
59	1	191.743	254.019	348	1.6183	0.80153	22.7215	0.461441	0.446447	4974	32	40	42.8255	32	40
60	1	187.464	444.44	155	1.91591	0.78121	13.1166	0.312163	0.407083	4282	31	39	27.7926	31	39
61	1	179.35	180.975	160	1.74376	0.83223	11.273	0.31954	0.431741	4174	27	31	27.915	27	31
62	1	182.463	312.953	177	2.04101	0.87101	13.0121	0.307621	0.516131	6096	34	41	34.4107	34	41
63	1	182.642	312.948	173	1.33589	0.71432	13.4739	0.481279	0.709539	8159	43	51	43.829	43	51
64	1	183.963	145.944	143	1.62117	0.80117	13.4735	0.489371	0.410559	4742	21	26	20.2457	21	26
65	1	182.925	326.351	131	1.32067	0.63064	11.9149	0.315714	0.719742	4343	31	34	31.1521	31	34
66	1	191.743	254.019	348	1.6183	0.80153	22.7215	0.461441	0.446447	4974	32	40	42.8255	32	40
67	1	187.464	444.44	155	1.91591	0.78121	13.1166	0.312163	0.407083	4282	31	39	27.7926	31	39
68	1	179.35	180.975	160	1.74376	0.83223	11.273	0.31954	0.431741	4174	27	31	27.915	27	31
69	1	182.463	312.953	177	2.04101	0.87101	13.0121	0.307621	0.516131	6096	34	41	34.4107	34	41
70	1	182.642	312.948	173	1.33589	0.71432	13.4739	0.481279	0.709539	8159	43	51	43.829	43	51
71	1	183.963	145.944	143	1.62117	0.80117	13.4735	0.489371	0.410559	4742	21	26	20.2457	21	26
72	1	182.925	326.351	131	1.32067	0.63064	11.9149	0.315714	0.719742	4343	31	34	31.1521	31	34
73	1	191.743	254.019	348	1.6183	0.80153	22.7215	0.461441	0.446447	4974	32	40	42.8255	32	40
74	1	187.464	444.44	155	1.91591	0.78121	13.1166	0.312163	0.407083	4282	31	39	27.7926	31	39
75	1	179.35	180.975	160	1.74376	0.83223	11.273	0.31954	0.431741	4174	27	31	27.915	27	31
76	1	182.463	312.953	177	2.04101	0.87101	13.0121	0.307621	0.516131	6096	34	41	34.4107	34	41
77	1	182.642	312.948	173	1.33589	0.71432									

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EV Table 1.doc

1	1	21.2164	266.125	66	1.29261	0.63032	0.02703	0.53112	0.72723	3.612	35.9063	57	44
2	1	24.0232	432.566	129	1.42446	0.49123	12.4019	0.521425	0.61518	2872	20.0155	30	23
3	1	35.4435	114.293	140	1.45781	0.72152	12.3512	0.50101	0.77167	5011	20.0071	31	25
4	1	39.3772	231.376	206	1.4679	0.79163	16.1952	0.523663	0.877432	9011	20.0071	32	26
5	1	38.3581	228.443	178	1.46005	0.331708	15.0315	0.521013	0.761111	9185	51.4539	32	38
6	1	39.3567	41.37	150	1.17173	0.327089	12.8198	0.521013	0.765206	4709	31.3933	33	38
7	1	38.3765	242.308	85	1.17148	0.421263	10.4031	0.494337	0.760132	2860	46.4223	33	38
8	1	45.323	21.1377	239	1.40448	0.702172	12.3204	0.555951	0.842474	4279	30.7462	32	33
9	1	37.3216	201.969	197	1.77115	0.425478	15.4304	0.511195	0.67857	4110	21.3819	24	18
10	1	55.4904	42.3214	166	1.41539	0.781573	12.4313	0.521051	0.653238	4298	28.4384	30	22
11	1	55.3416	176.373	124	1.31687	0.450110	11.4166	0.5	0.621309	4098	31.5228	33	33
12	1	65.4193	170.466	274	2.47252	0.219223	20.3108	0.73905	0.492142	16200	50	52	39
13	1	61.9115	232.138	195	1.45329	0.781019	15.1357	0.519811	0.711216	8025	46.2821	48	35
14	1	61.9115	214.022	160	1.7325	0.825678	15.1310	0.503232	0.714286	8730	48.5	50	37
15	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
16	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
17	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
18	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
19	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
20	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
21	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
22	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
23	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
24	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
25	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
26	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
27	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
28	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
29	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
30	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
31	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
32	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
33	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
34	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
35	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
36	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
37	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
38	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
39	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
40	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
41	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
42	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
43	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
44	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
45	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
46	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
47	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
48	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
49	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
50	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
51	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
52	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
53	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
54	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
55	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
56	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
57	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
58	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
59	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
60	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
61	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
62	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
63	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
64	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
65	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
66	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
67	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
68	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
69	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
70	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
71	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
72	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
73	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
74	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
75	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
76	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
77	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
78	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
79	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
80	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
81	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
82	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
83	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
84	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
85	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9	74	56
86	1	77.3137	232.032	43	1.4258	0.716165	7.9454	0.519811	0.714286	8730	74.9		

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EV Table 2.doc

Example of the summary output of AnalysedNA.m program
(summary for 10 3 by 3 montage images)

1	1107	153.912	79.3918	1.58809	0.398933	0.724441	0.137986	14.0012	3.315	0.303227	0.0350163	0.701248	0.035176	549.26	3186.35	01.539	18.332	02.3016	18.9393
2	1108	16.324	50.3908	22.4584	0.398942	0.727933	0.138402	14.2034	3.43288	0.304571	0.0311485	0.70177	0.0320218	6786.37	3895.1	02.2116	17.0945	03.0273	17.303
3	1109	15.0245	13.3382	31.3501	0.397933	0.727552	0.137845	14.0853	3.33904	0.305267	0.0348822	0.702591	0.0310005	6881.08	3335.81	01.7167	20.5916	05.3838	21.2287
4	1110	16.2032	54.4017	25.1765	0.389682	0.727142	0.140518	14.3331	3.61812	0.302766	0.0374234	0.695945	0.0333889	6997.68	0212.87	03.1798	19.4582	07.2561	20.5112
5	1111	13.5902	52.505	24.193	0.400493	0.721084	0.141704	14.318	3.61045	0.304132	0.0372988	0.70073	0.0336884	7050.22	4163.04	01.8559	21.4761	05.0322	21.1282
6	1112	16.5359	51.5367	24.3388	0.425512	0.728114	0.137721	14.0974	3.31686	0.304616	0.0342731	0.696204	0.0382855	6843.2	3974.12	04.286	19.3386	05.3105	19.4724
7	1113	15.112	84.4886	1.40512	0.451022	0.694813	0.185667	11.721	5.27441	0.83311	0.0481729	0.704526	0.0392393	5182.51	0302.11	24.9743	21.1569	26.16	21.0024
8	1114	17.0121	43.0619	25.1082	0.403167	0.727123	0.137559	14.3133	3.60593	0.306384	0.0357179	0.703574	0.0351602	6165.87	3787.46	02.2752	17.3133	03.9781	17.7821
9	1115	14.53	51.3961	20.356	0.404986	0.7268	0.138518	14.1402	3.47794	0.305208	0.0356298	0.700331	0.0366773	6576.34	4022.38	01.4066	17.4991	03.143	16.2488
10	1116	13.4606	82.618	1.51286	0.400372	0.717117	0.139015	13.8185	3.49536	0.804875	0.0378111	0.703759	0.03572	6587.18	3788.39	04.2141	20.7691	05.0765	20.125
11	1117	14.3182	53.7818	25.4084															

CLAIMS

What is claimed is:

1. A method of predicting a property of a manipulation of cells based
5 upon a descriptor, said method comprising:
 providing a plurality of cells;
 manipulating said plurality of cells;
 capturing a morphological value from said plurality of cells;
 assigning a degree of presence of said morphological value; and
10 storing said morphological value and said degree of presence;
 wherein said descriptor is derived from a first component of a cell and
a second component of said cell, said capturing said morphometric value from said
plurality of cells comprises determining a relationship between said first component
and said second component.
- 15 2. The method of claim 1 wherein said first component and said second
component are selected from a protein, a protein modification, a nucleic acid, a lipid,
a carbohydrate, a subcellular structure and an organelle.
3. The method of 1 wherein said step of manipulation occurs in a manner
selected from a electrical source, a chemical source, a thermal source, a gravitational
20 source, a nuclear source, a temporal source, and a biological source
4. The method of claim 3 wherein said chemical source is selected from a
pharmacological candidate and a drug screening library.
5. The method of claim 1 wherein said morphological value is selected
from a count, an area, a perimeter, a length, a breadth, a fiber length, a fiber breadth, a
25 shape factor, a elliptical form factor, an inner radius, an outer radius, a mean radius,
an equivalent radius, an equivalent sphere volume, an equivalent prolate volume, an
equivalent oblate volume, an equivalent sphere surface area, an average gray value, a
total gray value, and an optical density.
6. The method of claim 1 wherein said degree of presence is
30 multiple of a quantized value.

7. A computer program product for populating a database with manipulated biological information, said computer program product comprising:
- code for providing a plurality of cells in various stages of the cell cycle, said stages of the cell cycle including at least one selected from interphase, G0 phase, G1 phase, S phase, G2 phase, M phase, prophase, prometaphase, metaphase, anaphase, and telophase;
 - code for manipulating said cells in said various stages of cell cycle development to form a plurality of manipulated cells;
 - code for capturing an image of said plurality of manipulated cells;
 - code for determining a descriptor from said image for said manipulated cells;
 - code for populating a database with said descriptor;
 - wherein said image includes a first component of a cell and a second component of said cell; and
 - a computer readable storage medium for holding the codes.
8. The computer program product of claim 7 wherein said first component and said second component are selected from a protein, a protein modification, a nucleic acid, a lipid, a carbohydrate, a sub-cellular structure and an organelle.
9. The computer program product of claim 7 wherein said image is a digitized representation of said plurality of manipulated cells.
11. The computer program product of claim 9 wherein said digitized representation provides a density value of said plurality of manipulated cells.
11. The computer program product of claim 7 wherein said descriptors comprise numeric or logical values.
12. The computer program product of claim 11 wherein said values further comprises a nucleotide.
13. The computer program product of claim 11 wherein said values further comprises an amino acid letter.
14. A system for capturing images of cells or cell structures, the system comprising:
- a cell holder comprising a plurality of sites in a spatial orientation, each of the sites being capable of holding a plurality of cells to be imaged;

an image capturing device coupled to the cell holder, the image capture device being adapted to capture at least one image in at least one of the plurality of sites;

5 an illumination apparatus comprising a liquid light guide coupled to the plate for highlighting the plurality of cells in a relatively even spatial manner for image capturing purposes;

an image processing device coupled to the image capturing device, the image capturing device being adapted to convert the image into a digital representation; and

10 a database storage device comprising a database management element coupled to the image capturing device, the database storage device being adapted to retrieve the digital representation of the image from the image processing device and storing the digital representation.

15 15. The system of claim 14 further comprising a stage comprising a device for moving the cell holder in a spatial direction to traverse across the cell holder in the spatial orientation.

20 16. The system of claim 14 wherein the illumination apparatus comprises sub-elements, at least one of the sub-elements being positioned away from the image capturing device to prevent a possibility of vibration from the one sub-elements to be transmitted to the image capturing device.

17. The system of claim 14 wherein the digital representation comprises a plurality of regions and objects.

18. The system of claim 14 further comprising a computing device connected between the database storage device and the image processing device.

25 19. The system of claim 14 wherein the image capturing device comprises a magnification of at least 1X and greater to capture the image of the site.

20. The system of claim 14 wherein the plurality of sites comprises at least 96 sites.

30 21. The system of claim 14 wherein the liquid light guide characterized as a flexible member that substantially prevents vibration from the an element of the illumination apparatus to be transferred to the image capturing device.

22. The system of claim 14 wherein the spatial direction can be selected from an x-direction, a y-direction, or a z-direction in a Cartesian coordinate system.

23. The system of claim 14 wherein the each of the sites comprises
5 a volume that is sufficient to prevent a solution therein from evaporating in a substantial manner that may influence the image capturing.

24. A method for identifying a mechanism of action for a first compound, the method comprising the steps of:
receiving the first compound;
10 measuring at least one feature of a cellular phenotype to define a target phenotype;
identifying additional compounds providing a feature similar to the feature identified in the measuring step; and
characterizing the first compound in terms of distance from a specific
15 target phenotype having known characteristics.

25. The method of claim 24 comprising the further step of storing the additional compounds and their associated phenotypes in a database.

26. A method for identifying a mechanism of action for a cellular stimulus, the method comprising the steps of:
20 receiving cells of interest;
measuring at least one feature of the cells to define and quantify a target phenotype;
identifying additional compounds providing a feature similar to the feature identified in the measuring step; and
25 characterizing the first compound in terms of distance from a specific target phenotype having known characteristics.

27. A method for identifying information relevant to at least one of a mechanism of action and cellular activity by utilizing assay data to elucidate a phenotype, the method comprising the steps of:
30 identifying a target protein;
identifying positive and negative biochemical hits related to the target protein;
defining the target phenotype utilizing the positive and negative hits;

identifying other compounds providing similar features; and
characterizing the first compound in terms of distance from a specific
target phenotype having known characteristics.

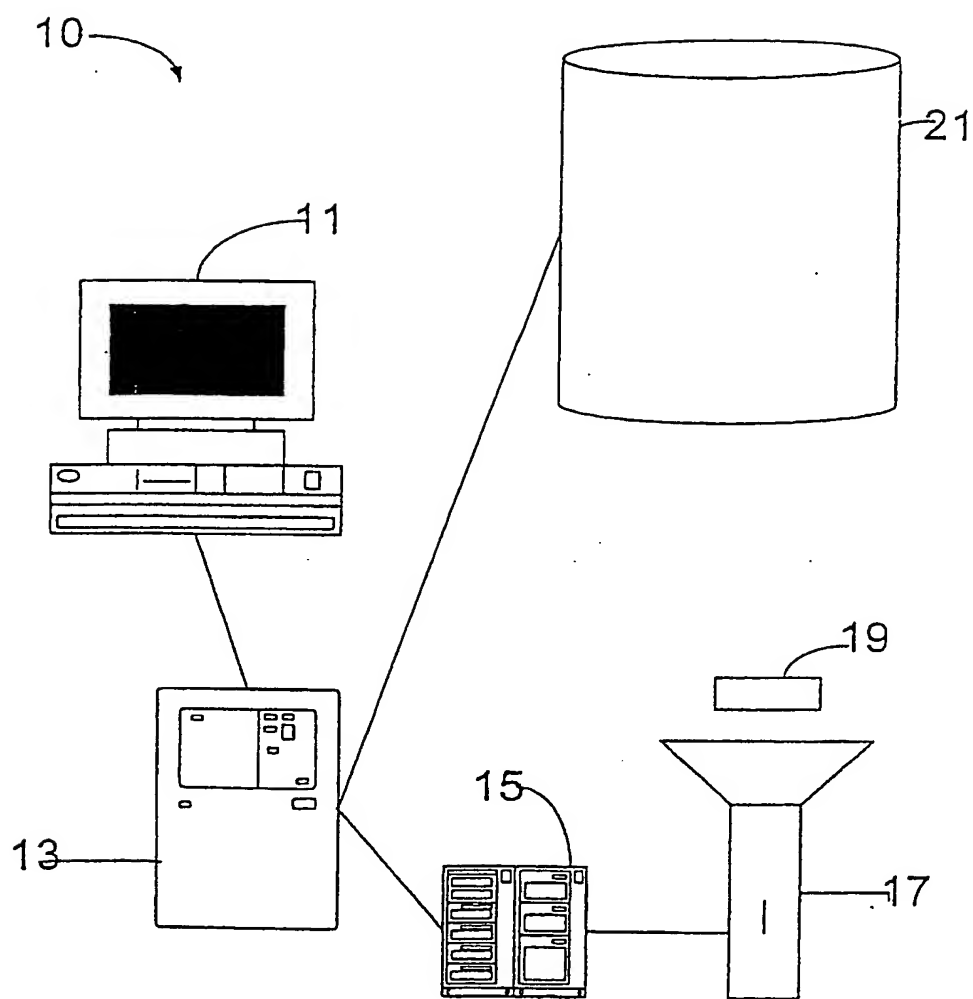


FIG. 1

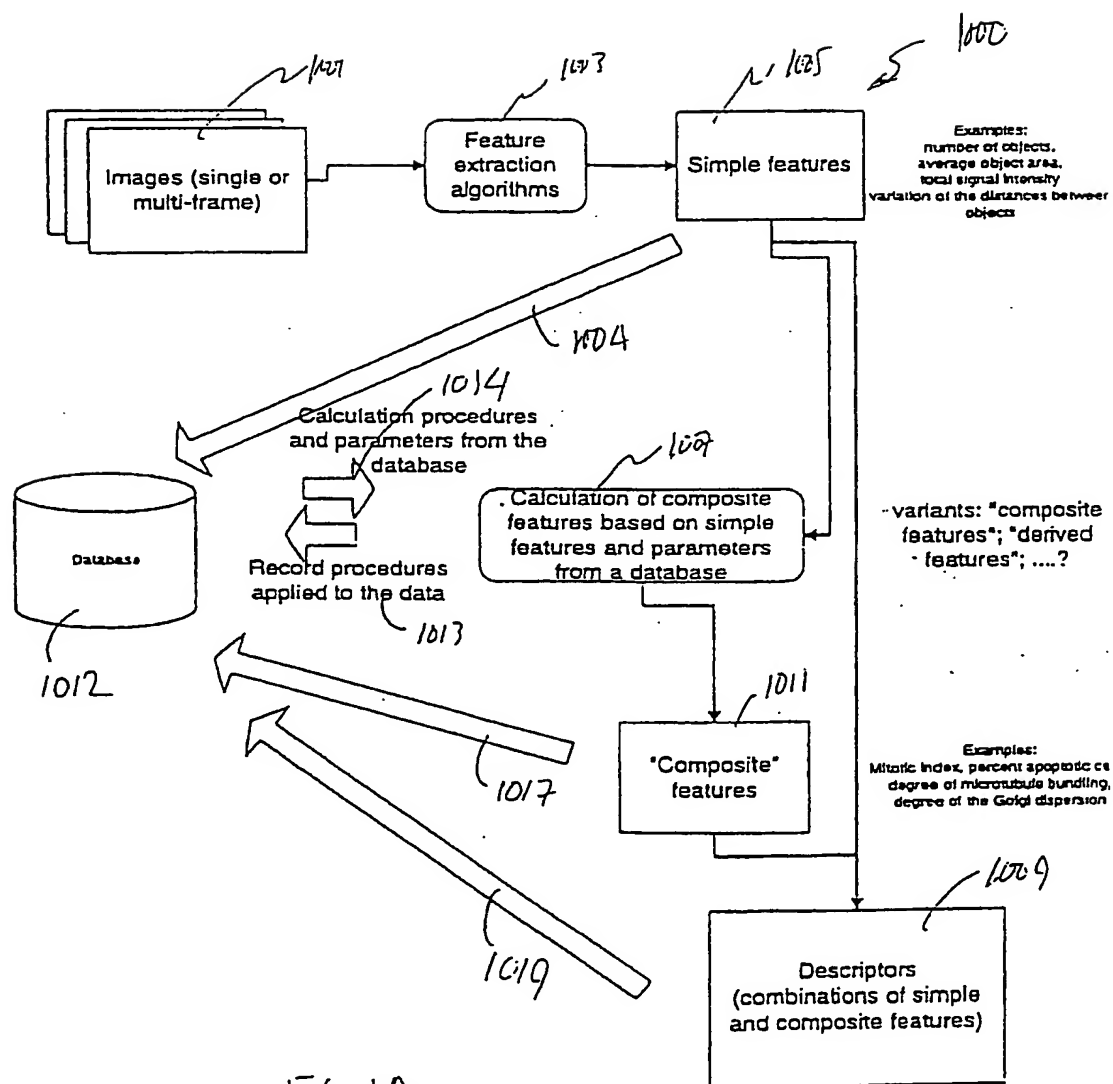
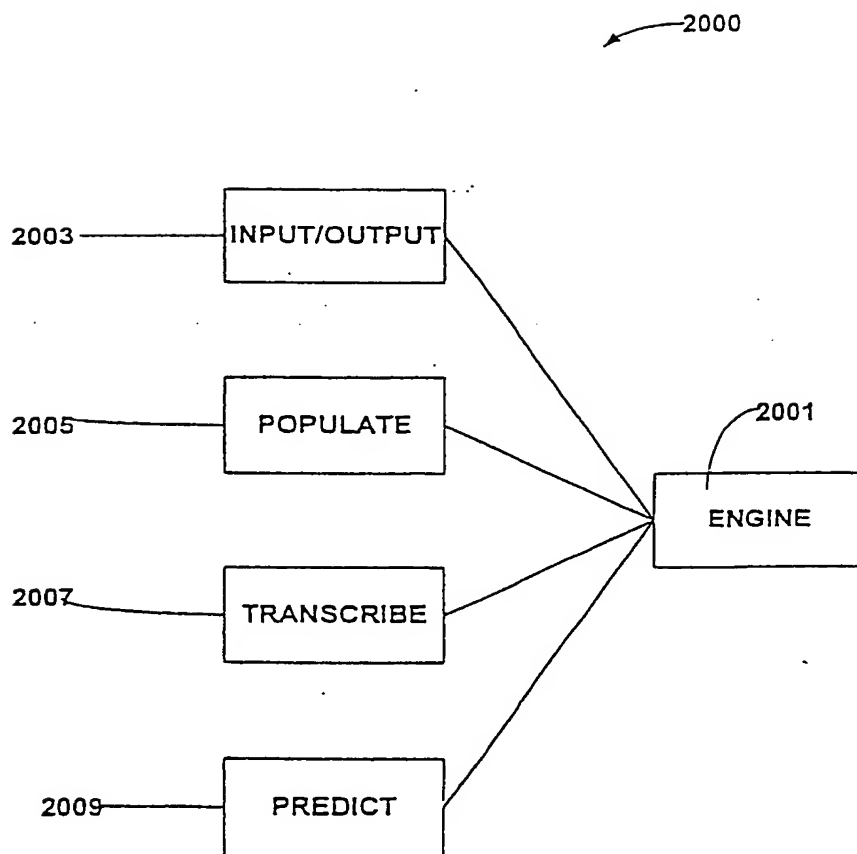


FIG. 1A

**FIG. 1B**

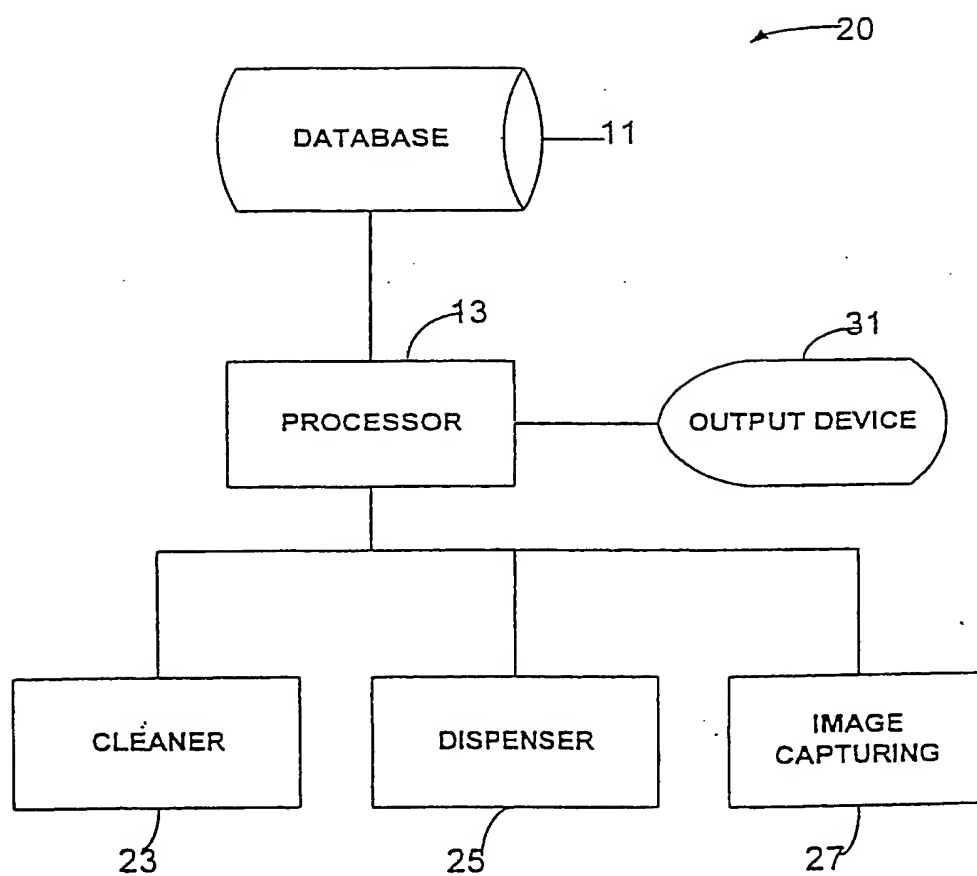
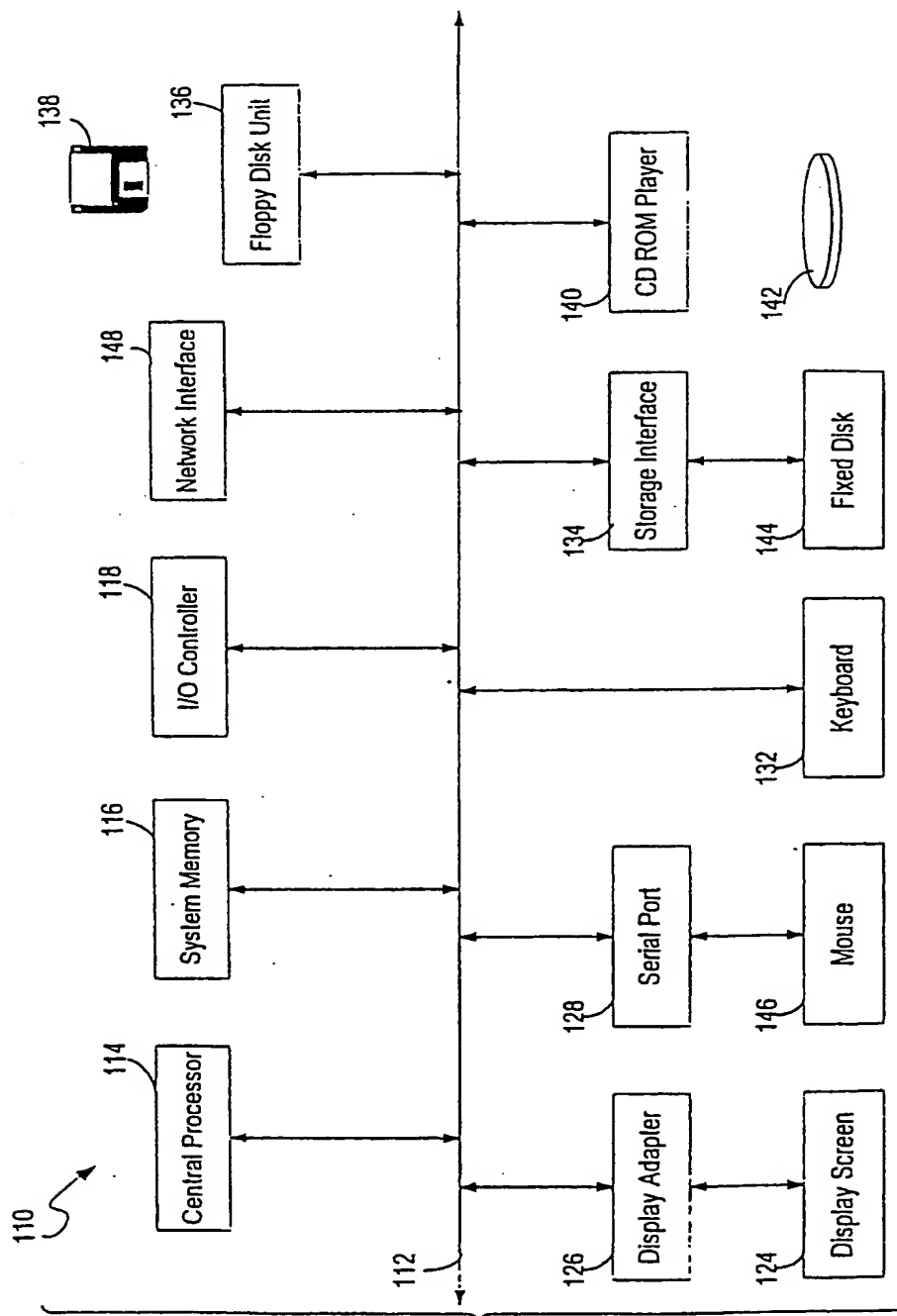


FIG. 2

*Fig 3*

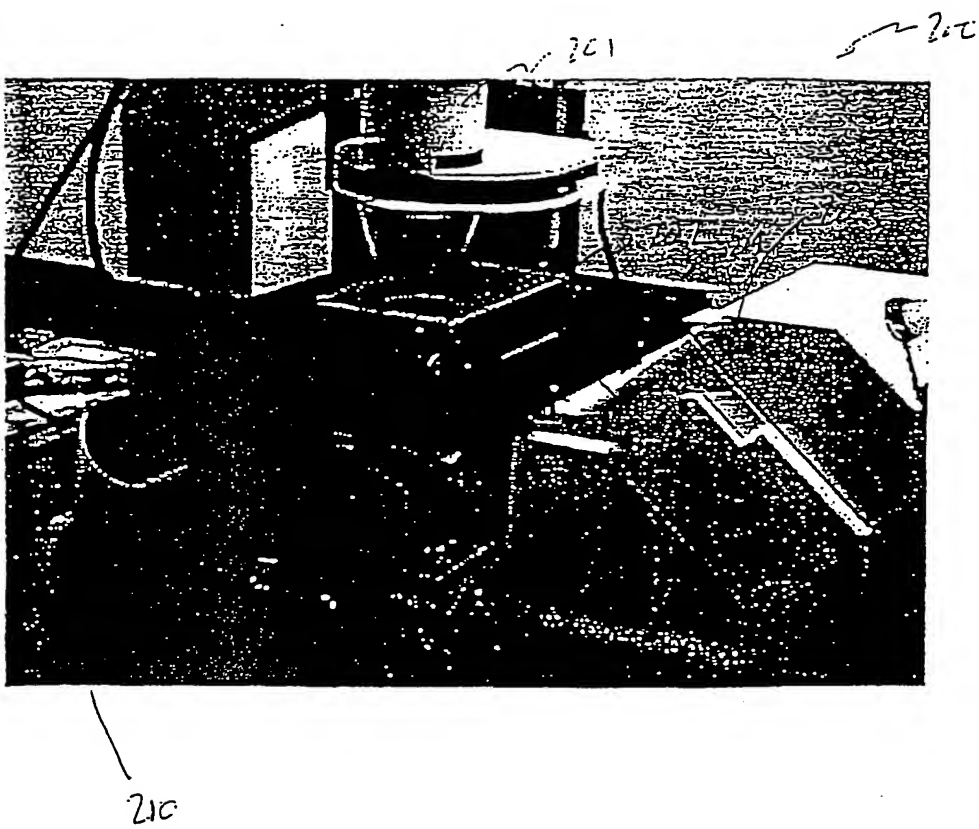


FIG. 4

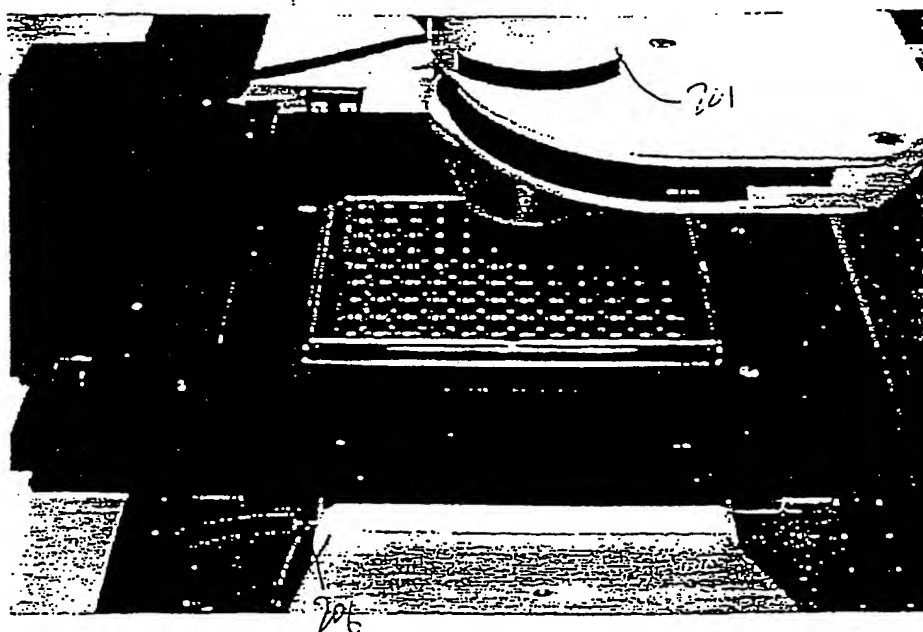


FIG 5

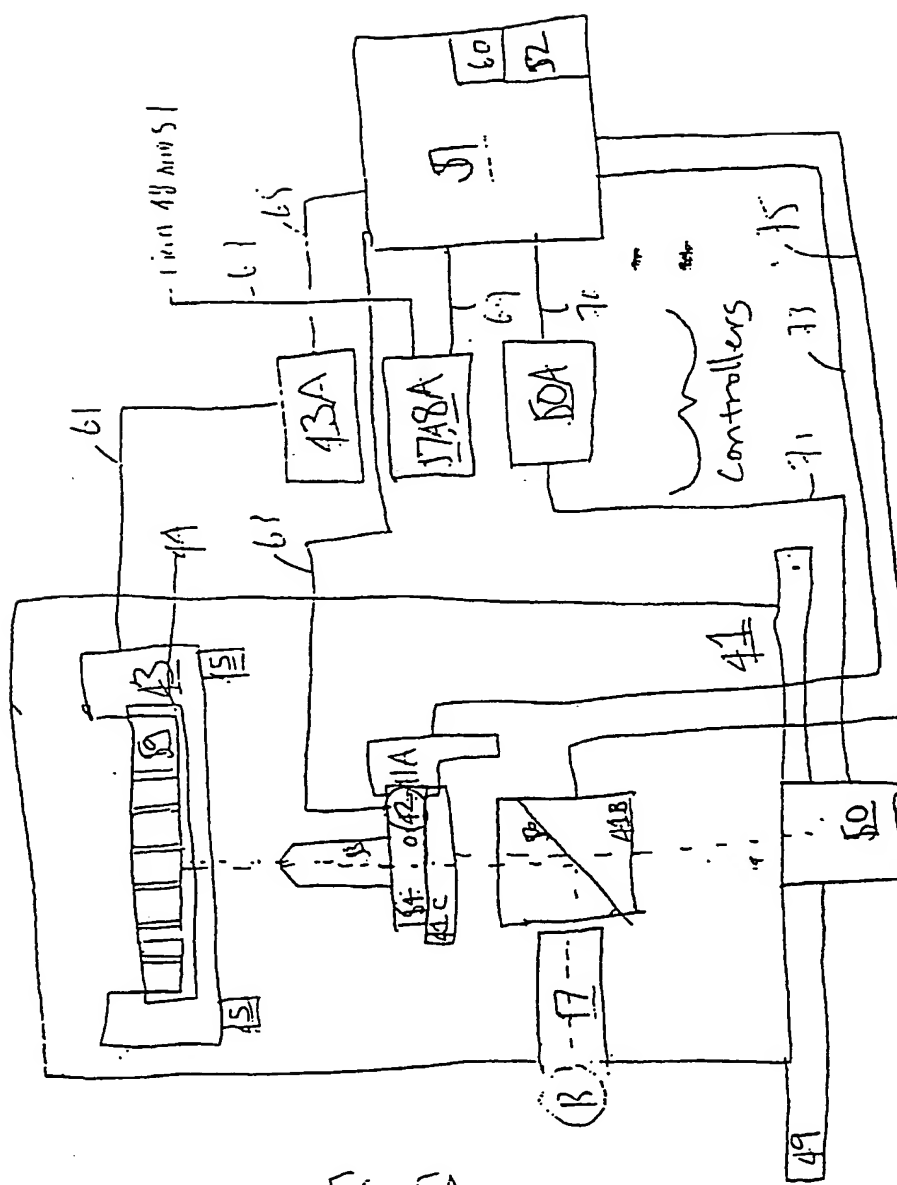


FIG. 5A

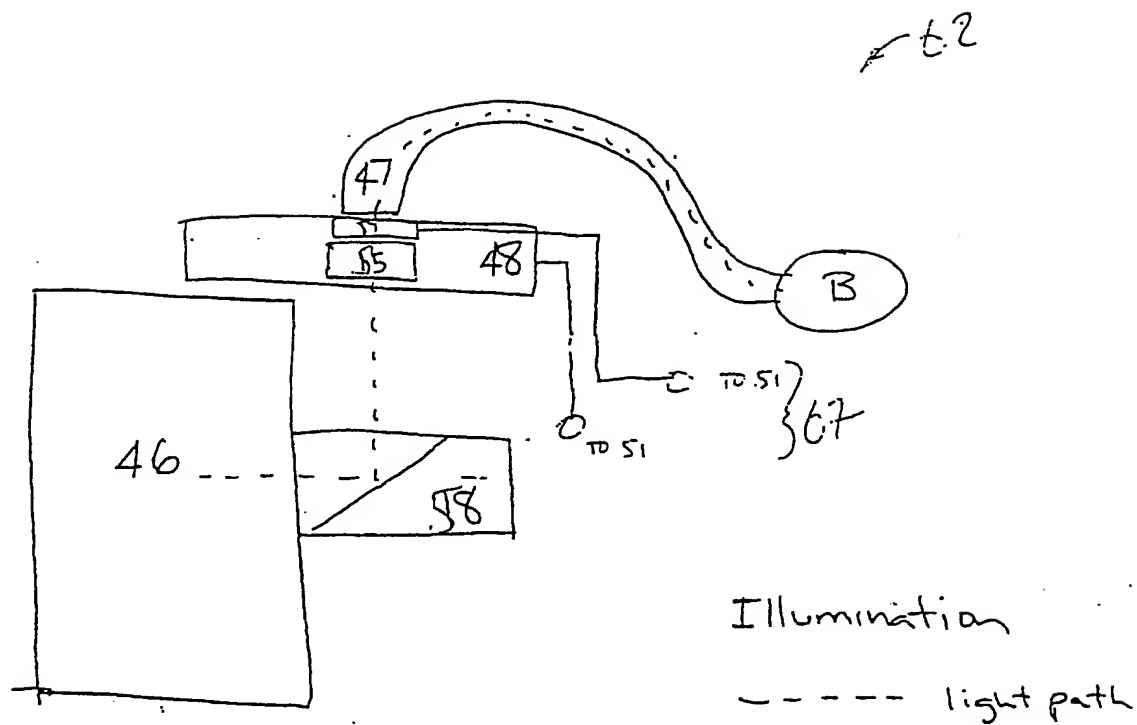


FIG. 5B

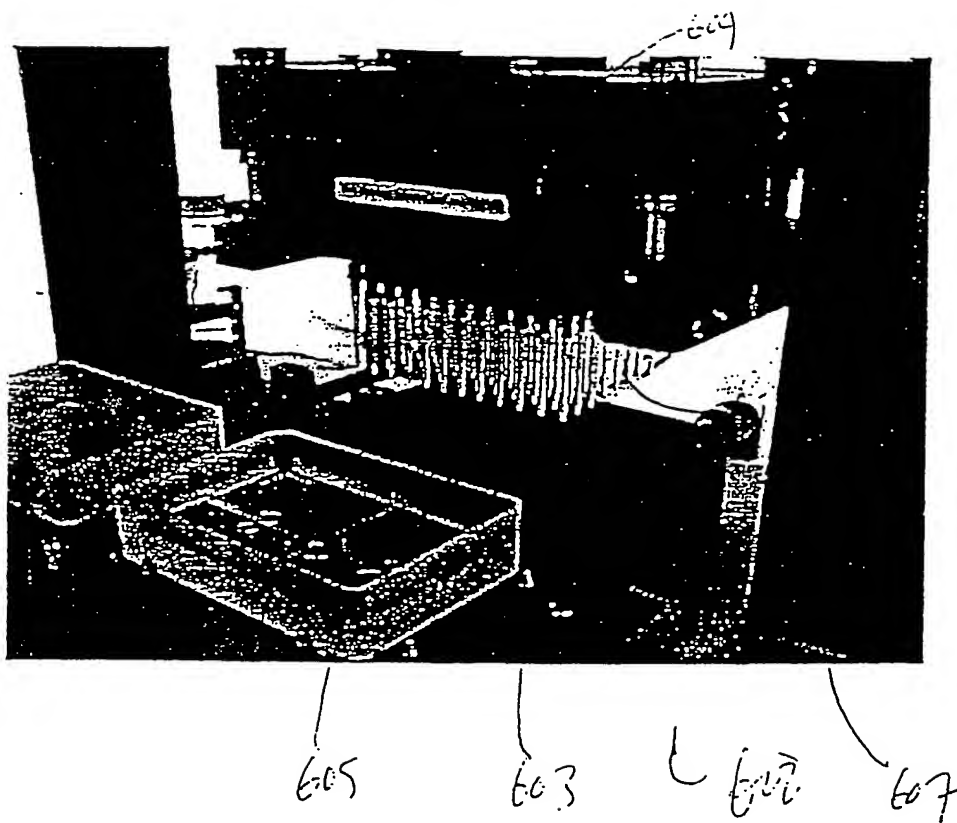


FIG. 6

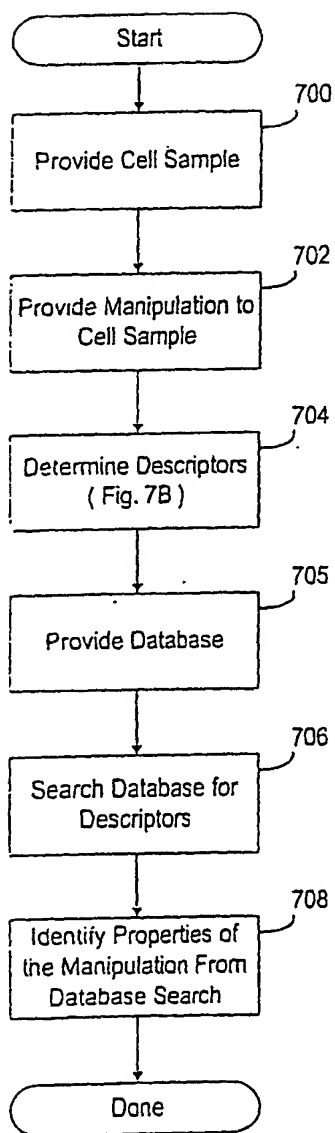


Fig. 7A

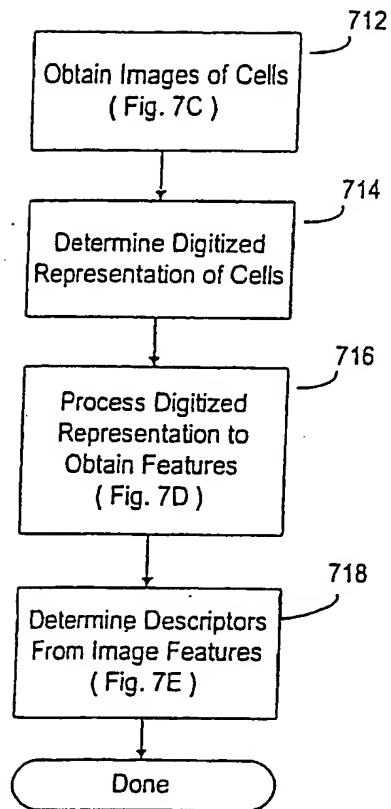


Fig. 7B
Step 704 of Fig. 7A

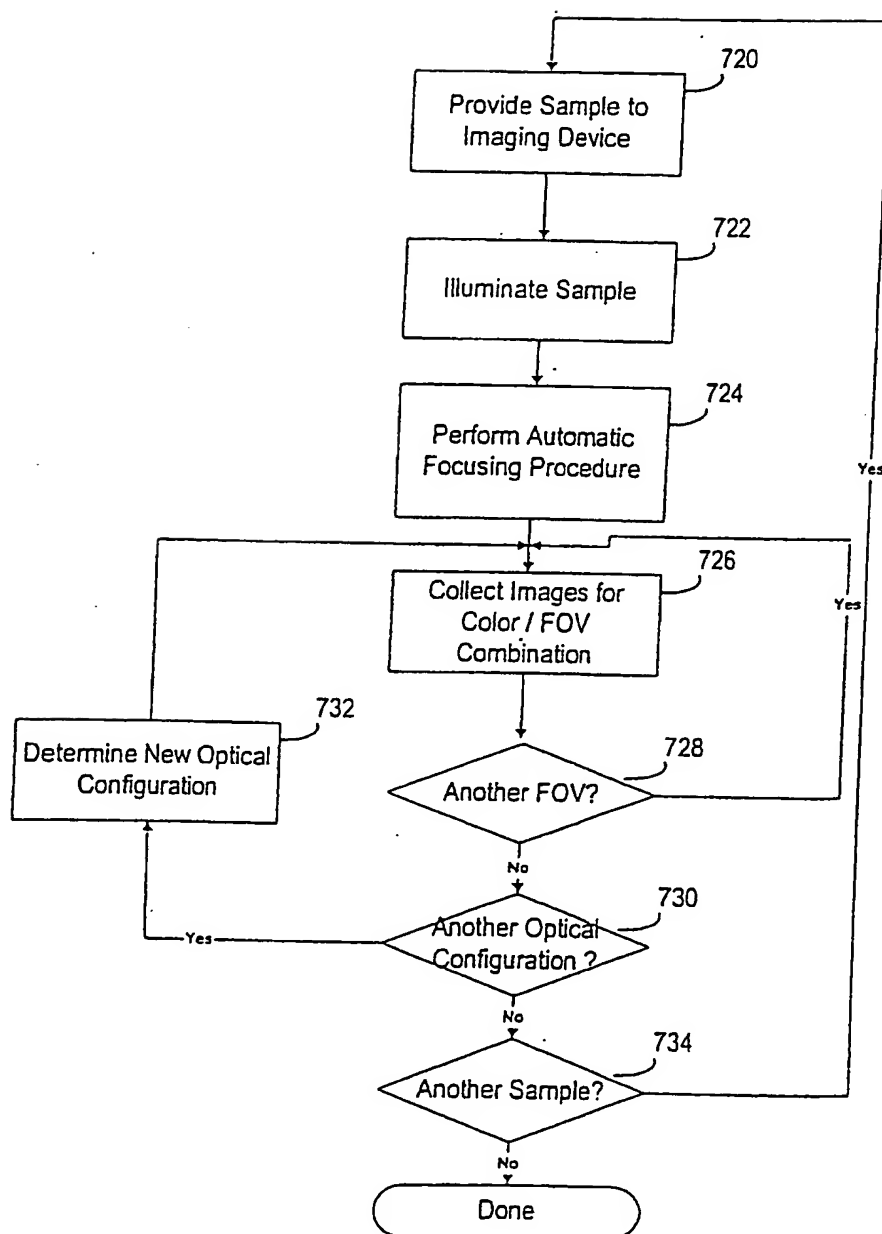


Fig. 7C
Step 714 of Fig. 7B

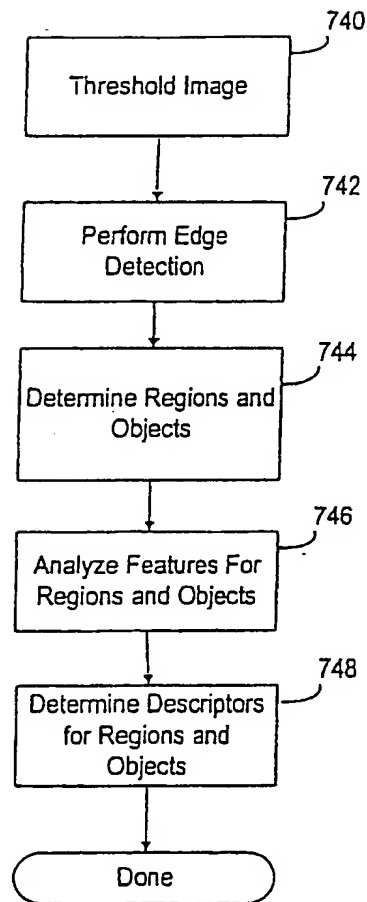


Fig. 7D
Step 716 of Fig. 7B

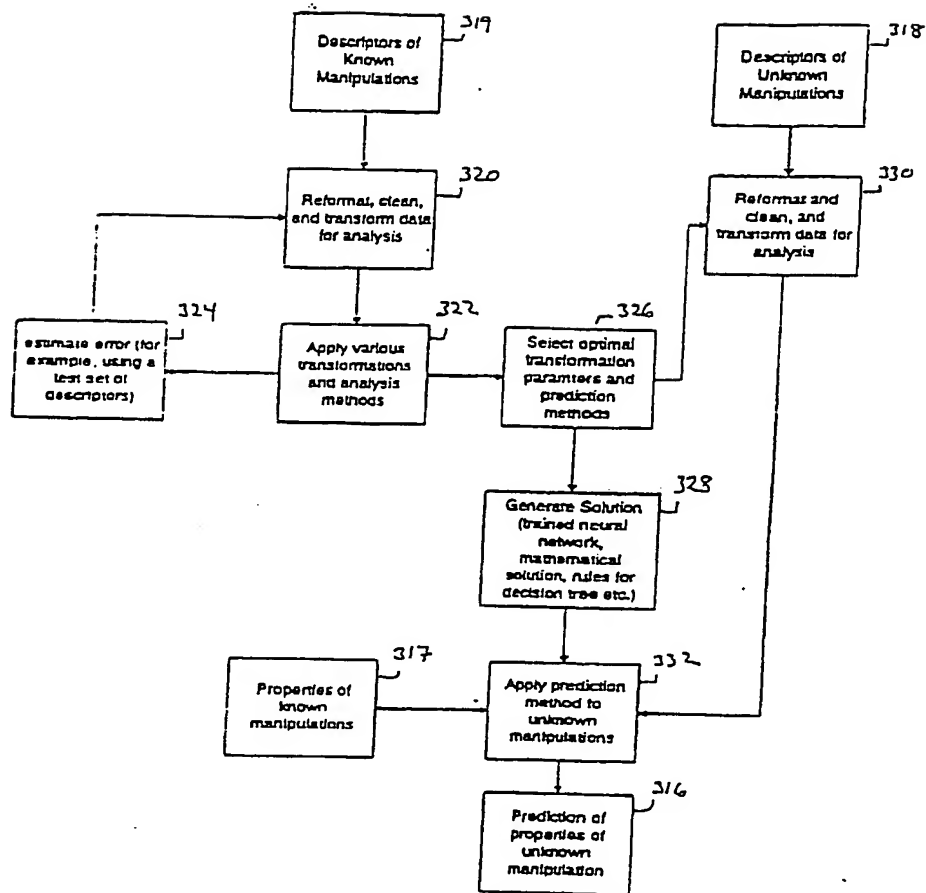


FIG. 7E

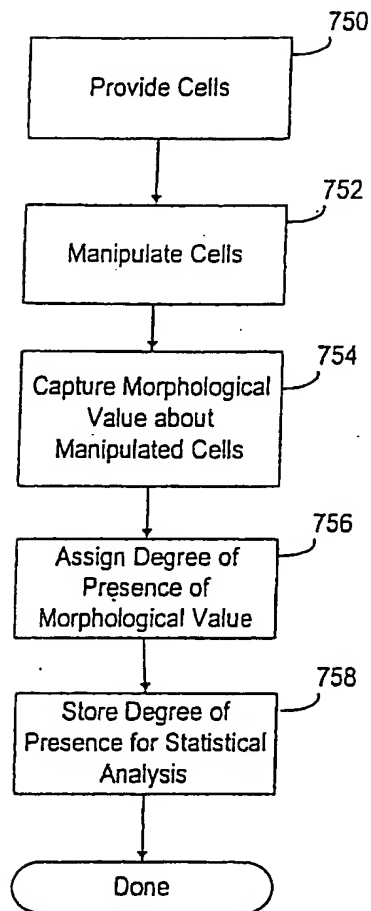


Fig. 7F

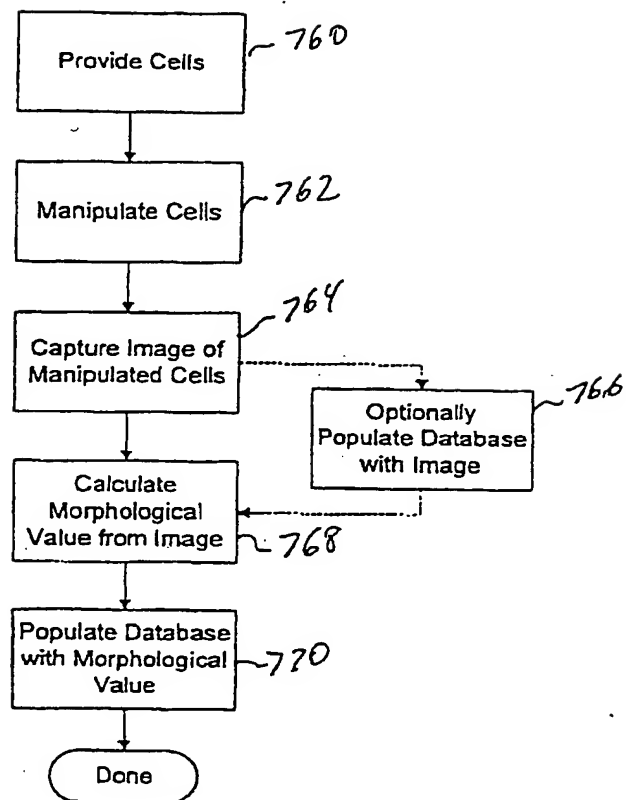


Fig 7G

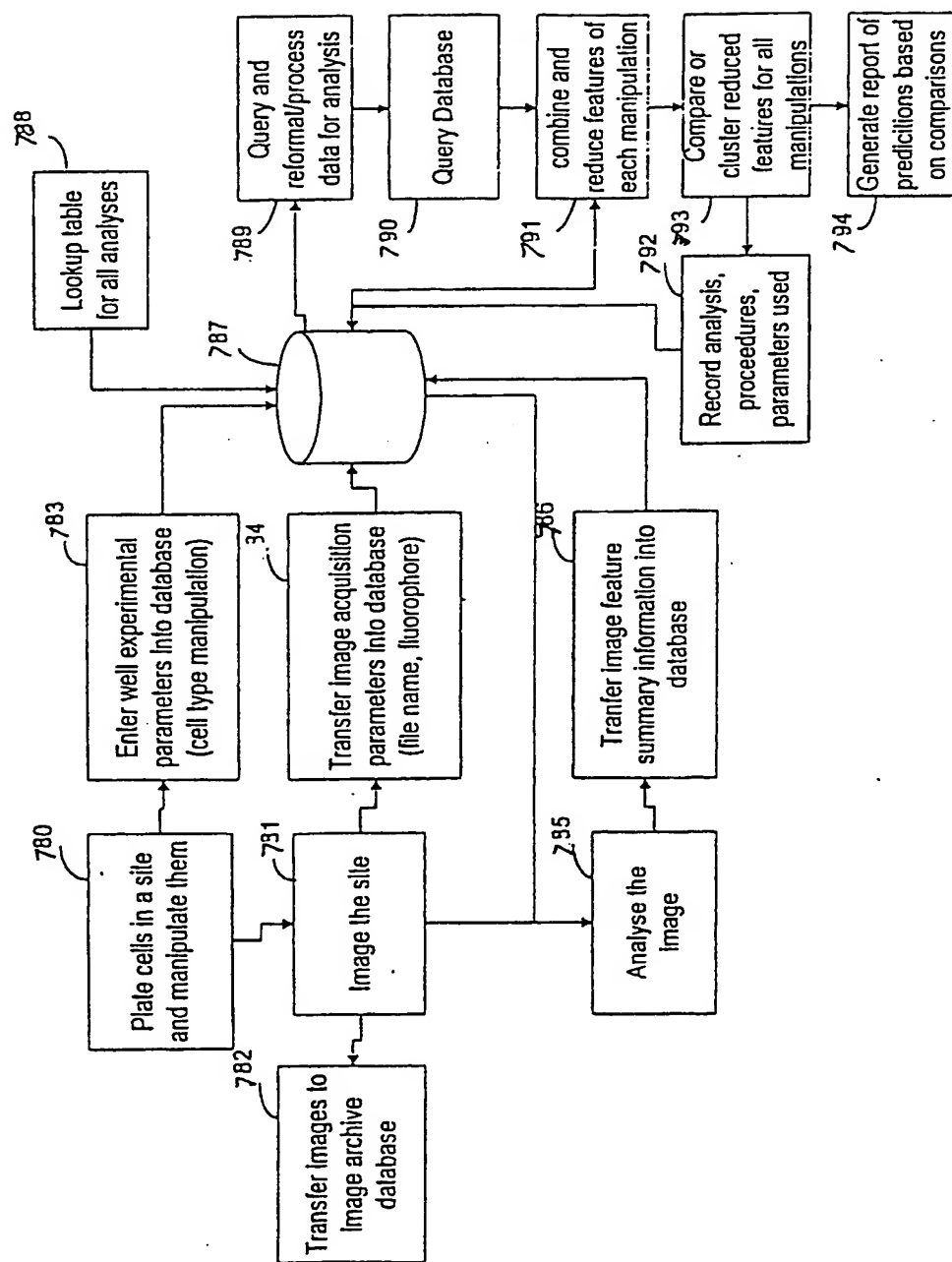


Fig. 7H

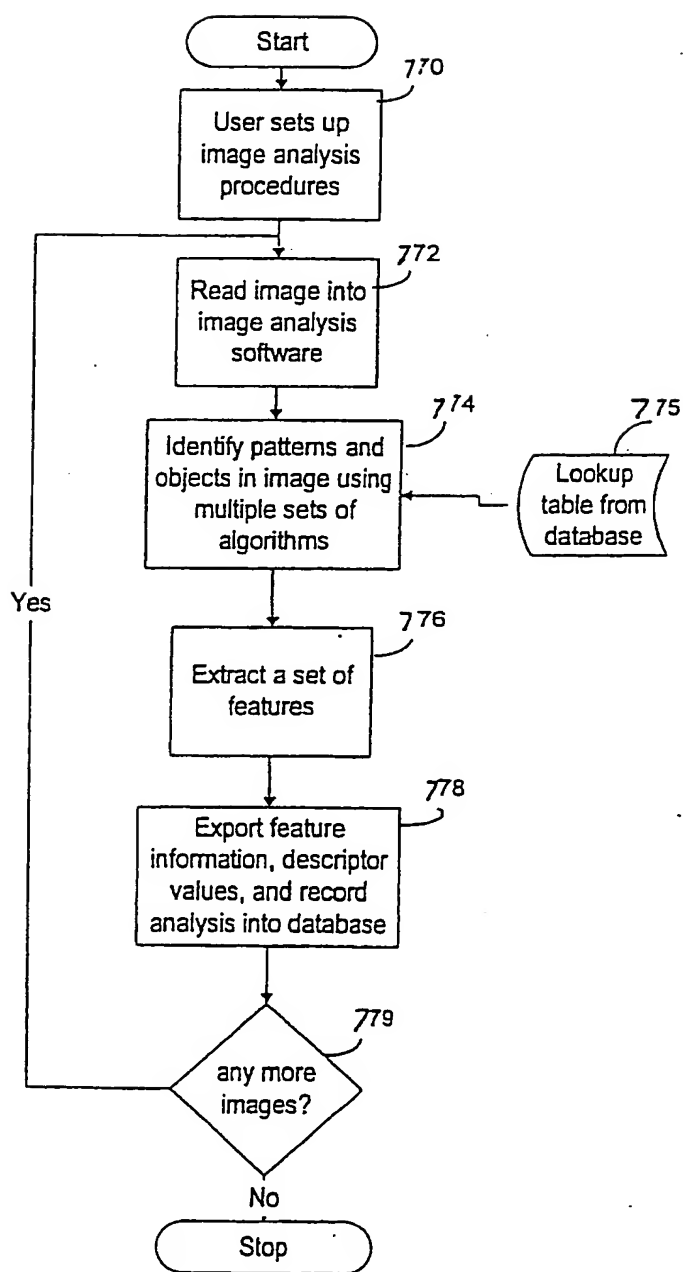


Fig. 71

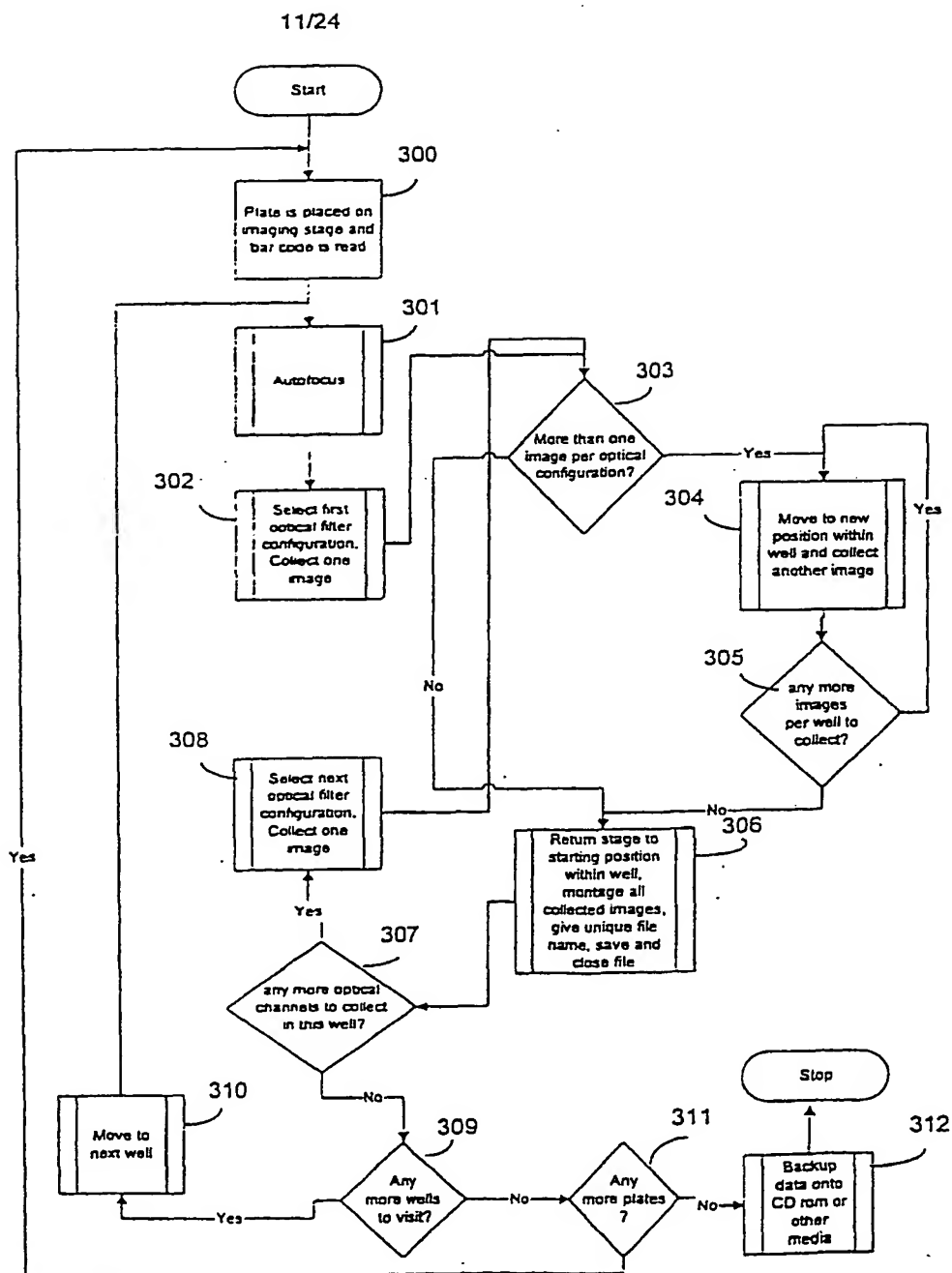


Fig. 7J

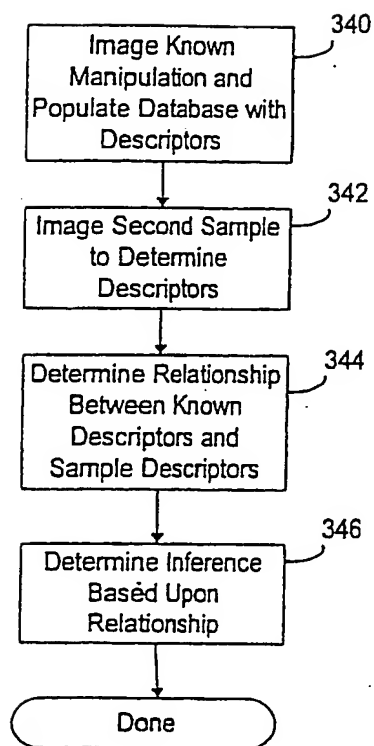


Fig. 7K

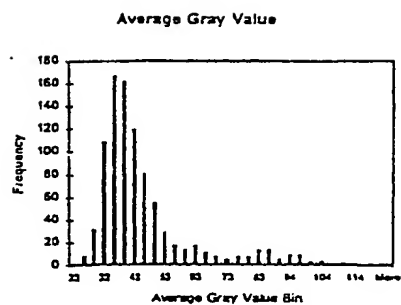


Fig. 8A

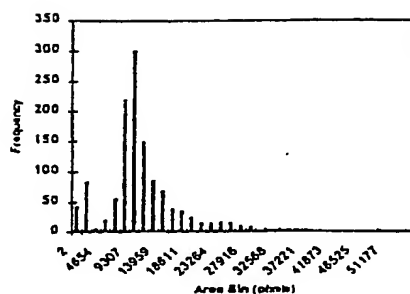


Fig. 8B

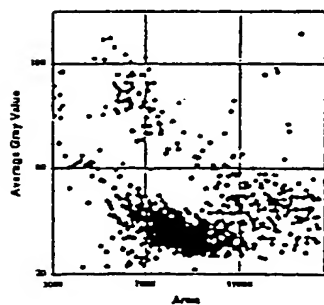


Fig. 8C

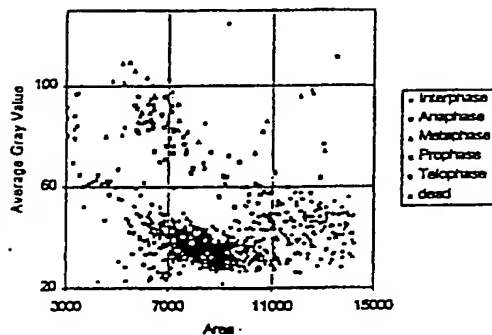


Fig. 8D

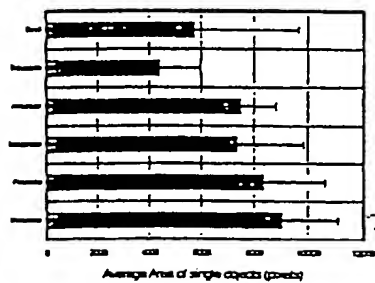


Fig. 8E

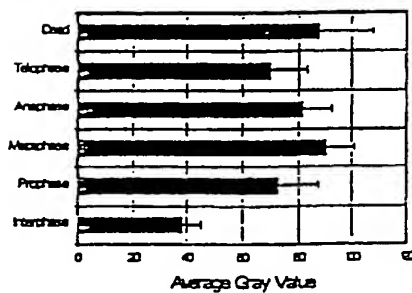


Fig. 8F

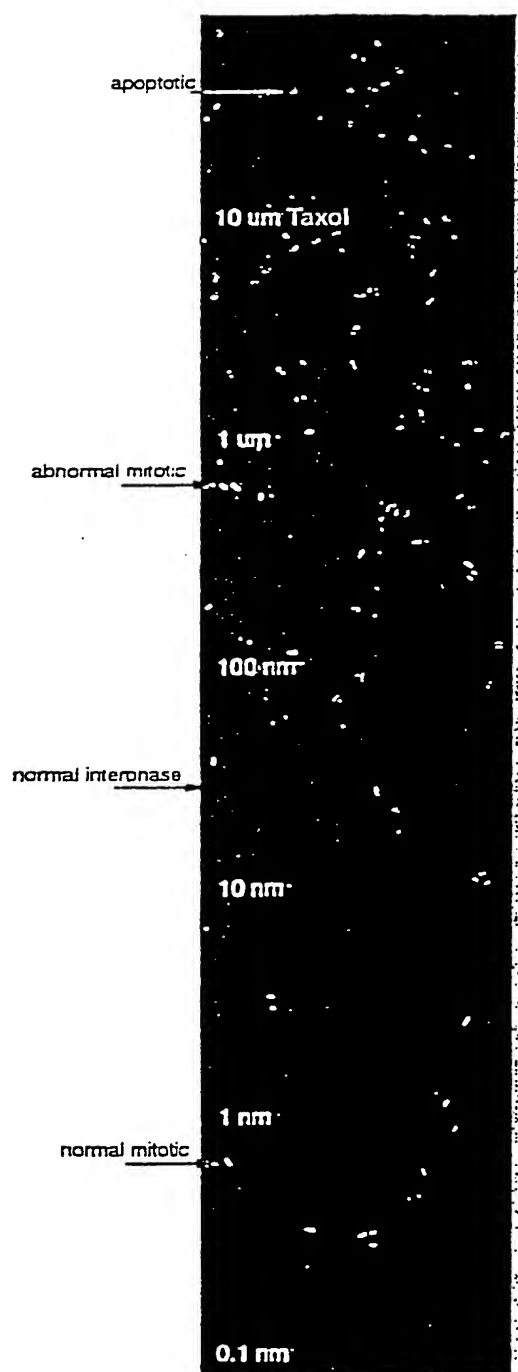


Fig. 9

MDCK cells treated with Taxol for 4.5 hours

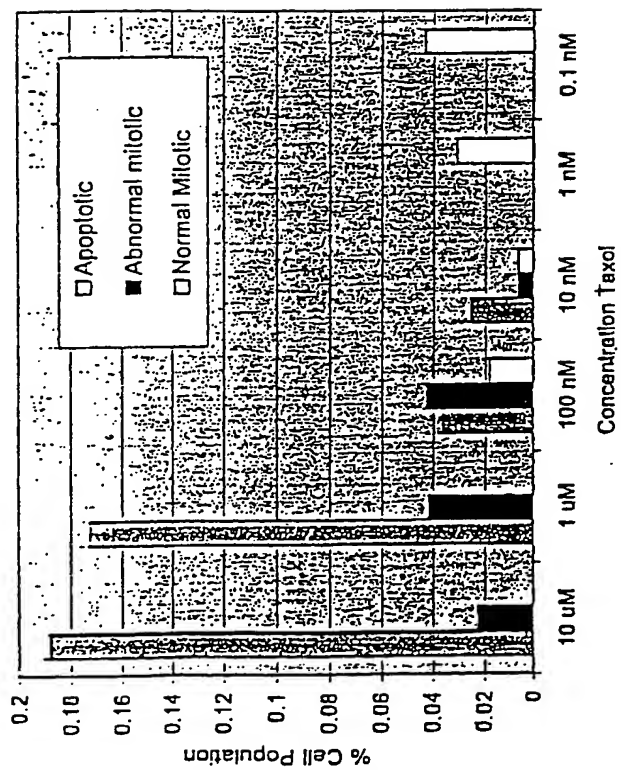
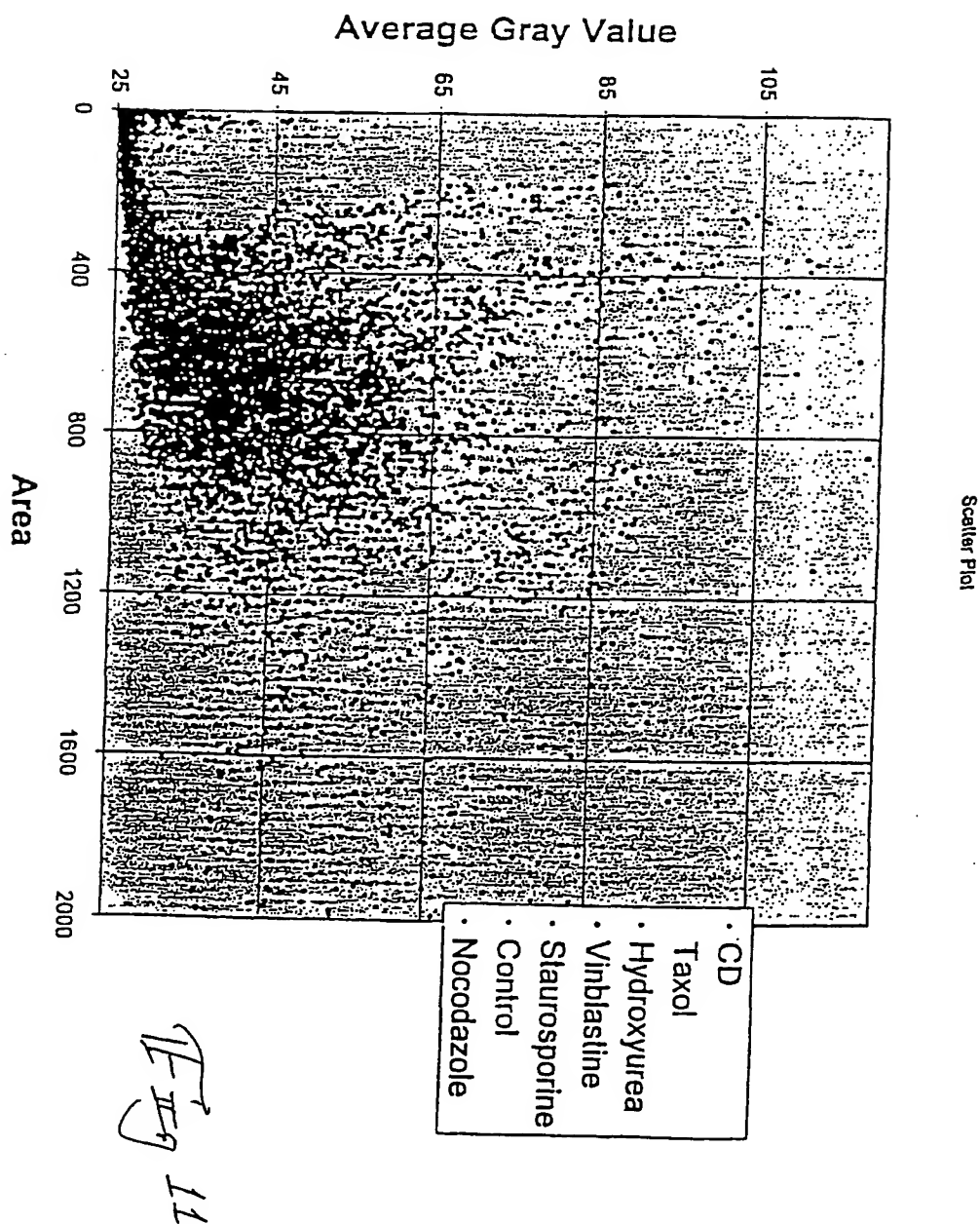
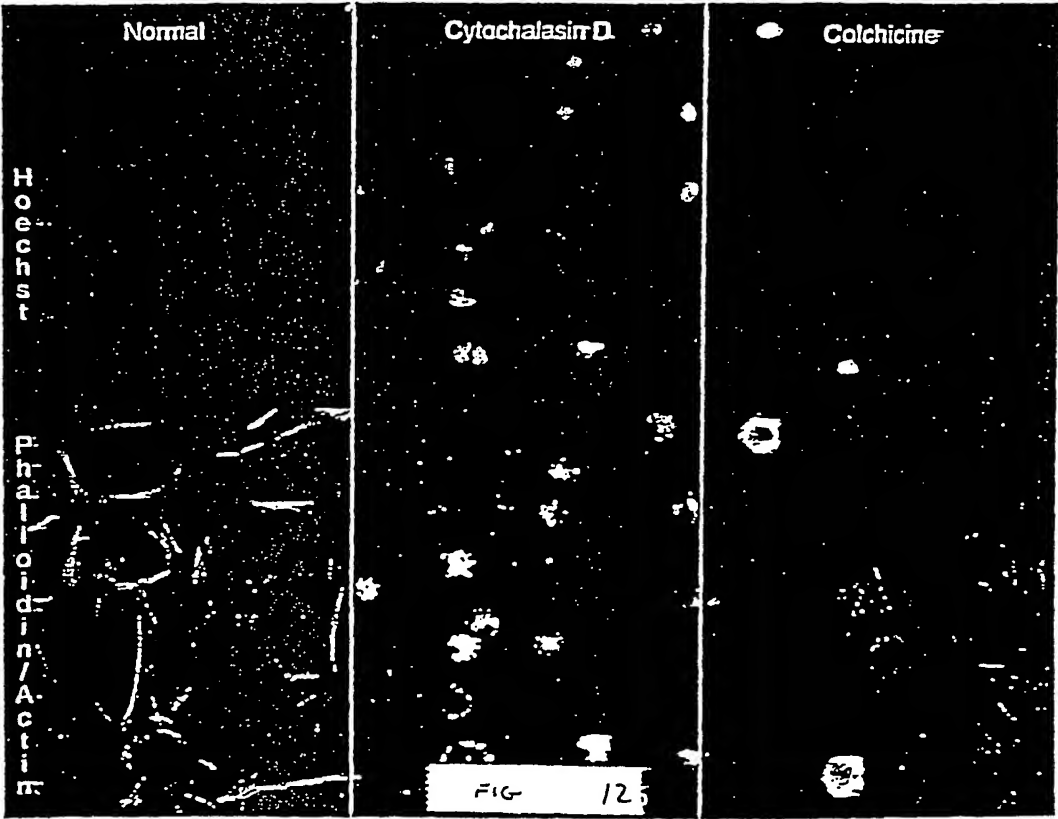


Fig. 10





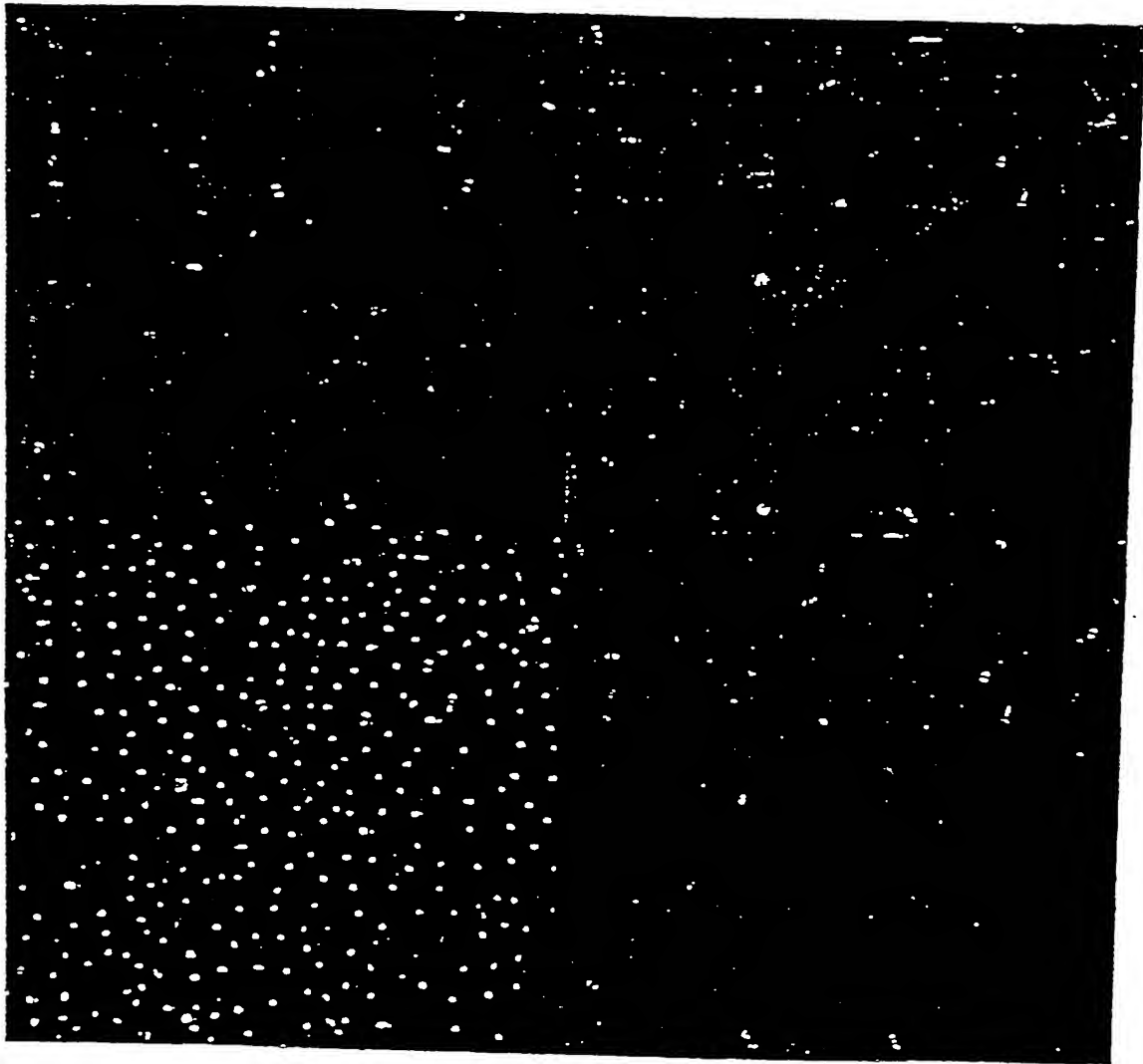


Fig 13

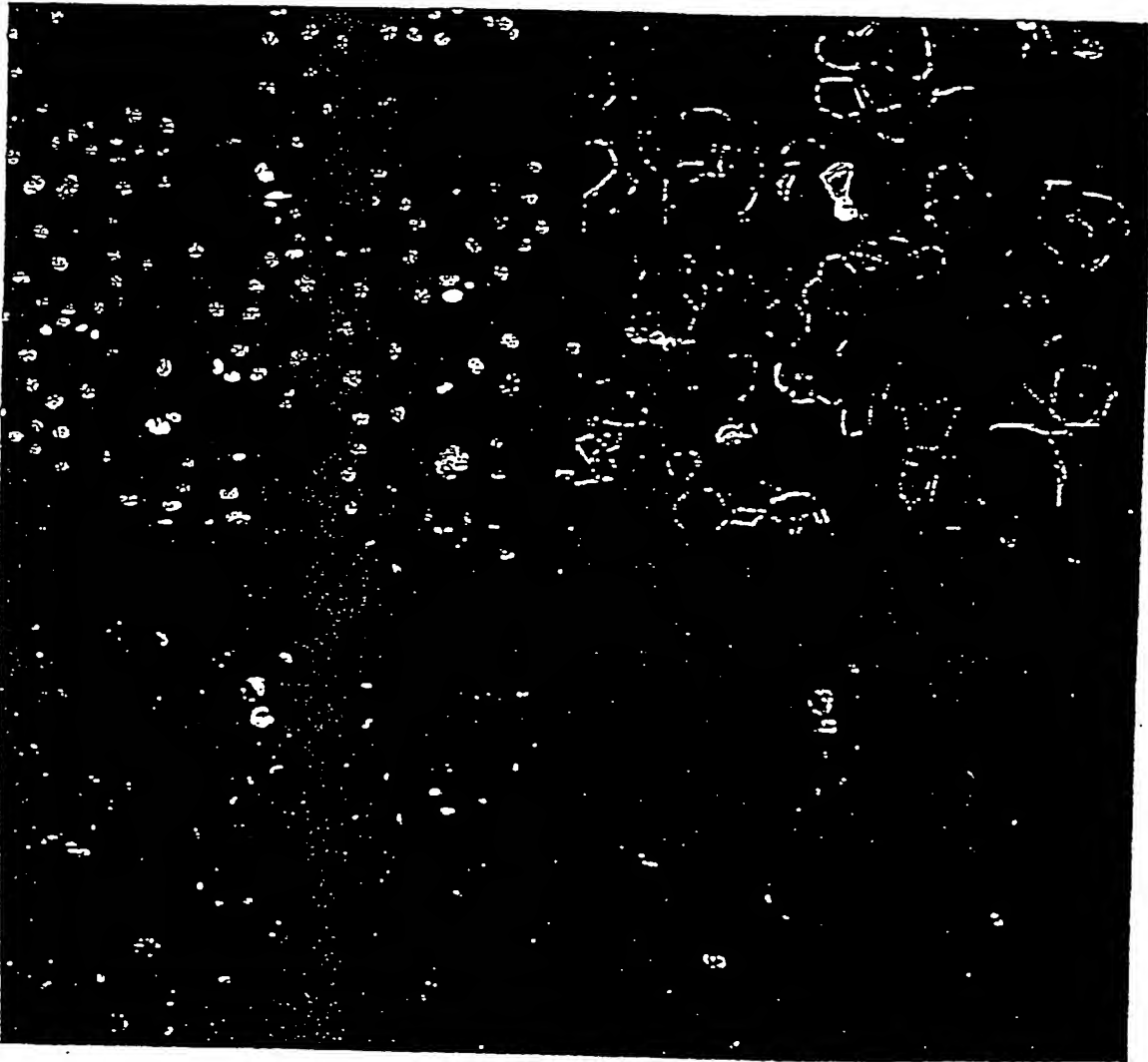


Fig 14

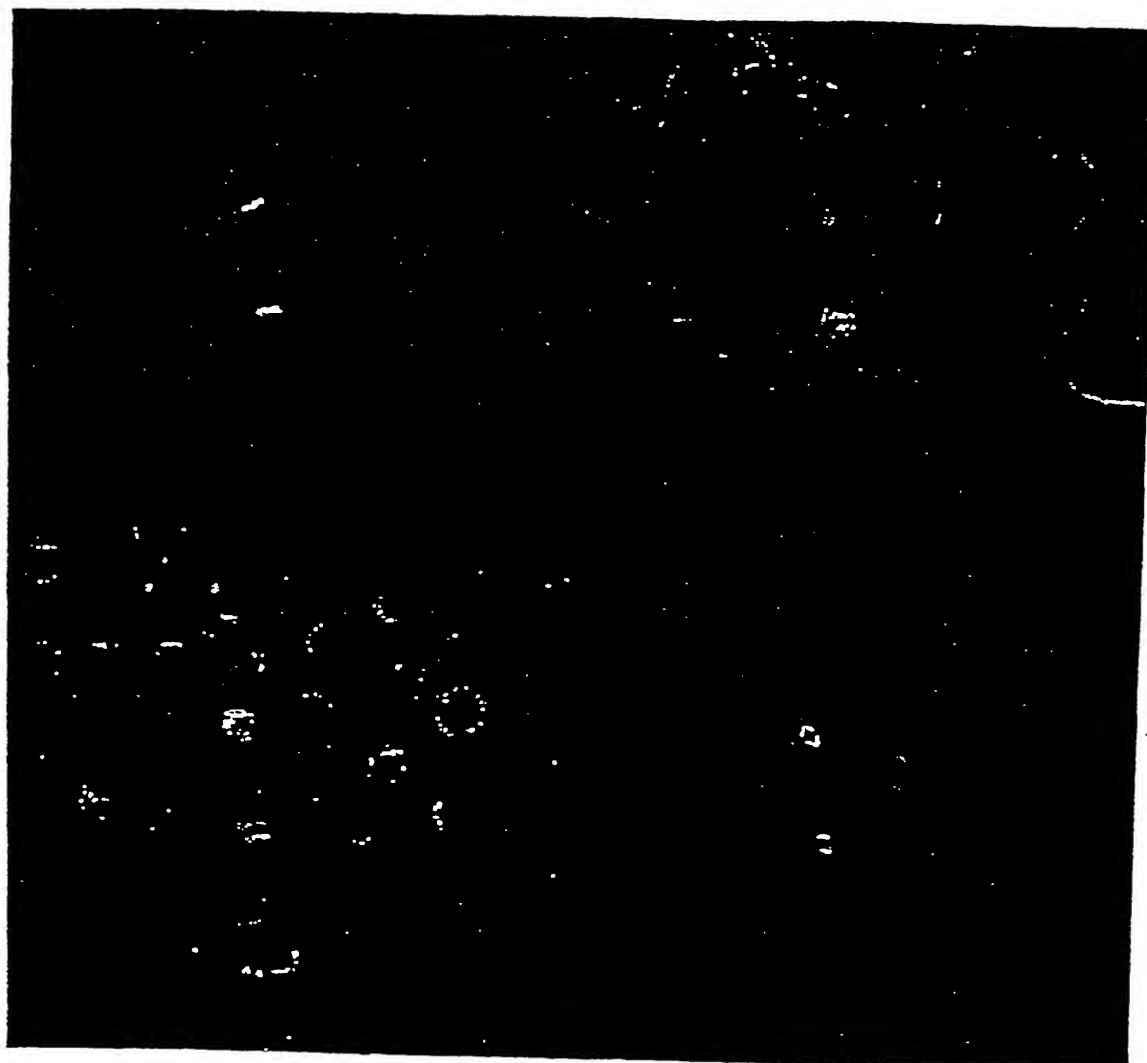


Fig 15

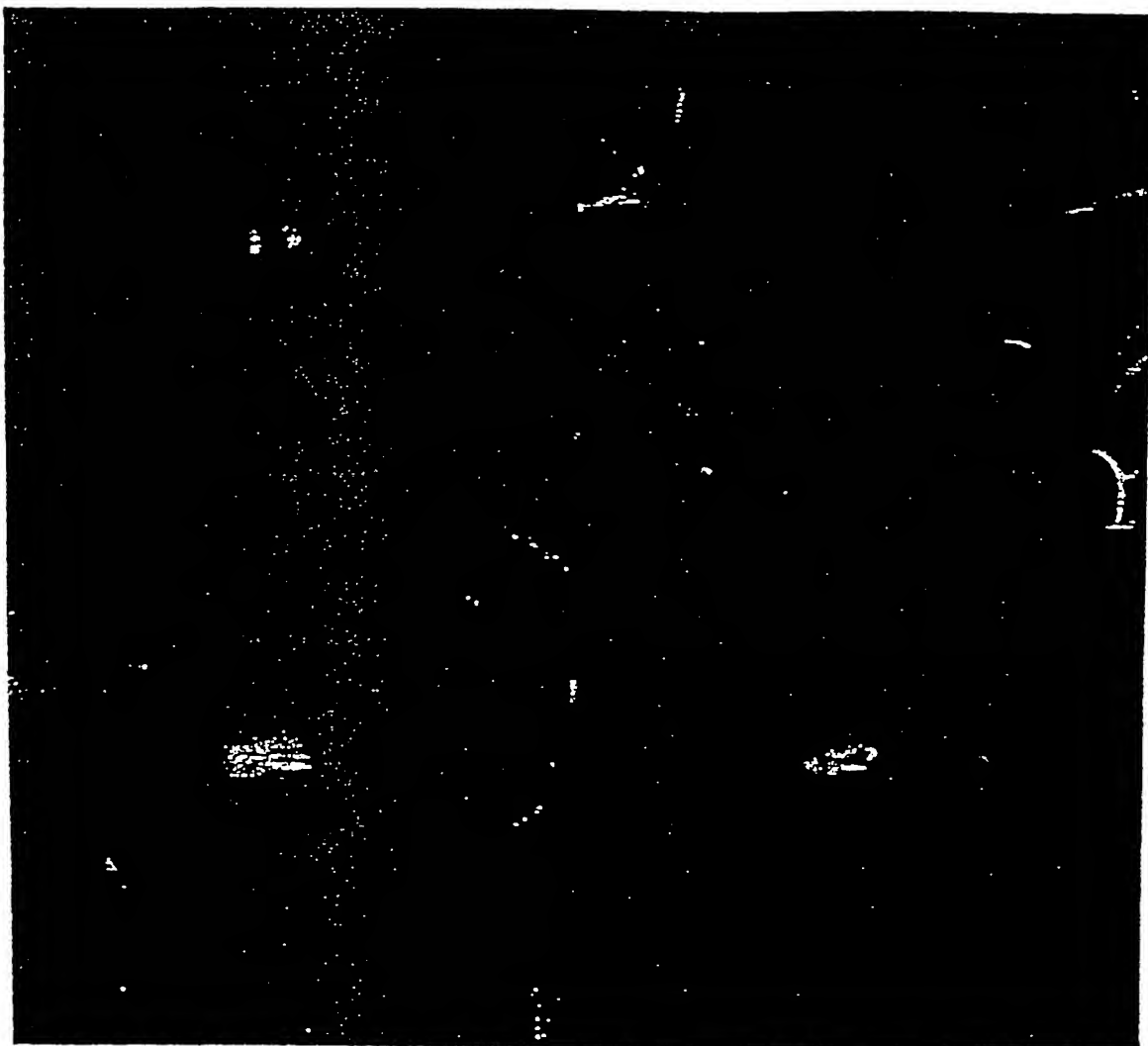


Fig 16

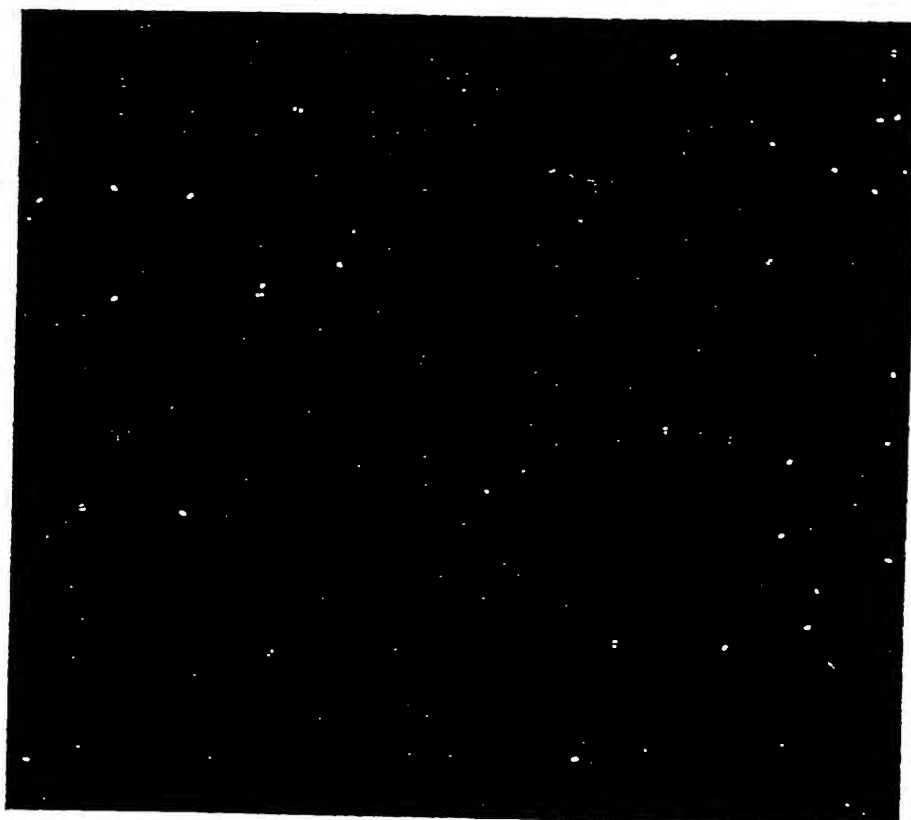


Fig 17

Conversion of morphometric parameters into nucleic acid code and clustering of the resulting sequences using Neighbor Joining method.

Compound:	Measurements																			
	Count	Area	Perimeter	Length	Breadth	Fiber length	Fiber breadth	Shape factor	Ell. form factor	Inner radius	Outer radius	Mean radius	Equiv. radius	Equiv. sphere vol.	Equiv. prolate vol.	Equiv. oblate vol.	Equiv. sphere surface area	Average gray value	Total gray value	Optical density
Control	t	t	t	t	t	t	t	t	t	t	t	t	t	t	t	t	t	t	t	t
Taxol	a	t	t	t	t	t	t	t	a	t	t	t	t	t	t	t	t	t	t	t
CD	c	a	a	a	t	a	t	t	c	a	a	a	a	a	a	a	a	t	a	a
Nocodazole	c	t	t	t	t	t	t	t	t	t	t	t	t	t	t	t	t	t	t	t
Staurosporine	g	g	c	a	a	t	a	a	t	g	a	a	a	t	g	g	g	a	a	t
Vinblastine	c	t	t	t	t	t	t	t	t	t	t	t	t	t	t	t	t	g	t	t
Hydroxyurea	g	t	t	t	t	t	t	g	t	t	t	t	t	t	t	t	t	t	c	t

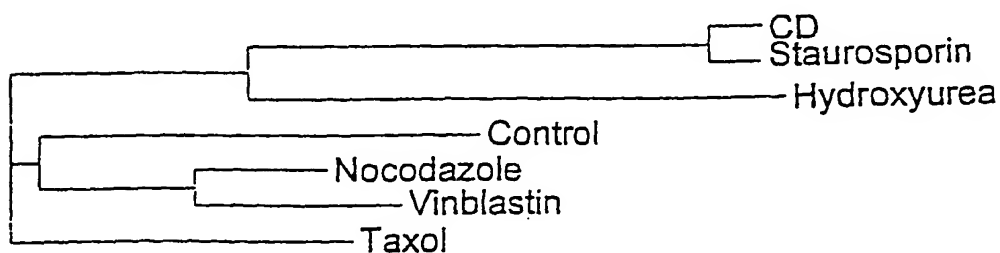


Fig 10

Conversion of morphometric parameters into amino acid codes and clustering of the resulting sequences using Neighbor Joining method.

	I	C	A	P	L	B	F	S	W	M	S	T	M	T	S	P	A	A	T	M	C	M	O	T	M	H	M	P	S	Y
Control	I	C	A	P	L	B	F	S	W	M	S	T	M	T	S	P	A	A	T	M	C	M	O	T	M	H	M	P	S	Y
Taxol	G	F	M	M	P	M	P	H	G	S	M	M	W	C	T	P	T	R	C	M	M	M	M	M	M	M	M	M	M	M
CD	F	G	G	G	M	G	M	K	A	G	G	G	G	G	G	G	G	H	G	G	G	G	G	G	G	G	G	G	G	G
Nocodazole	W	F	M	M	W	M	P	T	R	S	M	M	M	F	M	W	F	M	M	R	M	M	M	M	M	M	M	M	M	M
Staurosporine	N	V	A	G	G	M	G	G	Y	V	G	G	G	M	V	V	V	G	G	H	G	M	G	G	V	V	V	V	V	V
Vinblastine	F	W	W	M	W	W	C	W	D	S	M	W	W	M	M	M	W	M	V	E	M	M	M	M	F	P	P	P	P	P
Hydroxyurea	S	H	H	H	H	H	H	V	H	H	H	H	H	H	H	H	H	H	H	H	H	H	H	H	H	H	H	H	H	H

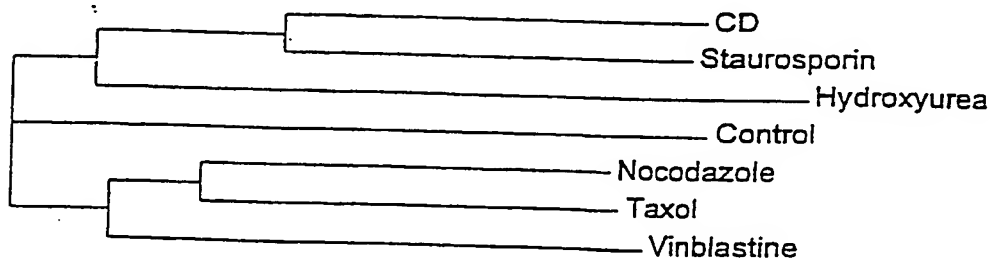


Fig 19

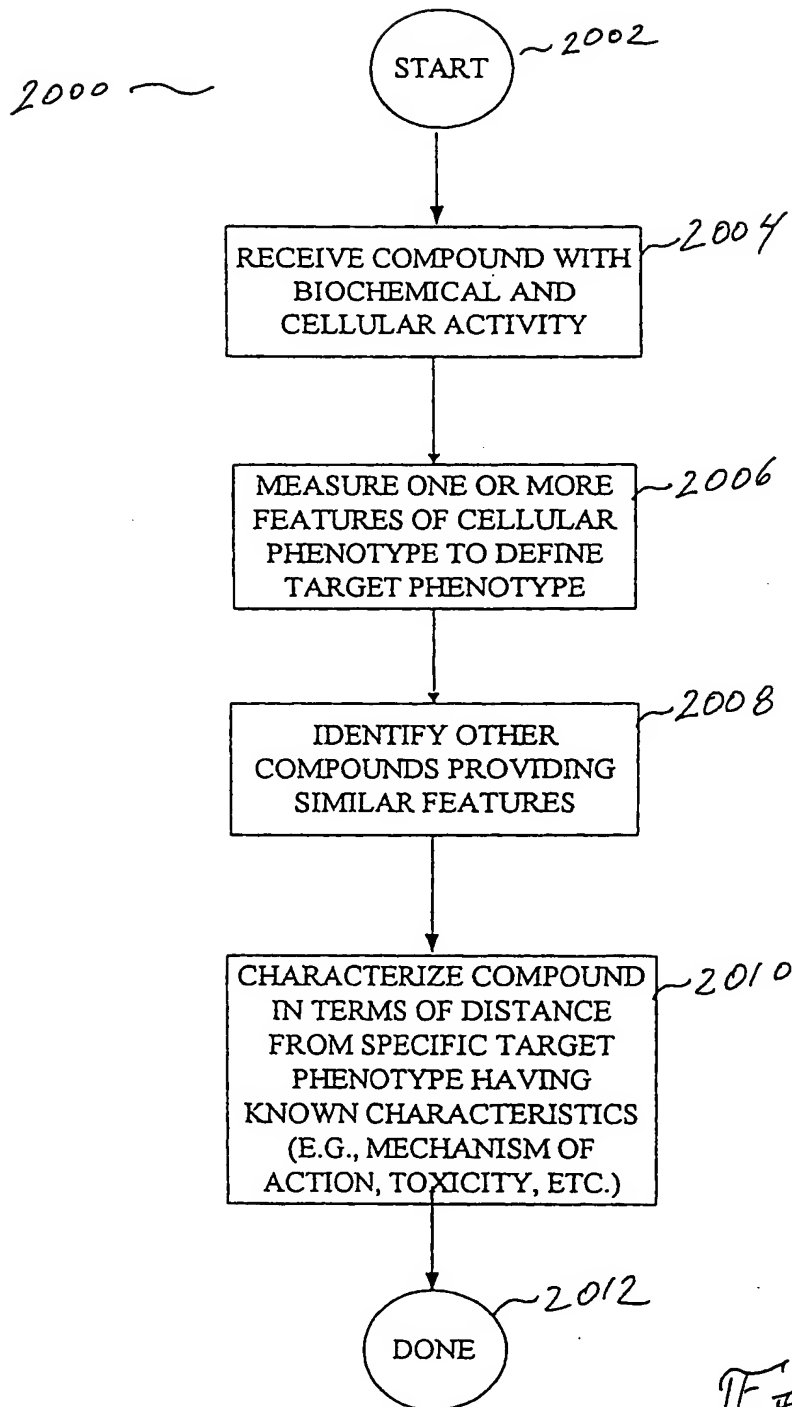


Fig 20

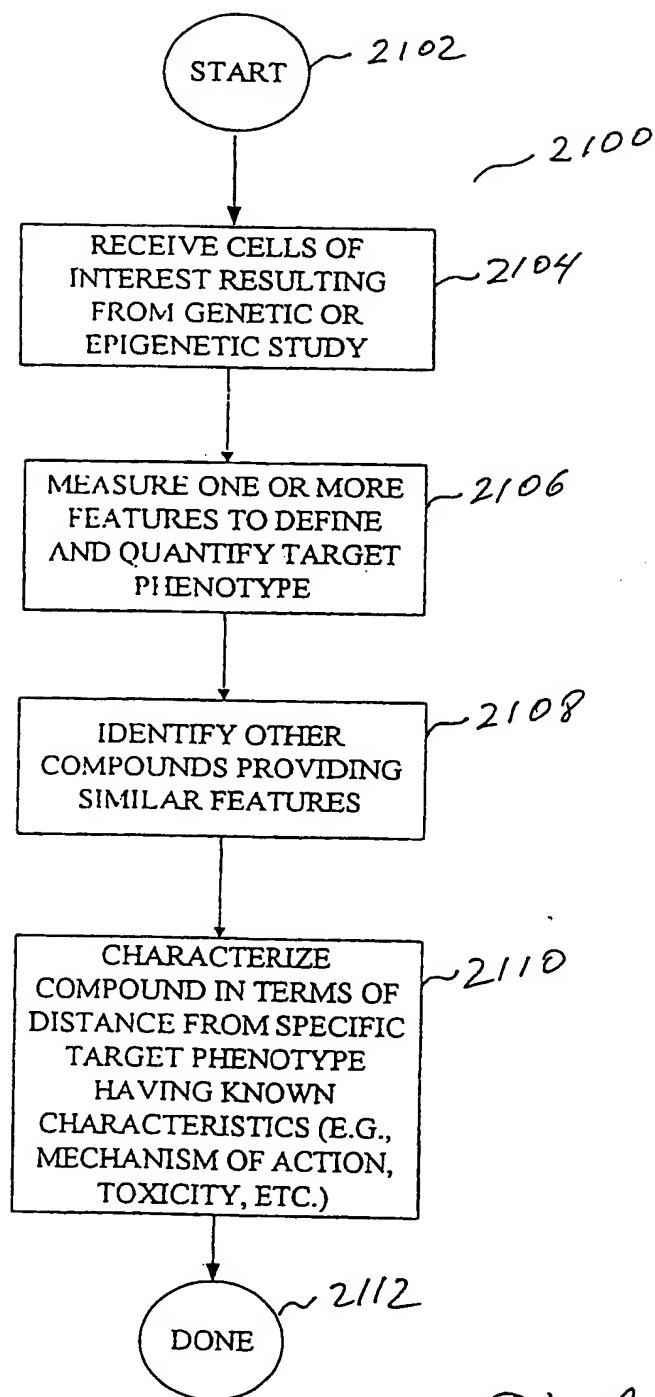


Fig 21

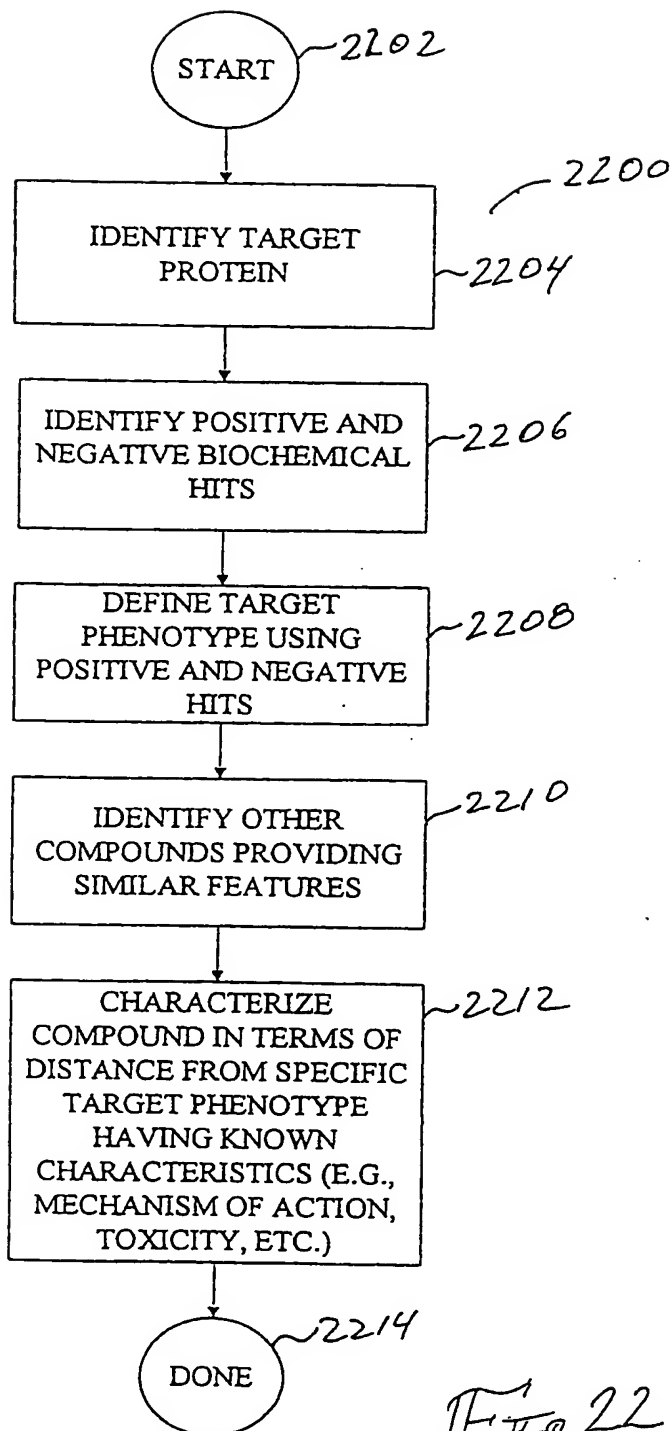


Fig 22

